

Alzheimer's Disease & Treatment

Chapter 2

Down Syndrome for A Better Understanding Alzheimer Disease

London J

Université Paris-Diderot, Sorbonne Paris Cité, BFA, UMR8251, CNRSF-72205, Paris, France

Email: london@univ-paris-diderot.fr

1. Introduction

A) Alzheimer disease (AD)

AD is the most common of dementia and affect millions individuals worldwide characterized mainly by loss of memory and cognitive decline but also by sensory and motor impairments. Most AD cases are sporadic (SAD 98% of cases) at age more than 60 years but some have a genetic inheritance (2% around) at age earlier than 60 years and some at even 40 years [1]. The autosomal dominant AD (ADAD) are related to mutations on presenilin 1 (*PSEN1* ; chromosome 14 ; 18-50% ADAD cases) and presenilin 2 (*PSEN2* ; chromosome 1 ; <5% ADAD cases) and on Amyloid Precursor Protein (*APP* ; chromosome 21; 10-15% ADAD cases).

At the biochemical level, SAD or ADAD is characterized by extracellular deposits of synaptotoxic β -amyloid ($A\beta$) peptides, mainly $A\beta_{40}$ and $A\beta_{42}$ peptides in fibrils structures forming amyloid senile plaques (SPs) and by intraneuronal neurofibrillary tangles of anormally phosphorylated Tau proteins (NTFs) [2,3].

These neuronal changes induce progressive neuronal and synaptic deficits leading to many deficit including a progressive cognitive decline. They are present may years before the apparition of the early clinical symptoms and this window of more than 10 years may offer not only methods for early diagnosis but also early treatments in the disease process before it is too severe.

The SPs begin according to Braak stages mainly in the neocortex for phase 1, spreading to the allocortex and then in phase 3 to the diencephalice nuclei, the striatum and the cholinergic

nuclei of the forebrain, and in phase 4 and 5 to other brain nuclei and cerebellum [4]. SPs accumulation precede by many years the first clinical symptoms of AD but some individuals never develop dementia despite the presence of SPs. A β peptides are present under many species derived from the clivage by α and β secretases of the APP protein derived from the clivage by β and γ secretases of the APP protein but the main species are A β 40 and A β 42; these peptides undergo abnormal configurations which by spreading lead to aggregates present in SPs mainly due the hydrophobic 25-35 sequence of the A β [5].

NFTs are present in cell bodies and apical dendrites and in abnormal neurites. The neurofibrillary lesions are mainly due to aggregates of the tau protein which is abnormally phosphorylated in specific sites mainly on threonine 212 (7-8 mol of phosphate in NTFs instead of 2-3 mol in control tau protein [6]. Hyperphosphorylated Tau protein tends to dissociate from microtubules and thus induces axonal impairment. NTFs develop in the brain through six Braak stages starting as silent in the transentorhinal stage then to the the limbic stage for the incipient AD and at the neocortical stage for full AD [7]. Moreover tau hyperphosphorylation can be induced by Abeta soluble dimers [8].

Besides SPs and NTFs, neurotrophins such as BDNF (Brain-derived neurotrophic factor) and monoamines such as serotonin (5-HT: 5-hydroxytryptamine) are involved in AD since they regulate cooperatively some aspects of neurogenesis, neural plasticity and survival during aging in order to allow the brain to respond to environmental demands which may be very detrimental. Many monoamines are modified in individuals with AD and in mice modelling AD [9]. In AD, BDNF is decreased in the hippocampus and the A β 42 peptide might compromise BDNF both production and signaling, leading to neuronal degeneration [10]. Reduced 5-HT levels is one of the markers of accelerating cognitive decline in AD [11]. Other monoamines DA, NA are also reduced in certain brain areas (frontal and temporal cortex, hippocampus) from individuals with AD but also in various brain areas from the APP/PS1 mouse model that develop cerebral amyloidosis [12]. Many patients with mild cognitive impairment (MCI) display some of the early clinical symptoms of AD. Among them some will be at risk for developing AD and some will not.

B) Down syndrome (DS)

DS is so far considered as the major genetic cause of intellectual disability. Down syndrome (DS) is a genetic disorder that results from the triplication of entire or part of chromosome 21 (Chr21) and occurs in 1 every 1000/1500 live births without family inheritance. Individuals with DS are not characterized by one symptom but present several pathological phenotypes which the accumulation of them induces the so-called DS phenotype which is highly variable among individuals and during aging [13]. Life expectancy in the DS population is shorter compared to non-DS individuals, but improvements in medical care and drug treatments have

significantly contributed to ameliorate the life quality of this population which now approaches 65-70 years and thus may be concerned with AD as the general population.

Although the increased dosage of Chr21 genes might be one of the causes of the phenotypical alterations of DS, the presence of trisomic genes also affects the expression of disomic genes, which, in turn, may gain aberrant expression and contribute to some clinical manifestations. Brain development defects induces intellectual disabilities which affect language, learning, memory but also motor, sensory and sleep deficits. Moreover it is reported that individuals with DS exhibit accelerating aging, behavioural abnormalities and early neurodegeneration which may be considered as major problems for this population.

Several early studies support the link between the DS phenotype and an increased risk of AD development [14 ; 15 ; 16]. The incidence of dementia among DS patients is 10% in the age range 35–50, 55% in the age range 50–60, and becomes 75% above the age of 60 years, but AD neuroanatomopathology is present in virtually all adults with DS older than 40 years. Despite these ubiquitous stigmates, the prevalence of dementia is variable ranging from 30-75% at age 65 and most of the cases of dementia in individuals with DS occur around 50-55 years. Nevertheless, there is a rather large subset of aged persons with DS who do not develop clinical signs of dementia at any age. Moreover it is reported that in peculiar cases, one woman of 78 years and one man of 65 years, the absence of the APP gene in triplicate leaves the individuals without the biochemical hallmarks of AD and consequently no dementia although they present the rest of the DS phenotype [17-18]. This rare patients confirm the crucial role of APP in the neuropathological and clinical findings of AD in individuals with DS. However, although plaques have been detected in young DS autopsy samples, and even in some fetal samples, it is only in late middle age that people with DS develop AD pathology [13;19]. The senile plaques are mainly due to the abnormal metabolism of the APP protein the gene of which is on chromosome 21 and the NTF are due to the abnormal phosphorylation of the Tau protein which might be partially due to some chromosome 21 phosphatases genes and also to some sAPP α dimers [8].

In the first study to comprehensively characterize DS samples with and without AD diagnosis in port-mortem brain samples and in the CSF, evident serotonergic and noradrenergic deficits were found in DSAD versus early AD individuals and to a lesser extent in DS versus healthy controls [20].

Although the clinical features and especially the time course might be different in AD in the general population and in individuals with DS, the biochemistry and pathology of AD in people with DS and in the typical population are essentially identical, the current « amyloid cascade hypothesis » is believed to apply to both populations. Thus, the population with DS represent the best human model to approach AD features several decades before the irreversible

dementia occurs.

2. Common Features between Individuals with DS and AD

The age of onset of dementia in individuals with DS is much earlier (35% at the age of 60 years) than the in SAD (70-80) but is similar (under 60) to the autosomal dominant AD (ADAD) [1].

Thus AD in DS individuals constitute the larger human group with a rather homogeneous genetic background to study the molecular pathways leading in some cases to dementia (DSAD) and thus allowing a window to test early markers and better understanding the course of this AD devastating disease.

Although, the definitive diagnosis for AD in the general population is often given post mortem by brain examination, many scales of evaluation of the cognitive deterioration are now used. Unlike familial AD, dementia in individuals with DS is still not well described although it accounts for the majority of genetic AD cases. The scales used for AD evaluations are not good enough for a population like DS that has as at baseline some cognitive impairments especially in comprehension, memory and language. Nevertheless, the CAMDEX-DS has been validated as a reliable tool for assessing clinical dementia in people with DS [21], the Rapid Assessment of cognitive function in Down syndrome (RADD) [22] and the novel Behavioral and Psychological Symptoms of Dementia in Down Syndrome (BPSD-DS) are reported in [23], which in combination with an amyloid PET scan should increase the confidence of dementia of Alzheimer's type in DS. A good presentation of the similarities and differences between clinical presentations in DS, AD (or SAD) and ADAD also named FAD is given in the updated review par Zis and Strydom in which they report also the comparison of DS and AD for biomarkers [24].

2.1. A β peptides

Briefly, in the plasma A β 40 and A β 42 levels are higher in individuals with DS; in DSAD, A β 42 tends to increase while A β 40 decrease thus the ratio A β 42/A β 40 has not the same time course to evaluate dementia as in AD. In CSF, A β levels have been shown to increase in childhood and then decrease when SPs start to deposit similarly to what is observed in AD [19].

Positron Emission Tomography imaging (PETscan) with ligands such as [11C]-Pittsburgh compound-B (PIB) enables « *in vivo* » quantification and localization of fibrillar A β deposits and also tangles [25].

Higher PIB binding levels in cortical and subcortical regions of the DS brain were observed in participants with higher age, lower cognitive performance on neuropsychological assessment,

and in those with a diagnosis of dementia [26,27,28]. In their work by PET and volumetric RMN [29]: they conclude that results from populations with amyloid overproduction (DS and ADAD) compared to the general population may be generalizable because all populations accumulate the same amyloid aggregates and experience the same overall temporal progression of AD in which amyloid accumulation precedes neurodegeneration and dementia.

Increased global amyloid- β was related to decline in verbal episodic memory, visual episodic memory, executive functioning, and fine motor processing speed. Participants who were consistently PiB+ demonstrated worsening of episodic memory [30]. Other modalities are now developed such as diffusion tensor or retinal imaging to improve early diagnostic [31].

2.2. Tau

Microtubule-associated-protein tau is hyperphosphorylated and abnormally phosphorylated in AD and aggregates in paired helical filaments (PHFs) in NTFs. The pretangle state, the intraneuronal tangles and the extraneurocellular NTFs can be differentiated by specific to particular Tau epitopes [32]. In general, NTFs follow a similar distribution in DS as in AD, starting in entorhinal cortex and spreading to hippocampus and later neocortex, but at a higher density in DS compared to AD brain. In the plasma the tau levels are higher in AD groups than in the general population. In individuals with DS these levels are also higher even in non demented ones [19].

2.3. APOE alleles and AD risk factor

APOE is a hepatocerebral lipoprotein that regulates the transport and deposition of cholesterol. Evidence suggests that harboring one or both apolipoprotein $\epsilon 4$ alleles (APO $\epsilon 4$) may increase the risk for AD due to the apoE as an essential catalyst of the amyloid cascade and a subsequent loss of function [33,34].

Inheritance of one copy of the allele increases AD risk four-fold while inheritance of two copies increases risk ten-fold in the general population. The APO $\epsilon 4$ allele also increases dementia risk in DS, albeit to a lower extent than in AD. At the biochemical level this increased risk is probably due to the presence of an amino-terminal apoE fragment in both the frontal cortex and hippocampus of the DS-AD brain [35].

2.4. Vascular pathology

Vascular pathology is common in the susceptibility to AD in the general population. The contribution of vascular pathology to dementia may play a similar role in age of onset and/or the rate of progression of AD in DS (DSAD) [36]. Microbleeds (Bs) were more frequent in DS cases relative to controls but present to a similar extent as sporadic AD. This aligned

with cerebral amyloid angiopathy (CAA) scores, with more extensive CAA in DS relative to controls in both brain regions. CAA was also more frequent in DSAD cases relative to sporadic AD [37]. Moreover, as reported in the recent study that vascular or metabolic imaging might provide earlier information regarding AD pathogenesis [38]. In the hippocampus of older DSAD individuals, Myoinositol (MI) is higher, Nacetylaspartate (NAA) is lower and glutamate-glutamine complex (Glx) is unchanged when compared to non-demented people with DS.

2.5. Oxidative stress

Oxidative stress (OS) results from accumulation of oxidized and damaged molecules which are not removed by the antioxidant defense system (superoxide dismutases (SOD 1, 2 and 3), glutathione peroxidase (GPx), catalase etc) and thus can damage various cellular and extracellular components. The chronic SOD1 overexpression in all cells that characterizes the trisomy 21 subjects and the consequent over-generation of endogenous hydrogen peroxide apparently is not adequately compensated by the relatively modest upregulation of catalase and GPx. Therefore, this chronic imbalance between the levels of both important antioxidant enzymes (SOD/CAT+GPX) and their corresponding substrates inducing the generation of the most deleterious hydroxyl radical (HO \cdot), might be the basis for DS disturbances. Biomarkers of oxidative stress are significantly elevated in DS [39]. The anti-oxidant system is affected in DS and this defect is worsened during aging [40]. The SOD1 has been shown to be elevated in the neocortex and the hippocampus of individuals with DS or AD [41]. Indeed loss of antioxidant enzymes and an increased in protein modification have been reported in AD [42, 43].

2.6. Mitochondria

Altered metabolism of APP might be related to mitochondrial dysfunction [44] and mutations in mitochondrial DNA have been related to AD changes in the general population as well as in the DS one [45].

Several studies have shown that alterations of mitochondrial structure and function associated with an impairment of reactive oxygen species (ROS) homeostasis are critically linked to DS pathogenesis. Deficits in energy metabolism due to mitochondrial dysfunctions negatively affect neuronal function, survival and central nervous system (CNS) development which requires ATP and occur as an early event in intellectual disability-linked diseases and several forms of dementia like AD [46]. Overexpression of APP may promote mitochondrial dysfunction in DS independent of aberrant A β deposition.

A recent review gives a survey of the role of mitochondria and its impaired functions in relation to oxidative stress both in DS and AD [47].

2.7. mTOR Pathway

mTOR influences A β deposition and tau aggregation and thus is associated with the pathogenesis and progression of AD and similarly in DS [28]. Briefly mTOR activation affects the regulation of A β generation/clearance and tau-phosphorylation by inhibition of the autophagy and by interaction with several key signaling pathways, including PI3K/Akt, GSK-3 β , AMPK and insulin/IGF. Inhibiting mTOR activation for the treatment and prevention of AD and AD-like dementia in DS has been pointed but much work need to be done before going to a trial [28].

2.8. Endosomes

Intracellular A β is localized to endosomes which are responsible for the turn-over and the degradation of the proteins within cells. Early endosomes are a major site of amyloid precursor protein (APP) processing and a convergence point for molecules of pathologic relevance to AD. Neuronal endosome enlargement, reflecting altered endocytic function, is a disease-specific response that develops years before the earliest stage of AD and DS [48]. Endosomal pathology contributes significantly to A β overproduction and accumulation in sporadic AD and in AD associated with DS and may signify earlier disease-relevant disturbances of the signaling functions of endosomes. One of the main point regarding endosomes is that large endosomes might be an hallmark for early detection for AD in DS and also for AD in the general population. More recently enlarged endosomes were detected in blood mononuclear cells and lymphoblastoid cell lines (LCLs) from individuals with AD using immunofluorescence and confocal microscopy showing that it may be a biomarker [49]. The volumes of enlarged endosomes correlate to [C11] PiB cortical index but not to the amyloid-beta, tau and phosphorylated tau levels measured in the cerebrospinal fluid. Moreover the enlargement of endosomes in DS is at least partially due to synaptojanin present in three copies [50].

2.9. Mis-segregation in AD

In their analysis [51] of thousands of cells from 27 AD and 13 control individuals showed that fibroblasts from AD patients were more than twice as likely to exhibit trisomy 21 compared to fibroblasts from control individuals. The increased frequency of trisomy 21 cells in fibroblasts from AD patients was significant and independent of age. Furthermore, the chromosome mis-segregation was associated with all types of AD, including sporadic and familial AD carrying a mutation in either PS1 or PS2.

3. Differences Between DS and AD Regarding Pathological Aging

3.1. Clinical aspects

In the DS population, the prevalence of dementia increases rapidly after the age of 30

years although data are very different from the very few cohorts studied. The overall prevalence of dementia in adults with DS is estimated at 6.8 % with an increase with age from 8.9 % in individuals up to 49 years to 32.1 % in individuals from 55 to 59 years old but it has also been reported higher as 33% among individuals with DS aged 30 to 39 years, 55% among those aged 40 to 59 years, and 77% for individuals above the age of 60 years. It should be point out that individuals who nowadays reach old ages are those who did not have cardiac abnormalities which most often were lethal without cardiac surgery available since 30 years in developed countries [19 ; 29]. A recent study shows some predictions about survival of people with DS according to the occurrence of AD [52].

By comparison, the prevalence of dementia in the general population is estimated as 4% below 65 years, 15% between 65 and 74 years, 43% between 75 and 84 years, and 38% over the age of 85 years [29].

3.2. Amyloid and tau aspects

The early study on 42 cases of DS under 40 years demonstrate the A β deposition within the hippocampus and parahippocampal gyrus prior to NFT in neurons or dystrophic neurites within SPs [53]. Moreover in this study and others it is reported that A β deposition in childhood and teens might be more frequent than previously estimated in DS and also than in the general population. Both infants, children and teens with DS accumulate intracellular A β prior to the accumulation of extracellular A β deposits [54].

A β production from APP nonspecific proteolytic cleavage leads to heterogeneous A β (including A β 40/42/43) in a possible endosomal/lysosomal location [55]. There is an accumulation of intracellular A β within neurons in DS at a much earlier age than in the general population. Moreover A β measured biochemically increase during aging suggesting an acceleration phase to disease development [56].

Schupf and colleagues reported that increasing levels of plasma A β 40 and decreased A β 42 were in DS good predictors of conversion to dementia [57]. Indeed, adults with DS with decreasing plasma 42 over time were 5 times more likely to become demented within 4 years. Similarly, in a separate prospective study on 405 persons with DS, those adults in the highest levels of plasma A β 42 and A β 40 also had the highest risk of developing dementia over an average 4.7 year period of time [58].

PIB binding in DS, first appearing in striatum, began around age 40 and was strongly associated with dementia and cognitive decline. The absence of a substantial time lag between amyloid accumulation and cognitive decline contrasts in sporadic/familial AD [27]. The study by PET imaging [59] has shown that in DS brain, A β binds to (18F) florbetaben and that this binding increases with age. Increased global amyloid- β was related to decline in verbal

episodic memory, visual episodic memory, executive functioning, and fine motor processing speed. Participants who were consistently PiB+ demonstrated worsening of episodic memory [60].

Regarding β amyloid peptides, it is interesting to note that BACE-2, a chromosome 21 protein which activates BACE1, was observed only in neurons of adults with DS but not in young people and in individuals with AD.

Few studies have focused on tau proteins in the course of AD in DS but a recent one shed a new light on this old controversy about the first hallmarks in AD [61].

3.3. Other biochemical aspects

As in the AD brain, the majority of proteins have been demonstrated to be oxidized (OS), thus the notion that aberrant protein oxidation in DS may contribute to AD development is relevant. In DS brain, despite increased OS levels, no changes in HO-1 protein levels has been observed in young subjects, whereas increased levels characterize adult DS subjects undergoing AD-like neurodegeneration. Interestingly, increased of HO-1 in DS/AD subjects is not comparable with that observed in AD subjects. This phenomenon seems likely linked to the trisomy of chromosome 21 BACH1 gene, which encodes for the nuclear repressor of HO-1 gene [62]. Indeed HNE modified proteins have been shown in excess in DS brain [63].

Accumulating evidence also suggests that impaired iron homeostasis is an early event in AD progression but also in DSAD [64]. Iron dyshomeostasis leads to a loss of function in several enzymes requiring iron as a cofactor, the formation of toxic oxidative species, and the elevated production of beta-amyloid proteins. DS might represent a specific case of genetically encoded OS. Indeed, there are a number of trisomic genes, that directly or indirectly affects ROS levels, either by causing increased ROS production or decreasing the antioxidant response [65]. The molecular mechanisms linking iron dysregulation to neurodegeneration in DS are still poorly understood.

4. Chromosome 21 Genes Involved Directly or Indirectly in AD

The entire sequence of human Chromosome 21 is now known and there are 233 coding genes, 299 long non-coding genes (Ensembl release 78) and 29 microRNAs (miRBase release 21). After investigation with Swiss-Prot and analysis with Gene Ontology Annotation, the 207 proteins found encoded on Chr 21: i) take part in 87 different biological processes, and 11 proteins are involved in signal transduction; ii) have 81 different molecular functions among which DNA binding and transcription factor activity are the most prevalent with 15 proteins; iii) are localized in 26 different cellular components, nucleus and the plasma membrane with 19 and 15 proteins, respectively, are the most predominant cellular localizations [66].

Some chromosome 21 genes might be important to better understand directly AD. Moreover some of chromosome 21 genes can regulate other genes and signalling protein involved in AD. The most important chromosome 21 genes known now to be involved in AD are : APP, BACE2, BACH1, DYR1A, ETS2, RCAN1, S100 β , SOD1, SYNAPTOJANIN. These genes encode proteins which have been shown to be important for some of the DS phenotype including the AD [67].

- **APP**

The gene coding for the APP protein is on chromosome 21 and its promoter is regulated by Ets-2 an another gene of chromosome 21 which is overexpressed in DS explaining at least partially the fact that APP is more than 1.5 expressed in DS cells and tissues. APP has already be mentionned previously as the source of the amyloid peptides which due to their abnormal folding will lead to agregates and later on to fibrills [68]. But it should be also point out that the proteolytic cleavage by α secretase gives the sAPP α extracellular fragment which has been shown to be beneficial for some cognitive functions especially memory, for neurite outgrowth, prevents A β generation and tau phosphorylation, counteract the A β effects and finally disrupt APP dimers [69]. The sAPP α extracellular fragment is present at lower levels in individuals with AD than in controls. Its potential beneficial role from an ombilical cord source has been recently assayed in animal models for AD and shown to be mediated by complement C1q [70].

- **BACE-2**

BACE-2 cleaves APP, is increased in DS and thus contributes to increased A β production [71]. Moreover BACE-2 activates BACE1, one of the proteolytic enzyme of the amyloid cascade [72].

- **BACH1**

BACH1 is a basic leucine-zipper protein and the nuclear repressor of HO-1 gene. Increased Bach1 expression levels are involved, overall, in repressing the induction of HO-1 in DS cases, thus reducing HO-1 overexpression in stress conditions as observed in AD. Overexpression of BACH1 is also related to oxidized species [62].

- **CBS**

CBS levels in DS brains are approximately three times greater than those in the normal individuals and CBS is localized to astrocytes and those surrounding senile plaques in the brains of DS patients with AD [73]. CBS is the main enzyme producing H₂S. CBS activity is reduced in AD brains and the decrease in H₂S may be involved in some aspects of the cognitive decline in AD [74]. Moreover, several experiments have shown in different rat or murine

models of AD that H₂S could be beneficial [75].

The CBS overexpression induces monoamine pathways alterations in various brain areas of CBS transgenic mice according to sex and age [76]. Thus these alterations by CBS overexpression might be involved in the developmental abnormalities in cognition in DS children and that may lead to AD in DS adults.

- **DYRK1A**

The DYRK1A gene encodes the protein DYRK1A which is a serine/threonine kinase. Dyrk1A is overexpressed in SPs from AD individuals and is a key molecule bridging β -amyloid production and tau phosphorylation in AD [77]. It phosphorylates tau protein making it a better substrate for GSK3 β phosphorylation, and DYRK1A phosphorylates alternate splicing factors leading to an increase ratio of 3R:4R tau, which is associated with neurodegeneration ; consistent with this finding, there is an increase in the number of DYRK1A-positive and 3R-positive NFTs in middle-aged and older DS brain compared to sporadic AD [78]. Overexpression of DYRK1A it at least partially responsible for the excessive synaptic inhibition in people with DS which can be reverses both in individuals with DS and in animal models by inhibiting DYRK1A with catechol compounds [79; 80; 81]. Moreover DYRK1A overexpression modulates monoamines neurotransmitters in transgenic mice altering both the serotonergic and the dopaminergic pathways [82].

- **ETS2**

Overexpression of ETS2 in DS may play a role in the pathogenesis of the brain abnormalities in DS and possibly AD [83]. Degeneration of DS neurons was reduced by dominant-negative ETS2, suggesting that increased ETS2 expression promotes DS neuronal apoptosis. In the human brain, ETS2 expression was found in neurons and astrocytes. Strong ETS2 immunoreactivity was observed in DS/AD and sporadic AD brains associated with degenerative markers such as Bax, intracellular A β , and hyperphosphorylated tau [84].

- **RCAN1**

RCAN1 levels are increased in the brain of DS and AD patients but also in the human brain with normal aging [85]. RCAN1 has been implicated in several neuronal functions including hippocampal plasticity. Its overexpression is involved in AD [86]. It is also involved to control the tightly coordinated process of fission/fusion in mitochondria (parra) and thus it induces mitochondrial defects seen in aging and AD [87]. Moreover RCAN1 increases susceptibility to oxidative stress.

- **S100 β**

S100 β expression levels are increased in both DS and AD astrocytes in association with neuritic plaques [88]. In addition, chronic overexpression of S100 β contributes to increased neuronal and neuritic APP expression with consequent accelerated amyloid deposition, as well as abnormal growth of neurites in β -amyloid plaques, similar to observations in middle-aged DS patients [89].

- **SOD1**

The SOD1 is localized in the neocortex and hippocampus from individuals with DS or AD [41]. The SOD1 activity is increased and not compensated by catalase or GPX thus leading to the overproduction of ROS.

SOD1 overexpression induces aberrant neuronal and mitochondrial proteins in hippocampus of transgenic mice [90] and alters the 20S proteasome during aging [91, 92]. The anti-oxidant system is affected in DS and this defect is worsened during aging [40]. It is also altered in AD [42].

- **Synaptojanin**

The phosphoinositide phosphatase synaptojanin 1 (SYNJ1) is key regulator of synaptic function Synaptojanin (Synj) and a dual phosphatase which regulates the Hedgehog pathway (Hh). It has been shown that reduction of Synaptojanin 1 (SYNJ1), the main phosphoinositol (4,5)-biphosphate phosphatase (PI(4,5) P2-degrading enzyme) in the brain and synapses, accelerates A β clearance in AD mice model [93]. It is overexpressed in DS brain and in APOE4 carriers with early AD and highly overexpressed in individuals with DS/AD [94].

Synaptojanin is involved in the homeostasis of inositols compounds and is overexpressed in DS brain [95]. Its metabolism is altered in the brain of Ts65Dn mice, the most commonly used model of DS. This defect is rescued by restoring SYNJ1 to disomy in Ts65Dn mice and is recapitulated in transgenic mice overexpressing SYNJ1 from BAC constructs [96]. It has been shown to be involved in the aging hippocampus memory deficits in three models of AD [97].

5. Animal Models to Study AD in DS Models

The first models for Down syndrome were transgenic mice for some key genes such as SOD1, APP, S100 β and CBS and some knock-out mice for some of them were also useful to understand the role of some of these genes either solely or in a complex genotype. Human chromosome 21 has orthologous genes on MMU 16, 17 and 10 and most of the partial trisomic murine models have been reviewed in [81; 98].

The first partial trisomic viable model, Ts65Dn, has been developed by Muriel Davisson

in the early 90's [99] and is trisomic for about 120 orthologs of Hsa21 protein encoding genes through a segmental trisomy for Mmu16 but it contains also a segment of about 10 Mb of the Mmu17, which contains 60 protein encoding genes, none of which are homologous to Hsa21 genes

Another mouse model for DS, Ts1Cje, has shorter Mmu16 trisomy than the Ts65Dn mouse, contains 62 orthologs of Hsa21 genes, and excludes the gene segment containing APP and SOD1.

The Ms1Ts65 mice are trisomic for the region of difference between Ts1Cje and Ts65Dn, contains 56 orthologs of Hsa21 genes within genetic segment from Mrp139 to Sod1. Another relatively novel model for DS is the Ts2Cje mouse model Ts (Rb (12.1716)) 2Cje (Ts2).

A more accurate mouse model for DS model have been developed by the team of Yu that carries complete aneuploidy (spanning the entire Hsa21 syntenic region) the triple aneuploid mice contains the Mmu10 (Ts1Yey), the Mmu 17 (Ts2Yey) and the Mmu16 (Ts3Yey).

A notable DS model was created from the trans-species insertion of Hsa21. The Tc1 mouse model for DS carries most of human chromosome 21 in addition to the normal complement of mouse chromosomes, and is trisomic for approximately 212 Hsa21 protein-encoding genes but not the APP gene [100].

The relationships between these models and their properties in relation to Alzheimer disease are presented in [101,102].

DS mouse models have also been investigated for better understanding the role of different gene segments of Hsa21 on AD pathology and memory impairment associated with DSAD. In these models some of the DS characteristics have been investigated regarding cognitive deficits biological mechanisms involved in DSAD including extracellular amyloid β protein (A β) accumulation, intraneuronal neurofibrillary tangles (NFTs) deposition, BFCN cell loss, neuron loss in locus coeruleus (LC), hippocampal abnormalities, imbalance of neurotrophic factors, alterations in long-term potentiation (LTP), abnormal endosomal signaling, presence of neuroinflammation and oxidative stress and more recently neurotransmitters.

The results obtained in these different models have been reported in recent studies [101;103] and most of them have been used to pre-clinical approaches. More recently a new model for studying AD and especially preclinical stages have been developed using a rat injected by adeno-associated viruses (AAV) coding for human mutant APP751 containing the Swedish and London mutations and PS1 (the M146L mutation) cDNAs into the hippocampi adult rats [104].

Before describing the pre-clinical approaches, we will developped a small paragraph

on neurotransmitters as we think that the abnormalities in neurotransmitters contents have not been enough considered in preclinical approaches.

5.1. Neurotransmitters

The monamine system has been studied in a few studies either from post-mortem brain tissues from individuals with AD or DS or both DSAD and in some animal models. Briefly in human tissues, a reduction of noradrenergic and serotonergic pathways were measured pathways in DSAD versus early onset of AD (EOAD) and to a lesser extent in DS versus controls [20]. DS and DSAD present similar monoaminergic profiles which might be related to early amyloid deposition in DS.

A recent study was performed on APP/PS1 mice [12] and showed no age effect in control mice according to age 6, 12, 16, 24 months but a region specific changes for all monoamines in 18 months APP/PS1 mice compared to controls.

The monoamines pathways evaluated in young Ts65Dn mice versus controls showed that a) the noradrenergic system was mainly affected by aging and not aneuploidy, b) the dopaminergic system was barely affected in Ts65Dn, c) the serotonergic was reduced (5-HT and 5-HIAA levels) in the hippocampus of young Ts65Dn versus controls but not in aged ones [105]. These results are quite different from those obtained transgenic mice overexpressing the single DYRK1A showing major deficits in serotonin contents for the four brain areas and major deficits in dopamine and adrenaline contents especially in the hypothalamus [82]. These differences between the two studies might be due to compensation between different chromosome 21 genes and non chromosome 21 genes present in the Ts65Dn mice.

The same type of studies should be performed on transgenic mice for important T21 genes related to DS and neurotransmission as it was done for CBS transgenic mice [76] and also in other models of T21 especially in rat models which seem to be more accurate to study AD in DS [106].

Neurotransmitters-bases strategies are plausible for targeting cognitive decline in AD and DS [107,108].

6. Pre-Clinical Approaches for AD in DS Models

A) *Search for biomarkers*

In the same time as researchers are trying to find therapeutical approaches, much work is done to find biomarkers for the conversion of AD in individuals with DS. These biomarkers found either in DS or in AD non DS will benefit for both of the populations.

In DS, the higher levels of A β 40 or A β 42/A β 40 correlates with the onset of AD in

DS. The formation of tangles is correlated with cognitive decline and PET studies of glucose metabolism might provide evidence of brain atrophy and AD changes in DS. Moreover as reported in the review by [109], many other markers show either decreases or increases; proNGF, MMP-1, TNF α , IL6, IL-10, SAH, SAM/SAH (from the chromosome 21 CBS gene) exhibit higher levels while serum MHPG, CSF orexin A show lower levels.

The combined use of DYRK1, BDNF (brain-derived neurotrophic factor) and homocysteine measured in the blood of two unrelated AD patient cohorts and age-matched controls has showed to give a sensitivity of 0.952, a specificity of 0.889 and an accuracy of 0.933 in testing for AD [110]. The same approach is currently used for DS in the progression of DSAD.

The rat model using AAV (AAV-AD) identified in the plasma of the various aged animals 41 proteins at 8 months and 21 proteins at 30 months which are specifically dysregulated in the course of AD pathology and thus identifying several steps before acute clinical hallmarks [104].

B) Inhibition of some DSAD hallmarks

Oxidative stress is a link between AD and DS [111]. Although since many years, treatments have been focused on antioxidants without real therapeutical progress, a study shed new light at the antioxidant status in the blood of DS children, before and after 6 months of daily antioxidant supplementation with vitamins E and C [112]. Before the antioxidant therapy, DS patients presented decreased GST activity and GSH depletion; elevated SOD, CAT, GR, GGT and MPO activities; increased uric acid levels; while GPx and G6PD activities as well as vitamin E and TBARS levels were unaltered. After the antioxidant supplementation, SOD, CAT, GPx, GR, GGT and MPO activities were downregulated, while TBARS contents were strongly decreased in DS. The same type of study should be done as the result obtained in a survey of trials with vitamin E only in mild cognitive impairment and Alzheimer's individuals fail to improve cognitive function, global severity or activities of daily living [113]. Other antioxidant compounds should be more investigated : Coenzyme Q10, curcumin to induce BDNF, melatonin.

- Inflammatory processes in BFCN degeneration can be modulated by minocycline treatment which inhibits microglial activation, prevents progressive BFCN decline and markedly improves performance of the Ts65Dn mice on a working and reference memory task [114].

- mTOR signalling and its aberrant modulation in DS and AD age-related cognitive decline affects crucial neuronal pathways. It was recently reported that intranasal rapamycin reverse many AD hallmarks. Indeed in the Ts65Dn mice this treatment could reduce APP

levels, APP processing and APP metabolites production, as well as, tau hyperphosphorylation and a reduction of oxidative stress markers [115].

- Modulation of neurotransmission. The group of Saheli has focused since many years on the possibility that neurotransmitter-based strategies and more specifically the noradrenergic system could be a target therapy for DS and AD [107; 108]. The GABAergic system is also a good approach and the use of antagonists of GABA receptors including pentylentetrazol (PTZ) to reduce perturbations of the excitatory/inhibitory balance towards an excess of GABA might be fruitful despite negative results up to date [81; 116]. Neurodegeneration can also be at least prevented by estrogen treatment which partially rescued working memory (T-maze test) and prevented neurodegeneration in aged Ts65Dn animals (11 to 17-months old) [117, 118].

- Regarding the endosome abnormalities, partial reduction of β -secretase1 (BACE1) by deleting one *BACE1* allele blocked development of age-related endosome enlargement in the medial septal nucleus, cerebral cortex and hippocampus, and prevented loss of choline acetyltransferase (ChAT)-positive medial septal nucleus neurons. It was also reported the possibility that studies focused on dysregulation of signaling endosome may identify some therapeutic targets for preventing DSAD [10].

- Exosome has been shown as a therapeutic approach for neurodegenerative disease [120]. Exosome secretion is enhanced in the brains of DS patients, in a mouse model of the disease and in DS fibroblasts. Furthermore, increased levels of the tetraspanin CD63, a regulator of exosome biogenesis, were observed in DS brains. As CD63 knockdown diminished exosome release and worsened endosomal pathology in DS fibroblasts, it was shown in the Ts65Dn mice that the increased CD63 expression enhanced exosome release to mitigate endosomal abnormalities in DS [121]. Exosome might be a biomarker for AD in DS [122].

- APP metabolism modulation

It was shown that long-term exposure to environmental enrichment reduces A β oligomers and rescues spatial-memory abilities in 12-month-old trisomic mice [123].

The Ts65Dn mice, as a model for DS and also for DSAD has been used for immunisation against A β oligomers [124]. In the States AC Immune and UCSD represents the first major clinical trial by a pharmaceutical company for Alzheimer's in the Down syndrome population. The study is focused on developing a vaccine that targets the amino acids of the A β peptides specifically involved in their misfolding, thus preventing their aggregation, the formation and plaque accumulation, and promoting plaque removal.

- Inhibition of DYRK1A

EGCG (Epigallocatechin-3-gallate) inhibits DYRK1A activity in vitro in several murine models of DS and mice treatment with EGCG restores some of the DS-associated deficits present in these models [79]. EGCG is not only an inhibitor of the biological activity of DYRK1A but it also binds the Tau protein in its phosphorylated region, hindering the access of this region to some kinase and modifying the tridimensional structure of the protein. These three combined functions of EGCG regarding Tau protein prevent its aggregation which is a key protagonist of neuronal cell death [125]. EGCG shows protective effects against A β -induced neurotoxicity and regulates secretory processing of sAPP α via the PKC pathway. Administration of EGCG (2 mg/kg) to mice for 7 or 14 days significantly decreased membrane-bound holoprotein APP levels, with a concomitant increase in sAPP α levels in the hippocampus. The role of EGCG in relation to AD is reviewed by [126]. Combined treatment with environment enrichment and EGCG ameliorates learning deficits and hippocampal alterations in Ts65Dn mice [127]. New DYRK1A inhibitors are currently studied in many laboratories [128, 129].

C) *Induced pluripotent stem cell (IPs) and deciphering AD in DS*

The use of pluripotent stem cell technology has given dramatic increase in our possibility for a better knowledge of the mechanism and the development of drugs in neurodegenerative disease [130]. A new inhibitor of DYRK1A, named ALGERNON (altered generation of neurons) was found in a screen for of neural stem cells (NSCs) [131]. This compound was found to rescue proliferative deficits in Ts65Dn-derived neurospheres and human NSCs derived from individuals with DS. Moreover, administration of ALGERNON to pregnant Ts1CJe dams rescued aberrant cortical formation in DS mouse embryos and prevented the development of abnormal behaviors in DS offspring. These data suggest that the neurogenic phenotype of DS can be prevented by ALGERNON prenatal therapy. As part of the neurogenic phenotype is also connected to AD, this type of experiments can lead to further evaluate if these corrected mice will present also some rescue of the DSAD phenotype although the Ts1CJe mice do not have the APP gene in triplicate.

Another recent study show promising results regarding AD, by the use of pluripotent IPSc from DS individuals [132]. It could be shown that in vitro generated DS neural cells have abnormal A β metabolism and increased expression of AD-associated chromosome 21 genes (BACE2, RCAN1, ETS2). These results show that it is possible to study AD-type pathology through the study of IPSC from DS individuals.

7. Conclusions

The close relationships between chromosome 21 genes involved in the onset of AD in the general population and their presence in third copies in individuals with DS yield new opportunities to better understand the course of AD, find biomarkers and innovative therapies for AD in the general population (SAD) through the precise study of the course of AD in

this special genetic population (DSAD). It is crucial to identify biomarkers for AD in this population so as to be able to determine the efficacy of any new treatment early in the course of the underlying disease process and well before the AD-related pathology and cerebral atrophy have become established. Thus the numerous mice models present for DS and their combination with others such as those silencing specific chromosome 21 gene are and will be very useful to assay new drug therapies. But mice are not men and the main important persons are those with DS or AD or both. In that perspective, the Eunice Kennedy Shriver National Institute of Child Health and Human Development (NICHD) and the National Institute on Aging (NIA), both parts of the National Institutes of Health, are partnering on an initiative to identify biomarkers and track the progression of Alzheimer's Disease in adults with DS. Moreover AC Immune and UCSD are trying to develop for individuals with DS a vaccine specific for the abnormal folding of the abeta peptides.

8. References

1. Bekris LM, Yu CE, Bird TD, Tsuang DW. Genetics of Alzheimer disease. *J Geriatr Psychiatry Neurol.* 2010; Dec; 23(4):213-27.
2. Selkoe DJ Alzheimer's Disease *Cold Spring Harb Perspect Biol.* 2011; Jul; 3(7): a004457.
3. Rikki Hullingera and Luigi Pugliellia Molecular and cellular aspects of age-related cognitive decline and Alzheimer's disease *Behav Brain Res.* 2017; March 30; 322(Pt B): 191–205 ; doi:10.1016/j.bbr.2016.05.008.
4. Thal DR, Rub U, Orantes M, Braak H Phases of Abeta-deposition in the human brain and its relevance for the development of AD. *Neurology* 2002; 58:1791-800.
5. Götz J Eckert A, Matamales M, Ittner LM, Liu X Modes of Abeta toxicity in Alzheimer disease. *Cell Mol Life Sci.* 2011; 68 : 3359-3375.
6. Kopke E, Tung YC, Shaikh S, Alonso AC, Iqbal K, Grundke-Iqbal I Microtubule-associated protein tau: Abnormal phosphorylation of a non- paired helical filament pool in Alzheimer disease. *J Biol Chem* 1993; 268(32): 24374-24384.
7. Braak H, Braak E Staging of Alzheimer' disease related to neurofibrillary changes. *Neurobiol Aging* 1995; 16: 271-278 (discussion 278-284).
8. Jin M, Shepardson N, Yang T, Chen G, Walsh D, Selkoe DJ Soluble amyloid β -protein dimers isolated from Alzheimer cortex directly induce Tauhyperphosphorylation and neuritic degeneration *Proc Natl Acad Sci U S A.* 2011; Apr 5; 108(14): 5819–5824. Published online 2011; Mar 18.
9. Šimić G, Babić Leko M, Wray S, Harrington CR, Delalle I, Jovanov-Milošević N, Bažadona D, Buée L, de Silva R, Di Giovanni G, Wischik CM, Hof PR Monoaminergic neuropathology in Alzheimer disease *Prog Neurobiol* 2017; 151 :101-138.
10. Chen XQ Sawa M, Mobley WC *Free Radic Biol Med* Dysregulation of neurotrophin signaling in the pathogenesis of Alzheimer disease and of Alzheimer disease in Down syndrome 2018; 114:52-61.
11. Trillo L, Das D, Hsieh W, Medina B, Moghadam S, Lin B, Dang V, Sanchez MM, De Miguel Z, Ashford JW, Salehi A. Ascending monoaminergic systems alterations in Alzheimer's disease. *Translating basic science into clinical care Neurosc Biobehav Reviews* 2013; 37: 1363-1379.
12. Von Linstow CU, Severino M, Metaxas A, Waider J, Babcock AA, Lesch KP, Gramsbergen JB, Finsen B Effect of

- aging and Alzheimer's disease-like pathology on brain monoamines in mice. *Neurochem Intern.* 2017; 108 : 218-245.
13. Lott IT Neurological phenotypes for Down syndrome across the life span *Prog Brain Res.* 2012; 197:101-21.
 14. Wisniewski KE et al Precocious aging in patients with Down's syndrome. *Biol Psychiatry.* 1978;13 (5): 619-27.
 15. Wisniewski KE Wisniewski HM, Wen GY Occurrence of neuropathological changes and dementia of Alzheimer's disease in Down's syndrome *Ann Neurol* 1985; 3:278-82.
 16. Mann DMA, Esiri MM The pattern of acquisition of plaques and tangles in the brain of patients under 50 years of age with Down's syndrome. *J Neur Sci* 1989; 89:169-179.
 17. Prasher VP, Farrer MJ, Kessling AM, Fisher EM, West RJ, Barber PC, Butler AC. et al. Molecular mapping of Alzheimer-type dementia in Down's syndrome *Ann Neurol* 1998; 43:380-383.
 18. Doran E, Keator D, Head E, Phelan MJ, Kim R, Totoiu M, Barrio JR, Gary W. Small GW, Potkin SG, Lott IT Down syndrome, partial trisomy 21, and absence of Alzheimer's Disease : the role of APP *J Alzheimer Dis* 2017; 56 (2):459-470.
 19. Head E. Lott IT, Wilcock DM, Lemere CA Aging in Down syndrome and the Development of Alzheimer's disease *Neuropathology Curr Alzheimer Res* 2016; 13(1): 18–29.
 20. Dekker AD, Vermeiren Y, Carmona-Iragui M, Benejam B, Videla L, Gelpi E, Aerts T, D Van Dam D, Fernández S, LMartin JJ Monoaminergic impairment in Down syndrome with Alzheimer's disease compared to early-onset Alzheimer's disease. *AlzheimersDement (Amst)* 2017; 10: 99-111.
 21. Ball SL, Holland AJ, Huppert FA, Treppner P, Watson P, Hon J. et al The modified CAMDEX informant interview is a valid and reliable tool for use in the diagnosis of dementia in adults with Down's syndrome *J Intellect Disabil* 2004; Res 48: 611-620.
 22. Walsh DM, Doran E, Silverman W, Tournay A, Movsesyan N, Lott IT Rapid Assessment of Cognitive Function in Down syndrome Across Intellectual Level and Dementia Status *J Intellect Disabil Res* 2015; 59(11): 1071-1079.
 23. Dekker AD, Vermeiren Y, Beugelsdijk G, Schippers M, Hassefras L, Eleveld J, Grefelman S, Fopma R, Bomer-Veenboer M, Oosterling GDE, Scholten E, Tollenaere M, Van Goethem G, Zu Eulenburg C, Coppus AMW, De Deyn PP. The Behavioral and Psychological Symptoms of Dementia in Down Syndrome (BPSD-DS) Scale: Comprehensive Assessment of Psychopathology in Down Syndrome *J Alzheimer's Dis.* 2018; 63: 797-820.
 24. Zis P, Strydom A Clinical aspects and biomarkers of Alzheimer's disease in Down syndrome *Free Radical Biology and Medicine* 2018; 114, 3-9.
 25. Mathis CA, Wang Y, Klunk WE. Imaging beta-amyloid plaques and neurofibrillary tangles in the aging human brain. *Curr Pharm Des.* 2004; 10(13):1469-92. Review
 26. Nelson LD, Siddarth P, Kepe V, Scheibel KE, Huang SC, Barrio JR, Small GW. Positron emission tomography of brain beta-amyloid and tau levels in adults with Down syndrome *Arch Neurol.* 2011 Jun; 68(6): 768-74.
 27. Annus T Wilson LR, Hong YT et al The pattern of amyloid accumulation in the brains of adults with Down syndrome. *Alzheimers Dement.* 2016; May12 (5): 538-45.
 28. Di Domenico F, Tramutola A, Foppoli C, Head E, Perluigi M, Butterfield DA mTOR in Down syndrome: Role in A β and tau neuropathology and transition to Alzheimer disease-like dementia. *Free Radic Biol Med.* 2018 Jan; 114: 94-101.
 29. Lao PJ, Handen BL, Betthausen TJ, Mihaila I, Hartley SL, Cohen AD, Tudorascu DL, Bulova PD, Lopresti BJ, Tumuluru RV, Murali D, Mathis CA, Barnhart TE, Stone CK, Price JC, Devenny DA, Mailick MR, Klunk WE, Johnson SC, Christian BT. Longitudinal changes in amyloid positron emission tomography and volumetric magnetic resonance

imaging in the nondemented Down syndrome population. *Alzheimers Dement (Amst)*. 2017 May 23; 9:1-9.

30. Hartley SL, Handen BL, Devenny D, Mihaila I, Hardison R, Lao PJ, Klunk WE, Bulova P, Johnson SC, Christian BT Cognitive decline and brain amyloid- β accumulation across 3 years in adults with Down syndrome *Neurobiol Aging* 2017; Oct 58:68-76.

31. Neale N, Padillab C, Fonseca LM, Hollandb T, Zamanb S Neuroimaging and other modalities to assess Alzheimer's disease in Down syndrome *NeuroImage Clin* 2018; 17, 263–271.

32. Augustinack JC, Schneider A, Mandelkow E-M, Hyman BT Specific tau phosphorylation sites correlate with severity of neuronal cytopathology in Alzheimer's disease. *Acta Neuropathol*. 2002;103: 26-35.

33. Rubinsztein DC Hon J, Stevens F, Pyrah I, Tysoe C, Huppert FA, Easton DF, Holland AJ Apo E genotypes and risk of dementia in Down syndrome. *Am J Med Genet*. 1999; 88(4):344-347.

34. Castellano JM, Kim J, Stewart FR, Jiang H, DeMattos RB, Patterson BW, Fagan AM, Morris JC, Mawuenyega KG, Cruchaga C, Goate AM, Bales KR, Paul SM, Bateman RJ, Holtzman DM. Human apoE isoforms differentially regulate brain amyloid- β peptide clearance. *Sci Transl Med* 2011; Jun 29 3(89):89ra57.

35. Day RJ, McCarty KL, Ockerse KE, Head E, Rohn TT Proteolytic Cleavage of Apolipoprotein E in the Down Syndrome Brain *Aging Dis*. 2016; 7(3): 267-277.

36. Wilcock DM, Schmitt FA, Head E Cerebrovascular contributions to aging and Alzheimer's disease in Down syndrome. *Biochim Biophys Acta* 2016; 1862(5):909-14.

37. Helman AM, Siever M, McCarty KL, Lott IT, Doran E, Abner EL, Schmitt FA, Head E Microbleeds and Cerebral Amyloid Angiopathy in the brains of people with Down syndrome with Alzheimer's disease. *J Alzheimers Dis* 2018; Nov 15.

38. Head E, Powell DK, Schmitt FA. Metabolic and vascular imaging biomarkers in Down syndrome provide unique insights into brain aging and Alzheimer disease pathogenesis. *Front Aging Neurosci* 2018; Jun 21; 10:191.

39. Jovanovic SV, Clements D, MacLeod K Biomarkers of oxidative stress are significantly elevated in Down syndrome. *Free Radical Biology & Medicine* 1998 ; 9: 1044-1048.

40. Perluigi M, Butterfield DA Oxidative stress and Down syndrome: a route toward Alzheimer-like dementia. *Curr Gerontol Geriatr Res*. 2012 ; 724904.

41. Furuta A Price DL, Pardo CA, Troncoso JC, Xu ZS, Taniguchi N, Martin LJ. Localization of the superoxidedismutases in Alzheimer's disease and Down's syndrome neocortex and hippocampus. *Am J Pathol*. 1995 116(2): 367-72.

42. Tramutola A, Lanzillotta C, Perluigi M, Butterfield DA Oxidative stress, protein modification and Alzheimer disease. *Brain Res. Bull*. 2017; 133:88-96.

43. Butterfield DA, Boyd-Kimball D Redox proteomics and amyloid β -peptide: insights into Alzheimer disease. *J Neurochem* 2018.

44. Busciglio J Pelsman A, Wong C, Pigino G, Yuan M, Mori H, Yankner BA Altered metabolism of the amyloid beta precursor protein is associated with mitochondrial dysfunction in Down's syndrome. *Neuron* 2002; 33: 677-688.

45. Coskun PE, Wyrembak J, Derbereva O, Melkonian G, Doran E, Lott IT, Head E, Cotman CW, Wallace DC Systemic mitochondrial dysfunction and the etiology of Alzheimer's disease and Down syndrome dementia. *J Alzheimers Dis* 2010; 20 Suppl 2:S293-310.

46. Moreira PI, Carvalho C, Zhu X, Smith MA, Perry G Mitochondrial dysfunction is a trigger of Alzheimer's disease pathophysiology. *Biochim Biophys Acta BBA-Mol. Basis Dis* 2010 ; 1802 2-10.

47. Valenti D, Braidy N, De Rasmio D, Signorile A, Rossi L, Atanasov AG, Volpicella M, Henrion-Caude A, Nabavi

- SM, Vacca RA Mitochondria as pharmacological targets in Down syndrome. *Free Radical Biology and Medicine* 2018 ; 114: 69-83.
48. Cataldo AM, Petanceska S, Terio NB, Peterhoff CM, Durham R, Mercken M, Mehta PD,
a. Buxbaum J, Haroutunian V, Nixon RA Abeta localization in abnormal endosomes: association with earliest Abeta elevations in AD and Down syndrome *Neurobiol Aging* 2004; 25(10):1263-72.
49. Corlier F, Rivals I, Lagarde J, Hamelin L, Corne H, Dauphinot L, Ando K, Cossec JC, Fontaine G, Dorothee G, Malaplate-Armand C, Olivier JL, Dubois B, Bottlaender M, Duyckaerts C, Sarazin M, Potier MC Clinical ImaBio3Team Modifications of the endosomal compartment in peripheral blood mononuclear cells and fibroblasts from Alzheimer's disease patients *Transl Psychiatry* 2015; 5(7): e595.
50. Cossec JC, Lavaur J, Berman DE, Rivals I, Hoischen A, Stora S, Ripoll C, Mircher C, Grattau Y, Olivomarin JC, de Chaumont F, Lecourtois M, Antonarakis SE, Veltman JA, Delabar JM, Duyckaerts C, Di Paolo G, Potier MC Trisomy for synaptojanin1 in Down syndrome is functionally linked to the enlargement of early endosomes. *Hum Mol Genet.* 2012; 21(14):3156-72.
51. Potter H, Granic A and Caneus J Role of Trisomy 21 Mosaicism in Sporadic and Familial Alzheimer's Disease *Curr Alzheimer Res.* 2016; 13(1): 7-17. Review.
52. Sinai A Mokrysz C, Bernal J, Predictors of Age of Diagnosis and Survival of Alzheimer's Disease in Down Syndrome *Journal of Alzheimer's Disease* 2018 ; 61 717-728.
53. Leverenz J, Raskind MA Early amyloid deposition in the medial temporal lobe of young Down syndrome patients: A regional quantitative analysis. *Exp Neurol* 1998; 150: 296-304
54. Gyure KA Durham R, Stewart WF, Smialek JE, Troncoso JC Intraneuronal Abeta-Amyloid precedes development of amyloid plaques in Down syndrome. *Arch Pathol Lab Med.* 2001; 125:489-492.
55. Glabe C Intracellular mechanisms of amyloid accumulation and pathogenesis in Alzheimer's Disease. *J Mol Neurosci* 2001; 17:137-145. Review.
56. Hirayama A, Horikoshi Y, Maeda M, Ito M, Takashima S Characteristic developmental expression of amyloid beta 40,42 and 43 in patients with Down syndrome. *Brain & Development* 2003; 25:180-185.
57. Schupf N, Zigman WB, Tang MX, Pang D, Mayeux R, Mehta P, Silverman W Change in plasma A β peptides and onset of dementia in adults with Down syndrome *Neurology* 2010; Nov 2; 75(18):1639-44.
58. Prasher VP Sajith SG, Mehta P, Zigman WB, Schupf N Plasma-beta amyloid and duration of Alzheimer's disease in adults with Down syndrome. *Int J Geriatr Psychiatry* 2010; 25(2): 202–207.
59. Jennings D Seibyl J, Sabbagh M, Lai F, Hopkins W, Bullich S, Gimenez M, Reiningger C, Putz B, Stephens A, Catafau AM, Marek K. Age dependence of brain β -amyloid deposition in Down syndrome: An [18F] florbetaben PET study *Neurology* 2015; Feb 3; 84(5): 500-7.
60. Hartley SL, Handen BL, Devenny D, Mihaila I, Hardison R, Lao PJ, Klunk WE, Bulova P, Johnson SC, Christian BT Cognitive decline and brain amyloid- β accumulation across 3 years in adults with Down syndrome. *Neurobiol Aging* 2017; Oct; 58:68-76.
61. Davidson YS, Robinson A, Prasher VP, Mann DMA. The age of onset and evolution of Braak tangle stage and Thal amyloid pathology of Alzheimer's disease in individuals with Down syndrome. *Acta Neuropathol Commun* 2018; Jul 4; 6(1):56.
62. Di Domenico F Pupo G, Mancuso C, Barone E, Paolini F, Arena A, Blarmino C, Schmitt FA, Head E, Butterfield DA, Perluigi M Bach1 overexpression in Down syndrome correlates with the alteration of the HO-1/BVR-a system: insights for transition to Alzheimer's disease. *J Alzheimers Dis* 2015; 44(4): 1107-1120.

63. Barone E, Head E, Butterfield DA, Perluigi M HNE-modified proteins in Down Syndrome: involvement in development of Alzheimer disease neuropathology. *Free Radic Biol Med.* 2017; 111: 262-269.
64. Peters DG Connor JR, Meadowcroft MD The relationship between iron dyshomeostasis and amyloidogenesis in Alzheimer's disease: two sides of the same coin. *Neurobiol Dis* 2015; Sep;81:49-65.
65. Barone E, Arena A, Head E, Butterfield DA, Perluigi M Disturbance of redox homeostasis in Down Syndrome: Role of iron dysmetabolism. *Free Radical Biology and Medicine* 2018; 114 84-93.
66. Antonarakis SE Lyle R, Dermitzakis ET, Reymond A, Deutsch S Chromosome 21 and Down syndrome: from genomics to pathophysiology. *Nat Rev. Genet* 2004; Oct;5(10): 5725-738. Review.
67. Lee JH Lee AJ, Dang LH, Pang D, Kisselev S, Krinsky-McHale SJ, Zigman WB, Luchsinger JA, Silverman W, Tycko B, Clark LN, Schupf N Candidate gene analysis for Alzheimer's disease in adults with Down syndrome *Neurobiol Aging* 2017; 56 : 150-158.
68. Galzitskaya OV Surin AK, Glyakina AV, Rogachevsky VV, Selivanova OM Should the Treatment of Amyloidosis Be Personified? Molecular Mechanism of Amyloid Formation by A β Peptide and Its Fragments *J Alzheimers Dis Rep* 2018; Oct 24;2(1):181-199.
69. Matthias Gralle M, Gralle Botelho M, Wouters FS Neuroprotective Secreted Amyloid Precursor Protein Acts by Disrupting Amyloid Precursor Protein Dimers *J Biol Chem.* 2009; May 29; 284(22): 15016–15025.
70. Habib A, Hou H, Mori T, Tian J, Zeng J, Fan S, Giunta B, Sanberg PR, Sawmiller D, Tan J. Human Umbilical Cord Blood Serum-derived α -Secretase: Functional Testing in Alzheimer's Disease Mouse Models. *Cell Transplant* 2018; 27(3):438-455.
71. Hussain I, Powell DJ, Howlett DR, Chapman GA, Gilmour L, Murdock PR, Tew DG, Meek TD, Chapman C, Schneider K, Ratcliffe SJ, Tattersall D, Testa TT, Southan C, Ryan DM, Simmons DL, Walsh FS, Dingwall C, Christie G ASP1 (BACE2) cleaves the amyloid precursor protein at the beta-secretase site. *Mol Cell Neurosci* 2000; 16(5):609-19.
72. Nistor M, Don M, Parekh M, Sarsoza F, Goodus M, Lopez GE, Kawas C, Leverenz J, Doran E, Lott IT, Hill M, Head E Alpha-and beta-secretase activity as a function of age and beta-amyloid in Down syndrome and normal brain. *Neurobiol Aging* 2007; 28(10):1493-506.
73. Ichinohe A, Kanaumi T, Takashima S, Enokido Y, Nagai Y, Kimura H Cystathionine beta-synthase is enriched in the brains of Down's patients. *Biochem Biophys Res Commun* 2005; 338(3):1547-50.
74. Eto K Asada T, Arima K, Makifuchi T, Kimura H Brain hydrogen sulfide is severely decreased in Alzheimer's disease *Biochem Biophys Res Commun* 2002; May 24 293(5):1485-8.
75. Liu Y Deng Y, Liu H, Yin C, Li X, Gong Q Hydrogen sulfide ameliorates learning memory impairment in APP/PS1 transgenic mice: A novel mechanism mediated by the activation of Nrf2. *Pharmacol Biochem Behav* 2016; 150-151: 207-216.
76. London J Ndiaye FK, Bui LC, Souchet B, Daubigney F, Magnan C, Luquet S, Dairou J, Janel N, Rouch C Alterations in the Serotonin and Dopamine Pathways by Cystathionine Beta Synthase Overexpression in Murine Brain *Mol Neurobiol* 2018; sep20.
77. Kimura R, Kamino K, Yamamoto M, Nuripa A, Kida T, Kazui H, Hashimoto R, Tanaka T, Kudo T, Yamagata H, Tabara Y, Miki T, Akatsu H, Kosaka K, Funakoshi E, Nishitomi K, Sakaguchi G, Kato A, Hattori H, Uema T, Takeda M The DYRK1A gene, encoded in chromosome 21 Down syndrome critical region, bridges between beta-amyloid production and tau phosphorylation in Alzheimer disease. *Hum Mol Genet.* 2007; 16(1):15-23.
78. Wegiel J, Kaczmarek W, Barua M, Kuchna I, Nowicki K, Wang KC, Wegiel J, Yang SM, Frackowiak J, Mazur-Kolecka B, Silverman WP, Reisberg B, Monteiro I, de Leon M, Wisniewski T, Dalton A, Lai F, Hwang YW, Adayev

- T, Liu F, Iqbal K, Iqbal IG, Gong CX et al. Link between DYRK1A overexpression and several-fold enhancement of neurofibrillary degeneration with 3-repeat tau protein in Down syndrome. *J Neuropathol Exp Neurol*. 2011; 70(1):36–50.
79. Souchet B, Guedj F, Penke-Verdier Z, Daubigney F, Duchon A, Herault Y, Bizot JC, Janel N, Creau N, Delatour B, Delabar JM Pharmacological correction of excitation/inhibition imbalance in Down syndrome mouse models. *Front Behav Neurosci* 2015; Oct20, 9: 267.
80. De la Torre R, De Sola S, Hernandez G, Farré M, Pujol J, Rodriguez J, Espadaler JM, Langohr K, Cuenca-Royo A, Principe A, Xicota L, Janel N, Catuara-Solarz S, Sanchez-Benavides G, Bléhaut H, Dueñas-Espín I, Del Hoyo L, Benejam B, Blanco-Hinojo L, Videla S, Fitó M, Delabar JM, Dierssen M; TESDAD study group Safety and efficacy of cognitive training plus epigallocatechin-3-gallate in young adults with Down's syndrome (TESDAD): a double-blind, randomised, placebo-controlled, phase 2 trial. *Lancet Neurol*. 2016; 15(8):801-810.
81. Herault Y Delabar JM, Fisher EMC, Tybulewicz VLJ, Yu E, Brault V. Rodent models in Down syndrome research: impact and future opportunities *Dis Model Mech* 2017; Oct 1;10(10):1165-1186.
82. London J, Rouch C, Bui LC, Assayag E, Souchet B, Daubigney F, Medjaoui H, Luquet S, Magnan C, Delabar JM, Dairou J, Janel N Overexpression of the DYRK1A Gene (Dual-Specificity Tyrosine Phosphorylation-Regulated Kinase 1A) Induces Alterations of the Serotonergic and Dopaminergic Processing in Murine Brain Tissues. *Mol Neurobiol* 2018; 55(5):3822-3831.
83. Wolvetang EW, Bradfield OM, Tymms M, Zavarsek S, Hatzistavrou T, Kola I, Hertzog PJ The chromosome21 transcription factor ETS2 transactivates the beta-APP promoter: implications for Down syndrome *Biochim Biophys Acta* 2003; 1628, 105–110.
84. Helguera P Pelsman A, Pigino G, Wolvetang E, Head E, Busciglio J ets-2 promotes the activation of a mitochondrial death pathway in Down's syndrome neurons. *J Neurosci*. 2005; 25(9):2295-303.
85. Lloret A Badia MC, Giraldo E, Ermak G, Alonso MD, Pallardó FV, Davies KJ, Viña J Alzheimer's amyloid- β toxicity and tau hyperphosphorylation are linked via RCAN1 *J Alzheimers Dis* 2011; 27(4): 701–709.
86. Ermak G and Kelvin J.A. Davies KJA Chronic high levels of the RCAN1-1 protein may promote neurodegeneration and Alzheimer disease 2013; *Free Radic Biol Med* 62: 47–51.
87. Wong H, Levenga J, Cain P, Rothermel B, Klann E, Hoefler C RCAN1 overexpression promotes age-dependent mitochondrial dysregulation related to neurodegeneration in Alzheimer's disease *Acta Neuropathol* 2015;130(6): 829-843.
88. Griffin WS, Sheng JG, McKenzie JE, Royston MC, Gentleman SM, Brumback RA, Cork LC, Del Bigio MR, Roberts GW, Mrak RE Life-long overexpression of S100beta in Down's syndrome: implications for Alzheimer pathogenesis *Neurobiol Aging* 1998; 19:401-405.
89. Royston MC, McKenzie JE, Gentleman SM, Sheng JG, Mann DM, Griffin WS, Mrak RE Overexpression of s100beta in Down's syndrome: correlation with patient age and with beta-amyloid deposition. *Neuropathol Appl Neurobiol* 1999 ; 25:387-393.
90. Shin JH, London J, Le Pecheur M, Weitzdoerfer R, Hoeger H, Lubec G Aberrant neuronal and mitochondrial proteins in hippocampus of transgenic mice overexpressing human Cu/Zn superoxide dismutase 1 *Free Radic Biol Med* 2004; 37(5): 643-53.
91. Le Pecheur M, Bourdon E, Paly E, Farout L, Friguet B, London J. et al Oxidized SOD1 alters proteasome activities in vitro and in SOD1 overexpressing mice *FEBS letters* 2005;579: 3613-8.
92. London J, Le Pecheur M (2016) Effect of SOD1 overexpression on the 20S proteasome during aging *Natural Science* 2016; 8: 295-304.

93. Zhu L, Zhong M, Zhao J, Rhee H, Caesar I, Knight EM, Volpicelli-Daley L, Bustos V, Netzer W, Liu L, Lucast L, Ehrlich ME, Robakis NK, Gandy SE, Cai D Reduction of Synaptojanin 1 Accelerates Clearance and Attenuates Cognitive Deterioration in an Alzheimer Mouse Model *J Biol Chem* 2013; 288(44):32050-63.
94. Martin SB, Dowling AL, Lianekhammy J, Lott IT, Doran E, Murphy MP, Beckett TL, Schmitt FA, Head E et al Synaptophysin and Synaptojanin-1 in Down Syndrome are Differentially Affected by Alzheimer's Disease. *J Alzheimers Dis* 2014; 42(3): 767-775.
95. Arai Y, Ijuin T, Takenawa T, Becker LE, Takashima S Excessive expression of synaptojanin in brains with Down syndrome. *Brain Dev.* 2002; 24(2):67-72.
96. Voronov SV, Frere SG, Giovedi S, Pollina EA, Borel C, Zhang H, Schmidt C, Akeson EC, Wenk MR, Cimasoni L, Arancio O, Davisson MT, Antonarakis SE, Gardiner K, De Camilli P, Di Paolo G Synaptojanin 1-linked phosphoinositide dyshomeostasis and cognitive deficits in mouse models of Down's syndrome. *Proc Natl Acad Sci USA* 2008; 105(27):9415-20.
97. Miranda AM, Herman M, Cheng R, Nahmani E, Barrett G, Micevska E, Fontaine G, Potier MC, Head E, Schmitt FA, Lott IT, Jiménez-Velázquez IZ, Antonarakis SE, Di Paolo G, Lee JH, Hussaini SA, Marquer C. Excess Synaptojanin 1 Contributes to Place Cell Dysfunction and Memory Deficits in the Aging Hippocampus in Three Types of Alzheimer's Disease *Cell Rep* 2018; 23(10): 2967-2975.
98. Gupta M, Dhanasekaran AR, Gardiner KJ Mouse models of Down syndrome: gene content and consequences *Mamm Genome* 2016; 27(11-12): 538–555.
99. Davisson MT, Schmidt C, Akeson EC Segmental trisomy of murine chromosome 16: a new model system for studying Down syndrome *Prog Clin Biol Res* 1990; 360, 263-280.
100. O'Doherty A, Ruf S, Mulligan C, Hildreth V, Errington ML, Cooke S, Sesay A, Modino S, Vanes L, Hernandez D, Linehan JM, Sharpe PT, Brandner S, Bliss TV, Henderson DJ, Nizetic D, Tybulewicz VL, Fisher EM. An aneuploid mouse strain carrying human chromosome 21 with Down syndrome phenotypes *Science* 2005; Sep 23;309(5743):2033-7.
101. Choong XY, Tosh JL, Pulford LJ, Fisher EM Dissecting Alzheimer disease in Down syndrome using mouse models. *Front Behav Neurosci* 2015; Oct 13; 9:268.
102. Hamlett ED, Boger HA, Ledreux A, Kelley CM, Mufson EJ, Falangola MF, Guilfoyle DN, Nixon RA, Patterson D, Duval N, Granholm AC Cognitive Impairment, Neuroimaging, and Alzheimer Neuropathology in Mouse Models of Down Syndrome *Curr Alzheimer Res.* 2016; 13(1): 35-52. Review.
103. Aziz NM, Guedj F, Pennings JLA, Olmos-Serrano JL, Siegel A, Haydar TF, Bianchi DW Lifespan analysis of brain development, gene expression and behavioral phenotypes in the Ts1Cje, Ts65Dn and Dp(16)1/Yey mouse models of Down syndrome *Dis Model Mech* 2018; Jun 12; 11(6) pii: dmm031013.
104. Audrain M, Souchet B, Alves S β APP Processing Drives Gradual Tau Pathology in an Age-Dependent Amyloid Rat Model of Alzheimer's Disease *Cerebral Cortex* 2017; 1–18.
105. Dekker AD, Vermeiren Y, Albac C, Lana-Elola E, Watson-Scales S, Gibbins D, Aerts T, Van Dam D, Fisher EMC, Tybulewicz VLJ, Potier MC, De Deyn PP et al Aging rather than aneuploidy affects monoamine neurotransmitters in brain regions of Down syndrome mouse models *Neurobiol Dis* 2017; 105:235-244.
106. Xu H, Wang Z, Zhu L, Sui Z, Bi W, Liu R, Bi K, Li Q Targeted Neurotransmitters Profiling Identifies Metabolic Signatures in Rat Brain by LC-MS/MS: Application in Insomnia, Depression and Alzheimer's Disease *Molecules* 2018;23(9): 2375.
107. Das D, Phillips C, Hsieh W, Sumanth K, Dang V, Salehi A Neurotransmitter-based strategies for the treatment of cognitive dysfunction in Down syndrome *Prog neuropharmacol Bio Psychiatry* 2014; 54:140-148.
108. Phillips C, Fahimi A, Das D, Mojabi FS, Ponnusamy R, Salehi A Noradrenergic system in Down syndrome and

Alzheimer's disease A target for therapy *Curr Alzheimer res.* 2016; 13(1) 68-83.Review.

109. Lee NC Chien YH, Hwu WL A Review of Biomarkers for Alzheimer's Disease in Down Syndrome. *Neurol Ther* 6(Suppl 1) 2017; 69-81.

110. Janel N, Alexopoulos P, Badel A, Lamari F, Camproux AC, Lagarde J, Simon S, Feraudet-Tarisse C, Lamourette P, Arbones M, Paul JL, Dubois B, Potier MC, Sarazin M, Delabar JM Combined assessment of DYRK1A, BDNF and homocysteine levels as diagnostic marker for Alzheimer's disease *Transl Psychiatry* 2017; 7(6): e1154.

111. Zana M, Janka Z, Kálmán J Oxidative stress: a bridge between Down's syndrome and Alzheimer's disease *Neurobiol Aging* 2007; May 28:648-76.Review.

112. Parisotto EB, Garlet TR, Cavalli VL, Zamoner A, da Rosa JS, Bastos J, Micke GA, Fröde TS, Pedrosa RC, Wilhelm Filho D Antioxidant intervention attenuates oxidative stress in children and teenagers with Down syndrome *Res Devl Disabil* 2014 Jun; 35 (2014) 1228-1236.

113. Farina N, Llewellyn D, MGEKN I, Tabet N. *Cochrane database Syst Rev.* 2017; 18;4: CD002854.

114. Hunter CL, Bachman D, Granholm AC Minocycline prevents cholinergic loss in a mouse model of Down's syndrome. *Ann Neurol.* 2004Nov; 56(5):675-88.

115. Tramutola A, Lanzillotta C, Barone E, Arena A, Zuliani I, Mosca L, Blarzino C, Butterfield DA, Perluigi M, Di Domenico F Intranasal rapamycin ameliorates Alzheimer like cognitive decline in a mouse model of Down syndrome. *Translational Neurodegeneration* 2018; 7:28.

116. Zorrilla de San Martin J Delabar JM, Bacci A, Potier MC GABAergic over-inhibition, a promising hypothesis for cognitive deficits in Down syndrome *Free Radical Biology and Medicine* 2018 ; 114: 33–39.

117. Granholm AC Sanders L, Seo H, Lin L, Ford K, Isacson O Estrogen alters amyloid precursor protein as well as dendritic and cholinergic markers in a mouse model of Down syndrome. *Hippocampus* 2003; 13(8):905-14.

118. Hunter CL Bimonte-Nelson HA, Nelson M, Eckman CB, Granholm AC Behavioral and neurobiological markers of Alzheimer's disease in Ts65Dn mice: effects of estrogen. *Neurobiol Aging* 2004; 25(7): 873-84.

119. Jiang Y, Rigoglioso A, Peterhoff CM, Pawlik M, Sato Y, Bleiwas C, Stavrides P, Smiley JF, Ginsberg SD, Mathews PM, Levy E, Nixon RA. Partial BACE1 reduction in a Down syndrome mouse model blocks Alzheimer-related endosomal anomalies and cholinergic neurodegeneration: role of APP-CTF *Neurobiol Aging* 2016; 39:90-8.

120. Sarko DK, McKinney CE. Exosome: Origins and therapeutic potential for neurodegenerative disease *Frontiers neurosci* 2017; Feb 27;11:82.

121. Gauthier SA, McKinney CE Enhanced exosome secretion in Down syndrome brain : a protective mechanism to alleviate neuronal endosomal abnormalities *Acta Neuropathologica Communications* 2017; 5:65.

122. Hamlett ED, Ledreux A, Potter H, Chial HJ, Patterson D, Espinosa JM, Bettcher BM, Granholm AC Exosomal biomarkers in Down syndrome and Alzheimer's disease *Free Radic Biol Med.* 2018 Jan; 114:110-121; doi: 10.1016/j.freeradbiomed. 2017.08.028. Epub 2017 Sep 5. Review.

123. Sansevero G, Begenisic T, Mainardi M, Sale A. Experience-dependent reduction of soluble β -amyloid oligomers and rescue of cognitive abilities in middle-age Ts65Dn mice, a model of Down syndrome. *Exp Neurol* 2016; 283(Pt A):49-56.

124. Pavel V, Madani R, Rey-Bellet L, Pihlgren M, Becker A, Plassard A, Vuillermot S, Giriens V, Nosheny RL, Kleschevnikov AM, Valletta JS, Bengtsson SK, Linke GR, Maloney MT, Hickman DT, Reis P, Granet A, Mlaki D, Lopez-Deber MP, Do L, Singhal N, Masliah E, Pearn ML, Pfeifer A, Muhs A, Mobley WC. An Anti- β -Amyloid Vaccine for Treating Cognitive Deficits in a Mouse Model of Down Syndrome *PLoS One* 2016; 11(3): e0152471.

125. Gueroux M, Fleau C, Slozeck M, Laguerre M, Pianet I Epigallocatechin 3-Gallate as an Inhibitor of Tau Phosphorylation and Aggregation: A Molecular and Structural Insight. *J Prev Alzheimers Dis* 2017; 4(4):218-225.
126. Xicota L, Rodriguez-Morato J, Dierssen M, de la Torre R Potential Role of (-)-Epigallocatechin-3-Gallate (EGCG) in the Secondary Prevention of Alzheimer Disease. *Curr Drug Targets* 2017;18(2):174-195.
127. Catuara-Solarz S, Espinosa-Carrasco J, Erb I, Langohr K, Gonzalez JR, Notredame C, Dierssen M Combined Treatment With Environmental Enrichment and (-)-Epigallocatechin-3-Gallate Ameliorates Learning Deficits and Hippocampal Alterations in a Mouse Model of Down Syndrome *eNeuro* 2016; 3(5): 0103-16.2016.
128. Neumann F, Gourdain S, Albac C, Dekker AD, Bui LC, Dairou J, Schmitz-Afonso I, Hue N, Rodrigues-Lima F, Delabar JM, Potier MC, Le Caër JP, Touboul D, Delatour B, Cariou K, Dodd RH. DYRK1A inhibition and cognitive rescue in a Down syndrome mouse model are induced by new fluoro-DANDY derivatives *Sci Rep* 2018; 8: 2859. Published online 2018 Feb 12.
129. Nguyen TL, Duchon A, Antigoni Manousopoulou A, et al Correction of cognitive deficits in mouse models of Down syndrome by a pharmacological inhibitor of DYRK1A *Dis Model Mech.* 2018;11(9): dmm035634.
130. Chang CY, Ting HC, Liu CA, Su HL, Chiou TW, Harn HJ, Lin SZ, Induced Pluripotent Stem Cells: A Powerful Neurodegenerative Disease Modeling Tool for Mechanism Study and Drug Discovery *Cell Transplant* 2018 ; 27(11): 1588–1602.
131. Nakano-Kobayashia A, Awaya T, Kii I, Sumida Y, Okuno Y, Yoshida S, Sumida T, Inoue H, Hosoya T, Hagiwara M. Prenatal neurogenesis induction therapy normalizes brain structure and function in Down syndrome mice. *PNAS* 2017 ; 114, 38 10268-10273.
132. Dashinimaev EB, Artyuhov AS, Bolshakov AP, Vorotelyak EA, Vasiliev AV Neurons Derived from Induced Pluripotent Stem Cells of Patients with Down Syndrome Reproduce Early Stages of Alzheimer's Disease Type Pathology *in vitro.* *J Alzheimers Dis* 2017; 56(2): 835-847.