

# Alzheimer's Disease & Treatment

## Chapter 4

### Fruit Fly (*Drosophila melanogaster*): A Viable Model for Screening Tropical Functional Foods for Neuroprotective Properties

Ganiyu Oboh<sup>1\*</sup>; Adedayo O Ademiluyi<sup>1</sup>; Ayokunle O Ademosun<sup>1</sup>; Opeyemi B Ogunsuyi<sup>1,2</sup>; Folasade L Oladun<sup>1</sup>

<sup>1</sup>Functional Foods and Nutraceutical Unit, Biochemistry Department, Federal University of Technology, P.M.B. 704, Akure, 340001, Nigeria.

<sup>2</sup>Department of Biomedical Technology, School of Health and Health Technology, Federal University of Technology, P.M.B. 704, Akure, 340001, Nigeria.

\*Correspondence to: Ganiyu Oboh, Functional Foods and Nutraceutical Unit, Biochemistry Department, Federal University of Technology, P.M.B. 704, Akure, 340001, Nigeria

Phone: +2347031388644; Email: [goboh2001@yahoo.com](mailto:goboh2001@yahoo.com) & [goboh@futa.edu.ng](mailto:goboh@futa.edu.ng)

#### Abstract

In the prevention of chronic disease such as neurodegenerative diseases, nutrition is critical; not just to meet nutritional requirements but more importantly to contribute to the total wellness of the consumer by either preventing and/or managing such disease conditions. This has further promoted the concept of functional foods and nutraceuticals. However, while several studies abound on the huge abundance and diversity of functional foods especially in tropical parts of the world, there is a still serious limitation to rapid and high throughput experimental screening for neuroprotective properties of several functional foods especially in developing nations. These limitations include modern, effective and accessible experimental models for rapid screening, cost of research and ethical issues with animal use among others. Fruit fly (*Drosophila melanogaster*) has emerged as a very useful model of neurodegenerative disease and could be more effective for therapeutic screening for neuroprotective properties of functional food and nutraceuticals especially from developing countries of tropical Africa. This

model organism has such advantages as short life span, high fecundity, low cost of maintenance, ease of handling and small genome size already sequenced and easy to manipulate. Therefore, this chapter review recent trends in functional food research especially of tropical African origin and how *D. melanogaster* can help optimize the effective screening of their neuroprotective properties.

## 1. Introduction

The saying “Let food be thy medicine and medicine be thy food” of Hippocrates about 2,500 year ago has attracted much scientific attention. This is clearly seen in the interest shown by scientists from various relevant fields in the role of specific food in enhancing health and well-being. While all foods could be said to have functionality mainly in terms of their nutritive values, however, the idea of functional food is not to be viewed as only necessary for living but is to also contribute to the total wellness of the consumer which involves prevention and reduction of disease risk factors, thereby enhancing the overall physiological function. In 1989, Stephen DeFelice, MD, (founder and chairman of the Foundation for Innovation in Medicine (FIM), Cranford) coined the word ‘nutraceutical’ from nutrition and pharmaceutical [1]. Nutraceutical can be described as a food (or part of a food) that provides medical or health benefits including the prevention and or treatment of a diseases. Nutraceuticals are often said to be products that are extracted or purified from animal, plant or marine sources which have shown physiological benefit or known to protect against chronic diseases.

Neurodegenerative diseases are pathologies with many ethology. Studies have shown that impairments in neurochemistry, oxidative stress and elevated metal ions deposits in the brain are few of the factors that contribute to the progression of neurodegenerative disease [2]. In order to set the key etiological factors as a focal point, it is very important to develop a multidimensional therapy that will prevent and manage these diseases. Most drugs designed all have short live span and side effects [3]. In order to overcome this limitation, dietary interventions as a complementary approach in management/prevention of neurodegenerative disease becomes imperative for holistic management.

Many studies have been published on therapeutic properties of functional foods especially of tropical African origins. In the last few decades, many interesting research publications have originated from Africa on therapeutic properties of several tropical functional foods. However, one major limitation to full evolution of functional food research in Africa has been adequate screening models. Many of published data on functional food from Africa has been from *in vitro* research with a good number on *in vivo* animal (usually mouse and rats) models. However, the current advocacy on ethical controls on laboratory rodent use is gradually challenging biomedical research generally in Africa. Therefore, it has become imperative to explore alternative models. *Drosophila melanogaster* also called fruit fly has emerged to the fore when it comes to therapeutic screening of functional foods, nutraceuticals, chemotherapeutic drugs

and lots more. Screening a large pool of therapeutics in search of few lead compounds is cost effective using *Drosophila*.

## **2. Functional Foods and Nutraceuticals: An Overview**

The term functional foods originates from Japan in the mid-1980s and it often referred to processed foods that possess active ingredients that aids specific function in the body in addition to its nutritional values. Around the world, Japan is foremost in creating specific regulatory approval process for functional foods. Functional food possesses a wide range of definition, one of which was published in European consensus publication as: “a food is said to be functional if it is satisfactorily demonstrated to affect beneficially one or more target functions in the body, beyond its adequate nutritional effects, in a way which is relevant to either the state of well-being and health or the reduction of the risk of disease”. Another definition is given by the Institute of Medicine of the National Academy of Sciences “as any food or food ingredient that may provide health benefit beyond the traditional nutrients it contains” [4]. Another simpler definition states that functional food are foods that are similar in appearance to conventional food and are often regarded as normal diet, but have been modified to provide physiological benefits apart from the basic nutrients they provide [5].

Functional food and conventional foods are often similar in appearance or even closely related. But they differ slightly in that functional foods has physiological benefits and are capable of reducing of the risk of chronic diseases beyond provision of nutritional values [6]. Functional food components are potentially beneficial components present in naturally occurring foods or functional ingredients added. These components include carotenoids, dietary fibres, fatty acids, flavonoids, phenolics acids, plant sterol, prebiotics and probiotics, soy proteins, vitamins and minerals, isothiocyanates. The concept of functional food and nutraceuticals are often used interchangeably but they can be distinguished. Functional food is a broad term used to describe food or part of food with specific function [7], whereas nutraceuticals deals with the expected result of the products which could be prevention or treatment of diseases. While functional food can also be food products that are required to be taken as part of usual diet so as to elicit beneficial effect (therapeutic effect) that goes beyond the known traditional nutritional benefit [8], nutraceuticals on the other hand could be described as purified products from plant or animal functional foods.

## **3. Functional Foods are Beyond Nutrient Sources**

A lot of scientific findings have proven that functional foods possess a wide range of therapeutic potential in the prevention and management of several diseases afflicting humans. Since functional food is mostly of plant origin the bioactive components present therein are majorly phytochemicals. These includes phenolic acids, flavonoids, alkaloids, ascorbic acids and vitamin E. These components have been reported upon to possess antioxidant property

which is one of the proposed mechanism through which they bring forth their therapeutic effects [9]. Antioxidants are essential molecules which are needed to counteract the deleterious effect caused by free radicals in biological tissues. Although, every biological organism has endogenous antioxidant systems to effectively manage the oxidative damages of free radicals, an overwhelming amount of free radicals could cause a tilt in this check and balance system which could lead to extensive tissue damage; this phenomenon is called oxidative stress. Oxidative stress often necessitates sourcing antioxidants from exogenous sources such as found in functional food. Furthermore, oxidative stress has been linked to the pathogenesis and progression of many human diseases such as diabetes, cardiovascular disease, inflammation, cancer and dementia among others. Interestingly, reports have shown potentials in the management of these diseases such as ginger and turmeric for the management of hypertension [10,11], green leafy vegetables for the management of dementia [12,13] and legume seeds for the management of type 2 diabetes [14,15]. The health benefits of fruits and vegetables in preventing chronic diseases including type 2 diabetes are attributed to their antioxidant constituents including polyphenols, carotenoids and ascorbic acid which could help prevent or ameliorate oxidative stress [16]. In addition to ameliorating oxidative stress, the bioactive component of fruits and vegetables can elicit their antidiabetic effect by stimulating insulin secretion and inhibiting carbohydrate absorption in the small intestine ultimately resulting in lower blood glucose level [17, 18]. In the past decades, fruits and vegetables have gained much interest in the management of cancer [19]. Although, most functional foods are of plant sources but some are also of animal origin that is quite interesting. Examples of such are (n-3) fatty acids from fish and conjugated linolenic acid present in milk and meat products [20]. In the management of cardiovascular disease, (n-3) fatty acid which are essential class polyunsaturated fatty acids have shown potential.

#### **4. Tropical Functional Foods for the Brain**

Functional foods that have neuroprotective properties are of higher focus due to the increased incidence of neurodegenerative diseases. A brief outline of research findings on some tropical functional foods with neuroprotective properties is given in table 1. Nevertheless, it is still clear that many outstanding *in vitro* findings do not often get to *in vivo* levels due to among others factors of cost and ethical issues with the use of mammalian models.

**Table1:** A brief outline of research findings on some tropical functional foods with neuroprotective properties

Class	Common name	Botanical name	Plant part	Major findings	Nature of study	Reference
Fruits and nuts	Citrus (orange, grape fruit and shaddock)	<i>Citrus spp.</i>	Peel	1. AChE, BChE and MAO inhibitory properties 2. Antioxidant properties	<i>In vitro</i>	[21]
	African star apple	<i>Chrysophyllum albidum</i>	Fruit			[22]
	Mangosteen	<i>Garcinia mangostana</i> Linn.	Fruit	1. Antioxidant properties 2. Protect against H <sub>2</sub> O <sub>2</sub> -induced oxidative stress in NG 108-15 neuroblastoma cells.	<i>In vitro</i>	[23]
Spices	Tumeric	<i>Curcuma longa</i>	Rhizome	1. Chronic dietary inclusion of turmeric rhizome protects against MPTP-induced PD in mice 2. Dietary inclusion of turmeric rhizome ameliorates alterations in activities of major neuronal enzymes in synaptosomes from the cerebral cortex of hypertensive rats	<i>In vivo</i>	[24, 25]
			Curcumin isolated from turmeric rhizome	1. Protect against ethanol-induced brain damage in rats 2. Protects blood-brain barrier integrity in cerebral ischemic rats 3. Synergize anti-amnesic and anticholinesterase properties of Donepezil in rats	<i>In vivo</i>	[26, 27]
	Pepper		Fruit	1. AChE and BChE inhibitory properties 2. Antioxidant properties	<i>In vitro</i>	[28, 29]
		<i>Capsicum spp.</i>		Ameliorate cyclophosphamide induced oxidative stress in rat brain	<i>In vivo</i>	[30]
			Capsaicin isolated from red pepper	Ameliorate biochemical markers in MPTP-induced PD in male C57BL/6J mice	<i>In vivo</i>	[31]

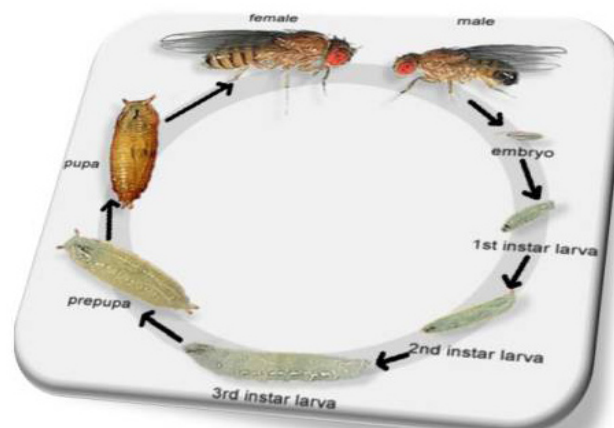
	Ginger		Rhizome	Enhance cognitive function and protect against neurochemical alterations in experimentally induced mammalian models of cognitive dysfunction		[32]
		<i>Zingiber officinale</i> Roscoe	Essential oil	Prevents oxotremorine-induced tremors and increased the latency of pilocarpine-induced seizures, as well as survival at 50 and 100 mg/kg in male swiss mice. However, higher dose of 100 mg/kg presents some cognitive impairments	<i>In vivo</i>	[33]
	Onion	<i>Allium cepa</i>	Bulb	Attenuation of ischemia-induced oedema and elevation in MDA level in mice brain	<i>In vivo</i>	[34]
				Protection against ischemic neuronal damage in Gerbil hippocampus		[35]
Vegetable	Tomato	<i>(Lycopersicon esculentum</i> Mill. var. <i>Esculentum</i> <i>Lycopersicon esculentum</i> Mill. var. <i>Cerasiforme)</i>	Fruit	1. AChE inhibitory property 2. Inhibit Fe <sup>2+</sup> and QA-induced lipid peroxidation in rat brain	<i>In vitro</i>	[36]
	African Jointfir	<i>Gnetum africanum</i>	Green leafy vegetable	AChE, BChE and MAO inhibitory properties	<i>In vitro</i>	[37]
	Fluted pumpkin	<i>Telfairia occidentalis</i> )	Green leafy vegetable	1. AChE inhibitory property 2. Inhibit Fe <sup>2+</sup> SNP and QA-induced lipid peroxidation in rat brain	<i>In vitro</i>	[12]
	Bitter leaf	<i>Vernonia amygdalina</i>	Green leafy vegetable	Antioxidant properties and Inhibit H <sub>2</sub> O <sub>2</sub> -induced lipid peroxidation in rat brain	<i>In vitro</i>	[38]
	Horseradish	<i>Moringa oleifera</i>	Green leafy vegetable	Ameliorate cognitive dysfunction in STZ-treated rats treated with acarbose	<i>In vivo</i>	[39]
	Black nightshade	<i>Solanum nigrum</i> L	Green leafy vegetable	Ameliorate cognitive dysfunction and neurochemical impairments in scopolamine-treated rats	<i>In vivo</i>	[13]
	African eggplant	<i>Solanum macrocarpon</i> L		AChE inhibitory and antioxidant properties	<i>In vitro</i>	[40]

AChE= Acetylcholinesterase. BChE= Butyrylcholinesterase, MAO= Monoamine oxidase, MDA= Malondialdehyde, MPRP= 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine, PD= Parkinson's disease, QA= Quinonic acid, SNP= Sodium nitroprusside, STZ= Streptozotocin

## 5. Fruit Fly (*Drosophila Melanogaster*) as a Therapeutic Screening Model for Functional Foods

### 5.1. Life cycle of *drosophila melanogaster* in brief

*Drosophila* has a very speedy life cycle (**Figure 1**). A single mating pair that is fertile can produce hundreds of offspring that are similar genetically in a period of 10- 12 days at room temperature. This is highly contrasting to the laboratory rodents that produce just a few offspring every 3- 4 months. The development of this fly occurs in stages (complete metamorphosis): the egg (embryo), larva, pupae and the full grown adult. The larva stage is actually in three phases; the first instar larva, second instar larva and the third instar larva. The third instar larva often wander in the culture media and it is at this phase the pupa develops. A great morphological changes often characterize the development of the third instar larva into pupae and eventually to the full grown adult. Under optimal condition life span of an adult fly is approximately 120 days. (For further reading, see Abolaji et al., [41] for more detailed review on *drosophila* development and husbandry).



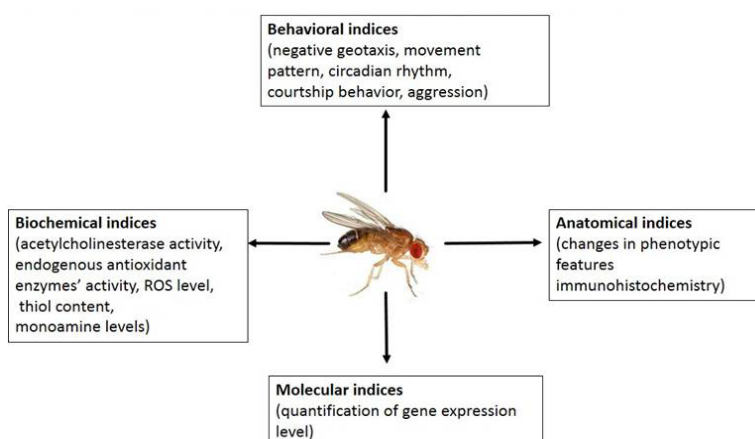
**Figure 1:** Life cycle of fruit fly (*Drosophila melanogaster*) (Abolaji et al., [42])

### 5.2. Potentials of *Drosophila melanogaster* as Model Organism

As stated earlier, functional food and nutraceuticals are of great interest to humans. Therefore, it becomes imperative to screen large pool of potential sources of functional foods and nutraceuticals for their perceived therapeutic functions; obviously, using humans as a screening model will be ambiguous, the extreme number of rats to be used will be unethical and *in vitro* studies will be insufficient. In search of a way out, the use of *Drosophila melanogaster* as a model animal comes to fore. Although, it might seem ironic that a ‘tiny’ fruit fly serving as a model for human therapeutic screening and physiologically speaking, there exist a wide range of differences between humans and the fruit fly. However, there is genetic homology between them which is the striking factor that makes research using *D. melanogaster* unique. Approximately 75% of diseases causing genes in human are conserved in *D. melanogaster* [43]. Another striking fact about *D. melanogaster* is the ease and cost effectiveness to manipulate and create a transgenic fly using high throughput genetic procedures [44]. With the availability

of these procedures, it is easier to generate models of human disease rapidly through various genetic engineering processes including mutation, genetic inactivation, or mis-expression of fly homologs of human disease genes and protein themselves.

*Drosophila* can also be used to monitor various pathological indices of neurodegeneration (figure 2). The wide varieties of these indices spanning biochemical, anatomical, molecular and behavioural aspects of neurodegeneration makes this organism quite useful as a research model. The flies present several anatomical features that can be monitored as indices of incidence and progression of neurodegeneration. Such features as simple as wing shape, eye colour and shape, fly size, larva size, and as complex as neuronal integrity, microtubule formation, synaptic formation and function could be used as markers of neurodegeneration as well as to monitor the potentials of any test therapeutic agent [45-48]. Such intrinsic anatomical alterations are often the phenotypic realities of several biochemical and molecular changes which ultimately reflect in their behavioural patterns such as their ability to climb (negative geotaxis), sleep-wake behaviour (circadian rhythm), copulation (aggressive behaviour) and their spatial orientation (movement pattern). Usually, fly models of neurodegeneration with marked anatomical and behavioural changes display concomitant biochemical and molecular modulations such as elevated level of reactive oxygen and nitrogen species, impaired cholinesterase activity, alterations in activities of antioxidant enzymes and levels of neurotransmitters, as well as modulation of gene expression levels of proteins of therapeutic importance; all these offers several useful points of evaluating therapeutic potentials of test compounds [46, 49-54]. Furthermore, the short and completely sequenced genome, coupled with the availability of relevant and accessible bioinformatics tools for *Drosophila* such as 'fly base' (<http://flybase.org>) makes investigations using *Drosophila* as model organism at the molecular level more achievable and attractive.



**Figure 2:** Chart showing the various pathological indices that *Drosophila melanogaster* can be used to monitor.

### 5.3. Current Possibilities

While the use of *D. melanogaster* for biomedical research in general and functional food research in particular in developing parts of the world such as Africa is still at its nascent



stage, it is interesting to note that a few interesting findings are already being published. One of such study accessed the toxicological implications of the popular condiment-monosodium glutamate and reported the toxicological implications of its consumption [55]. In this study, the author reported that feeding flies with monosodium glutamate up to 2.5 g/kg diet for five days significantly reduced their longevity, induced production of reactive oxygen and nitrogen species, hydrogen peroxide production and impaired activities of catalase and glutathione-S transferase antioxidant enzymes. Another study by Farombi et al., [56] recently used *D. melanogaster* as a model organism to show the ameliorative effect of kolaviron (the biflavonoid from bitter kola) on rotenone-induced toxicity. The biflavonoid was able to ameliorate the impaired locomotor performance, reduced life span, altered enzymes' activities and ROS/RONS production induced by rotenone. Another biflavonoid (hesperidin), a nutraceutical from citrus has been investigated for its therapeutic properties using *D. melanogaster* as model organism [57, 58,54]. Furthermore, curcumin which is adjudged the main polyphenol in turmeric was also shown to modulate acetylcholine gene expression level in *D. melanogaster* [53]; in this study, the authors reported that curcumin-supplemented diet improves survival ability increased antioxidant enzymes' activities but decreased AChE activities. The mRNA expression levels of AChE was similarly suppressed; the authors proposed this as one of the major mechanism behind the neuroprotective properties of this compound as previously reported [59-62]. Just recently, Abolaji et al., [63] studied the ameliorative effect of resveratrol on oxidative damage induced by 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine (MPTP) in *D. melanogaster*. MPTP has been extensive used to model Parkinson's disease in various animal models and as noted by the authors, this was the first time MPTP-induced toxicity will be studied in drosophila. These few are a testament to the possibilities of using *D. melanogaster* for functional food research especially in developing world. (For further reading, see 'Notes on Recent History of Neuroscience in Africa', by Russell, [64])

#### 5.4. Limitations

Although fly models can have high degrees of conservation and validity, providing opportunity for rapid screening and interpretation of results, however, modelling multifactorial human disease may be a bit complicated mainly due to the fact that such fly models usually express only certain aspects of the disease, making result interpretation more complex. Furthermore, while there seems to be a strong correlation of toxicity between the two organisms, nevertheless, due to metabolic differences, it is possible to observe that some drugs toxic to flies might not be in humans and vice versa [65]. In view of these limitations, it should be emphasized that *D. melanogaster* could be a useful model for rapid screening of a pool of functional foods and isolated nutraceuticals, as well as post screening validations to narrow down potential therapeutic candidates to a much smaller pool of lead substances/compounds, which could still be necessary to validate using conventional mammalian experimental

procedures. Nevertheless, with increasing attention being given to the use of *D. melanogaster* for biomedical research globally, which is attracting more sophisticated experimental protocols and re-validated research findings, these limitations might soon be overcome and a 'fly-to-bed' research approach, allowing direct clinical trials of validated fly-based research findings might not be too far away.

## 6. Acknowledgment

This chapter is written as part of a research funded by The World Academy of Science (TWAS) Grant No: 16-500 RG/CHE/AF/AC\_G – FR3240293300.

## 7. References

1. Biesalski, HK. Nutraceuticals: the link between nutrition and medicine. Nutraceuticals in Health and Disease Prevention New York: Marcel Dekker Inc. 2001: 1-26.
2. Bar-Am O, Amit T, Kupersmidt L, Aluf Y, Mechlovich D, Kabha H, et al. Neuroprotective and neurorestorative activities of a novel iron chelator-brain selective monoamine oxidase-A/moanoamine oxidase-B inhibitor in animal models of Parkinson's disease and aging, *Neurobiology of Aging*. 2015; 36:1529–1542.
3. Zarotsky V, Sramek JJ, Cutler NR. Galanthamine hydrobromide:an agent for Alzheimer ' s disease. *American Journal of Health-System Pharmacy*. 2003; 60: 446 –52.
4. Milner JA. Functional Foods and Health promotion. *The Journal of Nutrition*. 1999; 129: 1395S-1397S.
5. Bech-Larsen T, Grunert KG, Poulsen JB. The acceptance of functional foods in Denmark, Finland and the United states. A study of consumers' conjoint evaluations of qualities of functional food and perceptions of general health factors and cultural values. Working paper No. 73, University of Aarhus, Aarhus school of business, The MAPP Centre. 2001.
6. Arai, S. Studies on Functional foods in Japan- State of art. *Bioscience. Biotechnology and Biochemistry*. 1996; 60; 915.
7. Zeisel, SH. Regulation of " Nutaceuticals". *Science*. 1999; 285: 185-186.
8. Whitman M. Understanding the perceived need for complementary and alternative nutraceuticals: lifestyle issues. *Clinical Journal of Oncology Nursing*. 2001; 5: 190-194.
9. Oboh G, Ademosun AO, Ogunsuyi OB. Quercetin and its role in chronic diseases. *Drug discovery from mother nature*. 2016a: 377-387.
10. Akinyemi AJ, Thome GR, Morsch VM, Bottari NB, Baldissarelli, J, de Oliveira LS. et al. Effect of Ginger and Tumeric Rhizomes on Inflammatory Cytokines Levels and Enzyme Activities of Cholinergic and Purinergic Systems in Hypertensive Rats. *Planta medica*. 2016; 82: 612-620.
11. Wang Y, Yu H, Zhang X, Feng Q, Guo X, Li S, et al. Evaluation of daily ginger consumption for the prevention of chronic diseases in adults: a cross section study. *Nutrition*. 2016; 36: 79-84.
12. Oboh G, Akinyemi AJ, Ademiluyi AO, Bello, FO. Inhibitory effect of some tropical green leafy vegetables on key enzymes linked to Alzheimer's disease and some pro-oxidant induced lipid peroxidation in rats' brain. *Journal of Food Science Technology*. 2014; 51; 884–891.
13. Ogunsuyi OB, Ademiluyi AO, Oboh G, Oyeleye SI, Dada AF. Green leafy vegetables from two *Solanum* spp. (*Solanumnigrum* L and *Solanummacrocarpon* L) ameliorate scopolamine-induced cognitive and neurochemical

impairments in rats. *Food Science & Nutrition*.2018; 6: 860-870.

14. Oboh G, Ademiluyi AO, Akinyemi AJ, Henle T, Saliu JA, Schwarzenbolz, U. Inhibitory effect of polyphenol-rich extracts jute leaf (*Corchorus olitorius*) on key enzyme linked to type 2 diabetes ( $\alpha$ -amylase and  $\alpha$ -glucosidase) and hypertension (Angiotensin I converting ) in vitro. *Journal of Functional Foods*. 2012; 4: 450-458.

15. Ademiluyi AO, Oboh G, Boligon AA, Athayde ML. Effect of fermented soybean condiment supplemented diet on  $\alpha$ -amylase and  $\alpha$ -glucosidase activities in Streptozotocin-induced diabetic rats. *Journal of Functional Foods*. 2014; 9: 1-9.

16. Carter P, Gray LJ, Troughton J, Khunti K, Davies MJ. Fruit and vegetable intake and incidence of type 2 diabetes Mellitus: systemic review and meta-analysis. *BMJ*2010; 341: c4229.

17. Yavuz SM, Aygen B, Dogukan A, Tuzcu Z, Akdemir F, Komorowski JR, Atalay M, Sahin K. Chromium picolinate and chromium histidinate protects against renal dysfunction by modulation of NF- $\kappa$ B pathway in high-fat diet fed and streptozocin-induced diabetic rats. *Nutrition and Metabolism*. 2012: 9: 1-7.

18. Rafighi Z, Shiva A, Arab S, Yusuf R M. Association of dietary vitamin C and E intake and antioxidant enzymes in type 2 diabetes mellitus patients. *Global Journal of Health Sciences*. 2013; 3: 183-7.

19. Tramer F, Moze S, Ademosun AO, Passamonti S, Cvorovic J. Dietary anthocyanins: Impact on colorectal cancer and mechanism of action. *Colorectal Cancer- from Prevention to Patient Care*. 2011: 123-156.

20. De Deckere EAM, Verschuren PM. Functional Fat and spreads. In: functional foods. Gibson GR & Williams CM (eds). CRC Press, Cambridge, UK. 2015.

21. Ademosun AO, Oboh G. "Comparison of the Inhibition of Monoamine Oxidase and Butyrylcholinesterase Activities by Infusions from Green Tea and Some Citrus Peels". *International Journal of Alzheimer's Disease*. 2014.

22. Oboh G, Oyeleye SI, Akintemi OA, Olasehinde TA. Moringa oleifera supplemented diet modulates nootropic-related biomolecules in the brain of STZ-induced diabetic rats treated with acarbose. *Metabolic brain disease*. 2018b; 33: 457-466.

23. Weecharangsan W, Opanasopit P, Sukma M, Ngawhirunpat T, Sotanaphun U, Siripong P. Antioxidative and neuroprotective activities of extracts from the fruit hull of mangosteen (*Garcinia mangostana* Linn.). *Medical Principles and Practice*. 2006; 15: 281-287.

24. Mythri, R B, Veena J, Harish G, Rao BS, Bharath, MS. Chronic dietary supplementation with turmeric protects against 1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine-mediated neurotoxicity in vivo: implications for Parkinson's disease. *British journal of nutrition*. 2011; 106: 63-72.

25. Akinyemi A. J, Thome GR, Morsch VM, Stefanello N, da Costa P, Cardoso A, et al. Effect of dietary supplementation of ginger and turmeric rhizomes on ectonucleotidases, adenosine deaminase and acetylcholinesterase activities in synaptosomes from the cerebral cortex of hypertensive rats. *Journal of Applied Biomedicine*. 2016; 14: 59-70.

26. Rajakrishnan V, Viswanathan P, Rajasekharan KN, Menon VP. Neuroprotective role of curcumin from *Curcuma longa* on ethanol-induced brain damage. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*. 1999; 13: 571-574.

27. Jiang J, Wang W, Sun YJ, Hu M, Li F, Zhu DY. Neuroprotective effect of curcumin on focal cerebral ischemic rats by preventing blood-brain barrier damage. *European journal of pharmacology*. 2007; 561: 54-62.

28. Akinyemi AJ, Oboh G, Oyeleye SI, Ogunsuyi O. Anti-amnesic effect of curcumin in combination with donepezil, an anticholinesterase drug: involvement of cholinergic system. *Neurotoxicity research*. 2017; 31: 560-569.

29. Oboh G, Rocha, JBT. Hot Pepper (*Capsicum* spp.) protects brain from sodium nitroprusside-and quinolinic acid-induced oxidative stress in vitro. *Journal of medicinal food*. 2008; 11: 349-355.

30. Ogunruku OO, Oboh G, Ademosun AO. Water Extractable Phytochemicals from Peppers (*Capsicum* spp.) Inhibit Acetylcholinesterase and Butyrylcholinesterase Activities and Prooxidants Induced Lipid Peroxidation in Rat Brain In Vitro. *International Journal of Food Science*. 2014.
31. Oboh G, Ogunruku OO. Cyclophosphamide-induced oxidative stress in brain: protective effect of hot short pepper (*Capsicum frutescens* L. var. *abbreviatum*). *Experimental and Toxicologic Pathology*. 2010; 62: 227-233.
32. Fong GM. Mechanisms of neuroprotection by capsaicin, a red pepper extract. University of Sydney, Sydney Medical School, Discipline of Pathology. 2017.
33. Lim S, Moon M, Oh H, Kim HG, Kim SY, Oh MS. Ginger improves cognitive function via NGF-induced ERK/CREB activation in the hippocampus of the mouse. *The Journal of Nutritional Biochemistry*. 2014; 25: 1058-1065.
34. Felipe CFB, Fonseca KS, Bezerra JNS, de Franccedil MM, de Barros Viana GS. Alterations in behavior and memory induced by the essential oil of *Zingiberofficinale* Roscoe (ginger) in mice are cholinergic-dependent. *Journal of Medicinal Plants Research*. 2008; 2: 163-170.
35. Hyun SW, Jang M, Park SW, Kim EJ, Jung YS. Onion (*Allium cepa*) extract attenuates brain edema. *Nutrition*. 2013; 29: 244-249.
36. Hwang IK, Lee CH, Yoo KY, Choi JH, Park OK, Lim SS, et al. Neuroprotective effects of onion extract and quercetin against ischemic neuronal damage in the gerbil hippocampus. *Journal of medicinal food*. 2009; 12: 990-995.
37. Oboh G, Bakare OO, Ademosun AO, Akinyemi AJ, Olasehinde TA. Inhibition of Cholinesterases and Some Pro-Oxidant induced Oxidative Stress in Rats Brain by Two Tomato (*Lycopersicon esculentum*) Varieties. *International Journal of Biomedical Science*. 2015; 11: 48-53.
38. Nwanna EE, Oyeleye SI, Ogunsuyi OB, Oboh G, Boligon AA, Athayde ML. In vitro neuroprotective properties of some commonly consumed green leafy vegetables in Southern Nigeria. *The Journal of Nutrition and Food Science*. 2016;2: 19–24.
39. Alisi CS, Asiwe ES, Emejulu AA, Ene AC, Nwoguikpe RN. Neuroprotective and Free Radicals Scavenging Potentials of Some Common Leafy Vegetables Consumed in South-Eastern Nigeria. *Annual Research & Review in Biology*. 2014; 4: 3345.
40. Oboh G, Oyeleye SI, Akintemi OA, Olasehinde TA. *Moringa oleifera* supplemented diet modulates nootropic-related biomolecules in the brain of STZ-induced diabetic rats treated with acarbose. *Metabolic brain disease*. 2018a; 33: 457-466.
41. Olarewaju OA, Alashi AM, Taiwo KA, Oyedele D, Adebooye OC, Aluko, RE. Influence of nitrogen fertilizer micro-dosing on phenolic content, antioxidant, and anticholinesterase properties of aqueous extracts of three tropical leafy vegetables. *Journal of Food Biochemistry*. 2018:e12566.
42. Abolaji AO, Kamdem JP, Farombi EO, Rocha JBT. *Drosophila melanogaster* as a promising model organism in toxicological studies. *Arch Bas App Med*. 2013; 1: 33-38.
43. Lloyd TE, Taylor JP. Flightless Flies: *Drosophila* models of neuromuscular disease. *Annals of the New York Academy of Sciences*. 2010; 1184: e1-e20.
44. Dietzl G, Chen D, Schnorrer F, Su KC, Barinova Y, Fellner M, et al. A genome-wide transgenic RNAi library for conditional gene inactivation in *Drosophila*. *Nature*. 2007: 448; 151-156.
45. Wittmann CW, Wszolek MF, Shulman JM, Salvaterra PM, Lewis J, Hutton M, Feany, et al. Tauopathy in *Drosophila*: neurodegeneration without neurofibrillary tangles. *Science*. 2001; 293: 711-714.
46. Dias-Santagata, D, Fulga TA, Duttaroy A, Feany MB. Oxidative stress mediates tau-induced neurodegeneration in *Drosophila*. *The Journal of clinical investigation*. 2017; 117: 236-245.

47. Chakraborty R, Vepuri V, Mhatre SD, Paddock BE, Miller S, Michelson SJ, et al. Characterization of a *Drosophila* Alzheimer's disease model: pharmacological rescue of cognitive defects. *PloS one*. 2011; 6: e20799.
48. Mhatre SD, Satyasi V, Killen M, Paddock BE, Moir RD, Saunders AJ, Marendra DR. Altered synapses in a *Drosophila* model of Alzheimer's disease. *Disease models & mechanisms, dmm-*. 2014: 012104.
49. Hosamani R, Ramesh SR. Attenuation of rotenone-induced mitochondrial oxidative damage and neurotoxicity in *Drosophila melanogaster* supplemented with creatine. *Neurochemical research*. 2010; 35: 1402-1412.
50. Krishnan N, Rakshit K, Chow ES, Wentzell JS, Kretzschmar D, Giebultowicz, JM. Loss of circadian clock accelerates aging in neurodegeneration-prone mutants. *Neurobiology of disease*. 2012; 45: 1129-1135.
51. Adedara, I. A., Abolaji, A. O., Rocha, J. B., & Farombi, E. O. (2016). Diphenyl diselenide protects against mortality, locomotor deficits and oxidative stress in *Drosophila melanogaster* model of manganese-induced neurotoxicity. *Neurochemical research*, 41, 1430-1438.
52. Olakkaran, S., & Antony, A. (2017). *Convolvulus pluricaulis* (Shankhapushpi) ameliorates human microtubule-associated protein tau (hMAPt) induced neurotoxicity in Alzheimer's disease *Drosophila* model. *Journal of chemical neuroanatomy*
53. Akinyemi AJ, Oboh G, Ogunsuyi O, Abolaji AO, Udofia A. Curcumin-supplemented diets improve antioxidant enzymes and alter acetylcholinesterase genes expression level in *Drosophila melanogaster* model. *Metabolic brain disease*. 2018; 33; 369-375.
54. Poetini MR, Araujo SM, de Paula MT, Bortolotto VC, Meichtry LB, de Almeida, et al. Hesperidin attenuates iron-induced oxidative damage and dopamine depletion in *Drosophila melanogaster* model of Parkinson's disease. *Chemico-biological interactions*. 2018; 279: 177-186.
55. Abolaji AO, Olaiya CO, Oluwadahunsi OJ, Farombi EO. Dietary consumption of monosodium L-glutamate induces adaptive response and reduction in the life span of *Drosophila melanogaster*. *Cell biochemistry and function*. 2017; 35: 164-170.
56. Farombi EO, Abolaji A O, Farombi TH, Oropo AS, Owoje OA, Awunah MT. Garcinia kola seed biflavonoid fraction (Kolaviron), increases longevity and attenuates rotenone-induced toxicity in *Drosophila melanogaster*. *Pesticide Biochemistry and Physiology*. 2018; 145: 39-45.
57. Abolaji AO, Babalola OV, Adegoke AK, Farombi EO. Hesperidin, a citrus bioflavonoid, alleviates trichloroethylene-induced oxidative stress in *Drosophila melanogaster*. *Environmental toxicology and pharmacology*. 2017; 55; 202-207.
58. Manjula A, Subashini R, Punitha R, Subramanian P. Modulating effects of hesperidin on circadian pattern indices of rotenone induced redox homeostasis in clock mutant (*cry b*) of *Drosophila melanogaster*. *Biological Rhythm Research*. 2017; 48: 897-906.
59. Thiyagarajan M, Sharma SS. Neuroprotective effect of curcumin in middle cerebral artery occlusion induced focal cerebral ischemia in rats. *Life sciences*. 2004; 74; 969-985.
60. Zbarsky V, Datla KP, Parkar S, Rai DK, Aruoma OI, Dexter DT. Neuroprotective properties of the natural phenolic antioxidants curcumin and naringenin but not quercetin and fisetin in a 6-OHDA model of Parkinson's disease. *Free radical research*. 2005; 39: 1119-1125.
61. Cole, G. M., Teter, B., & Frautschy, S. A. (2007). Neuroprotective effects of curcumin. The molecular targets and therapeutic uses of curcumin in health and disease : 197-212.
62. Pan R, Qiu S, Lu D, Dong J. Curcumin improves learning and memory ability and its neuroprotective mechanism in mice. *Chinese Medical Journal*. 2008; 121: 832-839.
63. Abolaji AO, Adedara AO, Adie MA, Vicente-Crespo M, Farombi EO. Resveratrol prolongs lifespan and improves

1-methyl-4-phenyl-1, 2, 3, 6-tetrahydropyridine-induced oxidative damage and behavioural deficits in *Drosophila melanogaster*. *Biochemical and biophysical research communications*. 2018.

64. Russell VA. Notes on the Recent History of Neuroscience in Africa. *Frontiers in neuroanatomy*. 2017; 11: 96.

65. Rand MD. Drosophotoxicology: The growing potential for *Drosophila* in neurotoxicology. *Neurotoxicology and Teratology*. 2010; 32: 74-83.