

Advances in Biotechnology

Chapter 3

A Comprehensive Review on the Physical and Chemical Properties of the Three Generations of Biofuels

Mehdi Ardjmand^{1} and Farid Jafarighighi¹, Mohammad Salar Hassani¹, Neda Bazel¹, Hasanali Bahrami²*

¹Department of Chemical Engineering, South Tehran Branch, Islamic Azad University, Tehran, Iran.

²Department of Mechatronics, Arak University, Iran

**Correspondence to: Mehdi Ardjmand, Head of Department of Chemical Engineering, South Tehran Branch, Islamic Azad University, Tehran, Iran.*

Tel: +989121513117; Email: m_arjmand@azad.ac.ir

Abstract

The shortage of fossil fuel resources and the dramatic increase in population have raised many concerns about fuel supply in the years ahead. Researchers are now focusing on renewable fuels, and biodiesel is one of those renewables. Four generations of biodiesel have been reported today and many studies have been done to optimize and enhance their performance. The present review article examines the physical and chemical properties of three generations of biodiesel. It was observed that the physical and chemical properties of the biodiesel vary based on the feed stocks and have a significant effect on the dynamic characteristics of emission level and performance of engine. All properties have the highest and lowest ranges for each feed. All the oils that have been studied for three generations to date have been fully reported, and these properties have been studied and compared for each of the three generations.

Keywords: Biodiesel; First Generation; Second Generation; Third Generation; Physical and Chemical Properties

1. Introduction

policies and dramatic fuel price changes in fuel-producing countries have caused many crises in the world. Lack of adequate resources and fossil fuel contamination in the world are other causes of global energy issues. (**Figure 1**) shows the increase of the demanding for the crude oil and the price changes in the world. These reasons have made the need for alternative fuel in the world completely necessary. Nowadays, biofuels have attracted a lot of attention as an alternative to fossil fuels [1-5]. Biofuels are included several advantages and the most important of them are related to the environmental benefits. Biodiesel can diminish emissions that cause environmental difficulties such as acid rain and global warming. Also, health issues as a consequence of emissions exposure are significantly declined by the cleaner emissions of biodiesel [5, 6]. Biodiesel is the non-petroleum based diesel fuel. It is contained of the mono-alkyl esters of the long-chain fatty acids derived from the renewable lipid sources [7-9]. Quality of biofuels is always dependent on many factors. Some of them are included the feedstock, fatty acid composition, production process, handling and storage, and postproduction parameters [10].

The close similarities between the properties of biodiesel and diesel fuels make that biodiesel is a good alternative to diesel fuels. The viscosity of biodiesel is so close to the diesel fuel. The conversion of triglycerides into ethyl or methyl esters via the transesterification procedure diminishes the molecular weight and viscosity and rises the volatility gradually. The cetane number is around 50-60 for biodiesel and it's higher than diesel fuels, however, the heating value of the diesel fuel is greater than biodiesel. The flashpoint and density of biodiesel are much higher than diesel fuels, while the cloud point for diesel fuels normally is better than biodiesel fuel. The sulfur compounds in petrodiesel provide much of the lubricity, however, Biodiesel comprises virtually no sulfur and this is frequently applied as the additive to ultra-low-sulfur diesel (ULSD) fuel to help with lubrication [11-13].

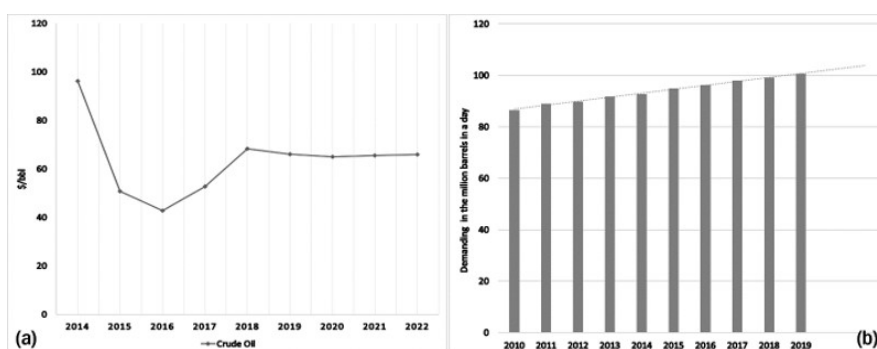


Figure 1: World Bank: average crude oil prices (a) and daily demand for crude oil worldwide (b)

1.1. Biodiesel feedstocks

The different types of feedstocks are used for the production of biodiesel. The choice of feedstocks relies on the economic aspects and availability of the concerned country. The major biodiesel feedstocks for different regions of the world are shown in (**Figure 2**). Four genera-

tions of biodiesel fuels are applied worldwide.



Figure 2: Main biofuel producers by region

The first generation of biodiesel is edible oils such as palm, soybeans, rapeseed, and sunflower oil. Some of the main advantages of the first generation of biodiesel are shown in Fig 3. However, the use of edible oil sources as biodiesel fuel has caused great concern in the world. These concerns include the possibility of food shortages in the world and rising food prices. This generation has also required arable land for the production and it creates serious ecological imbalances due to that countries start cutting down forests for plantation purposes. Therefore, the demand for biodiesel increases, it will cause severe damage to the environment and wildlife, due to the greater need for arable land and larger scale deforestation [3, 5].

The second generation of biodiesel is non-edible oils. The mahua, jatropha, tobacco seed, jojoba oil are examples of second-generation biodiesel. This generation of biodiesel has many advantages, as shown in Fig 3. This generation also has some limitations for worldwide using. They may not be abundant enough to substitute transportation fuels. The performance of this generation has some restriction in cold temperatures [14].

The third generation of biodiesel are included microalgae, animal fats, and waste cooking oils. Some advantages of this generation are shown in (Figure 3) [3]. While, this generation requires huge amount of money for producing. According to research, the production of algae biofuel still requires a lot of work, mainly in the process of the oil extraction and low yields as well as it emits captured carbon dioxide.

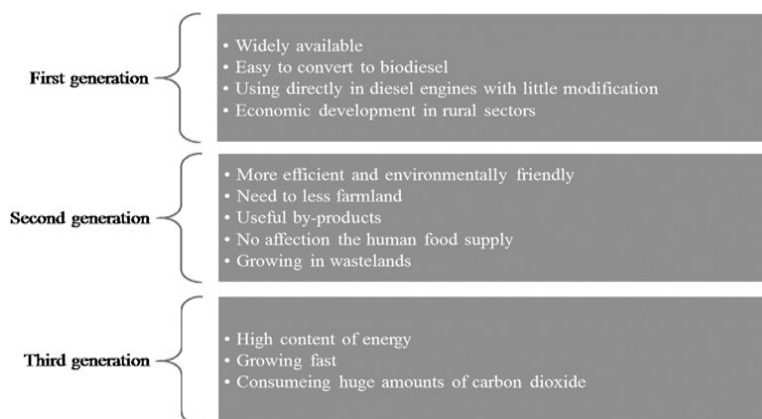


Figure 3: The advantages of the first, second, and third generation

Limited information is available on fourth generation biodiesel. This has led to a complete lack of scrutiny in this area. The Synthetic Genomics Company is applied genetic engineering in production of biofuel. The genetically modified microorganisms is used to generate fuel directly from the carbon dioxide on the industrial scale. Furthermore, the fourth generation biofuels are gained from genetically modified crops in which they spend more carbon dioxide from the atmosphere than they release over combustion which makes it a carbon negative fuel.

1.2. The standards of biodiesel in the worldwide

Several studies indicated that the physical properties of biodiesel have a huge effect on emission and combustion. The physical and chemical properties of the produced biodiesel must reach the standard value defined in the different regions for using. Some of the standards are included EN 14213/EN 14214, SANS 1935, JASO M360, ASTM D 6751, ANP 42, and IS 15607 which used in EU, South Africa, Japan, U.S, Brazil, and India, respectively [10]. Some of the most important physical characteristics of biodiesel are included density, cetane number, kinematic viscosity, flash point, pour point and cloud point, calorific value, acid value, copper strip corrosion, ash content, sulfur content, glycerine, and oxidation stability [12, 15]. Standards set guidelines for testing the biodiesel fuels and propose the proper ranges for different physical and chemical properties of the fuel.

There have been limited studies of the physical properties of biodiesel, but no reports to date have examined the physical properties of all three generations of biodiesel and comparing them to one another. A recent study has surveyed all the physical properties of three-generations of biodiesel from all oil sources which effect on the engine performance and emission features and also shown comparisons between them. The report also mentioned the lowest highest values of properties and shows the range for all biodiesels.

1.3. Characteristics and properties of three generations of biodiesel

The important physical properties of three-generation biodiesels are summarized in Table 1, 2, and 3. The Physicochemical property ranges are illustrated for pure biodiesels. All tables show the most important properties of biodiesels such as Density (kg/m^3), Kinematic Viscosity 40°C (mm^2/s), Calorific Value (Mj/Kg), Higher (gross) Heating Value (Mj/Kg), Lower (net) Heating Value (Mj/Kg), Acid Value (Neutralization number) (mg KOH/g), Flash Point ($^\circ\text{C}$), Cetane Number, Oxidation Stability (h), Cloud Point ($^\circ\text{C}$), and Pour Point ($^\circ\text{C}$).

There have been numerous reports on the types of vegetable oils and different amounts have been reported. About 69 first generation oils that have been the most researched are shown in (Table 1).

Taramira (<i>Eruca sativa</i>)	at 15°C: 881.1 [183] at 40°C: 871±0.15 [184]	5.71±0.21, 5.9 [184] [183]	0.40 [183]	197.3±2.1 [184]	48, 59.08±1.34 [183] [184]	6 [183]	1.5±0.7 [184]	-2.97±0.17 [184]
Tea seed (<i>Camellia</i>)	at 20°C: 884 [185]	4.95 [185]	37.512 [185]	120 [185]	52 [185]
Thistle (<i>Silybum marianum</i>)	at 20°C: 863 [186] at 15.6°C: 0.8788 ± 0.0012 [187] at 40°C: 899 [188]	4.46, 6.32 [186] [188]	16.984 [188]	...	0.10 ± 0.01, 0.44 [187] [186]	115.0 ± 0.50, 153 [187] [186]	44, 51 [188] [186]	2.1 [186]	-1, 7 [186] [188]	-6, 10 ± 1.0 [188] [187]
Tobacco seed (<i>Nicotiana tabacum</i>)	at 15°C: 886.8, 888.5 [189] [137] at 25°C: 870 [190] at 20°C: 882 [191]	3.5, 5.2 [189] [191]	38, 39.811 [190] [189]	...	0.3, 0.66 [137] [191]	165.4 [137]	51, 52 [189] [190]	0.8 [137]
Tomato seed	at 40°C: 915.1 [192]	28 [192]	35.9 [192]	189 [192]	54.71 [192]
Castor	at 20°C: 917 [90]	13.5, 14.4 [43] [90]	0.42, 3.9 [43] [90]	165 to 186.5 [193]
Colza (<i>Brassica rapa</i>)	at 20°C: 882±25 [30]	4.02±0.12 [30]	0.22±0.03 [30]	185±3 [30]	...	11.2±0.1 [30]
Radish	...	4.6 [37]	53 [37]
Tigernut (Nut-sedge, <i>Cyperus esculentus</i>)	at 30°C: 866 [194]	2.34 [194]	186 [194]

Niger seed (Asteraceae , Guizotia abyssinica)	at 15°C: 890 [163]	4.10, 4.30 [163] [164]	128, 157 [163] [164]	57 [164]	1.02 [164]	3, 4 [163] [164]	2.5 [163]
Okra seed (Abelmoschus esculentus)	at 15°C: 870 [104]	4.1 [104]	0.22 [104]	126 [104]	...	1.6 [104]	6 [104]	6 [104]
Papaya seed	at 40°C: 840, 900 [165] [166] at 15°C: 895 [134]	3.53, 6 [165] [167] 42.085 [41]	...	38.49, 38.97319 [165] [134]	0.35, 0.72 [167] [166]	112, 147 [165] [134]	77.3, 48.29 [168] [165]	...	1, 2 [167] [168]	-1, 1 [167] [168]
Pequi (Caryocar brasiliense)	at 20°C: 69.69 ± 0.07, 866.3 [169] [170]	4.27 ± 0.03, 5.64 [169] [170]	0.7909 [170]	4.5±0.3, 5.85 [169] [170]
Pomegranate seed (Punica granatum)	at 40°C: 892 [171] at 20°C: 895.2 [135]	3.31, 5.65 [135] [171] 39.49, 40.048 [135] [171]	130 [171]	26.1, 45.02 [135] [171]
Poppyseed	893 [41]	3.5, 4.63 [49] [41]
Pracaxi (Pentaclethra macroloba)	at 20°C: 869±6 [172]	5.8 ±0.03 [172]	3.2±0.2 [172]
Rice bran	at 40°C: 868.1 [26] at 15°C: 876±15.7, 892 [173] [174]	4.14, 5.37 [175] [114] 39.957, 42.2 [26] [176]	...	39.5, 43.10±0.98 [114] [173]	0.09, 0.586 [175] [26]	174.5, 183 [26] [176]	73.6, 51 [26] [114]	1.61, 1.63±0.12 [86] [173]	-10, 9 [174] [176]	-11, -2.00±0.14 [174] [173]
Sacha inchi	...	4.66 [177]	0.39 [177]
Sapote	at 40°C: 864 [178]	4.5, 5.43 [179] [178]	...	37.12 [179]	0.16 [179]	174 [178]	52 [178]	-6 [179]
Shea butter	At 5°C: 877, 883 [180] [181]	4.42, 5.93 [180] [181]	38.9 [182]	37.93, 37.98 [180] [182]	0.16, 0.28 [181] [180]	130, 171 [181] [180]	47, 58 [181] [180]	...	3,12 [182] [181]	3, 10 [180] [181]

Cocklebur (Xanthium)	at 15°C: 896.89 [148]	6.877 [148]	38.527 [148]	166 [148]	42.3 [148]	...	-1 [148]	-19 [148]
Coriander seed	...	4.21 [92]	...	40.1 [92]	37.5 [92]	0.1 [92]	14.6 [92]	...	-19 [92]
Date seed	at 15°C: 870 to 890 [149]	3.7 to 4.2 [149]	122 to 131 [149]	53.56 [149]	...	3 to 7 [149]	...
Dika (Irvingia gabonensis)	...	3.2 [150]	39 [150]	...	39 [151]	0.01 [150]	140 [150]	-14 [151]	-6 [151]
False flax (Camelina sativa)	...	2.9 to 3.15, 4.37 [152] [51]	45.05 to 46.15 [152]	0.04 [51]	179 [51]	41.26 to 51.17 [152]	...	4.1 [51]	-11 to -8, 0 [152] [51]
Grape seed	at 20°C: 882 [135] at 32°C: 890 [126]	4.04, 4.1 [135] [38]	39.73 [135]	0.27, 0.31 [38] [126]	175 [38]	48, 48.6 [38] [135]	0.5 [38]	...	-22 [126]
Hemp	at 15°C: 884, 891.5 [153] [154] at 40°C: 858, 872 [155] [156]	3.48, 4.23 [155] [156]	39.81 [155]	0.01, 0.67 [156] [153]	120, 175 [154] [157]	-5, -2.5 [153] [154]	-17, -4 [155] [154]
Kapok seed (Ceiba pentandra)	at 15°C: 876.9, 890 [158] [104] at 40°C: 875 [159]	4.2, 5.4 [104] [159]	40.493 [158]	36.292 [159]	...	0.24, 0.38 [104] [158]	156, 167 [159] [104]	57.2 [158]	0.8, 4.42 [104] [158]	2, 3 [104] [158]	1, 2.8 [104] [158]
Kenaf seed (Hibiscus cannabinus)	at 15°C: 879.5 [160]	4.8 [160]	0 [160]	...	54 [160]	0.35 [160]	3.8 [160]	1.7 [160]
Marula (Sclerocarya birrea)	at 25°C: 877 [161]	4.12 [161]	171 [161]	6 [161]	3 [161]
Meadowfoam seed	...	6.18, 6.22 [162] [51]	...	40.591 [162]	38.499 [162]	0.02, 0.06 [162] [51]	205 [51]	-6.6, -6 [51] [162]	-10 [162]
Mustard	at 40°C: 866 [36]	4.1 [36]	...	41.3 [36]	169 [36]

Açaí (Euterpe oleracea)	...	4.5 [138]	200 [138]	...	1.5 [138]
Black seed (Nigella sativa)	at 15°C: 886.1 [139] at 40°C: 867.7 [139]	4.5026 [139]	39.967 [139]	0.26 [139]	...	172.5 [139]	...	1.32 [139]	-1 [139]	-1 [139]
Blackcurrant seed (Ribes nigrum)	at 15°C: 892.6 [140]	3.84 [140]	0 [140]	40.5 [140]	1.67 [140]	-2.2 [140]	-12 [140]
Borage seed (Borago officinalis)	at 15°C: 887.09 [140]	4 [140]	0 [140]	45.6 [140]	1.67 [140]	4.5 [140]	-2 [140]
Evening promise (Oenothera biennis)	at 15°C: 878 [141]	5.68 [141]	0.37 [141]	...	196 [141]	51 [141]	...	7 [141]	4 [141]
Flaxseed (linseed, Linum usitatissimum)	at 40°C: 870, 888.2 [32] [142]	2.8, 4.07 [32] [25]	...	40.84, 41.82 [32] [142]	37.267 [25]	142, 184 [36] [138]	41, 55 [37] [142]	2.2 [138]
Apricot	at 15°C: 884.3 [143]	4.92 [143]	39.95 [143]	111 [143]
Apple seed	at 20°C: 881 [135] at 40°C: 865 [144]	4.12, 5.9 [135] [144]	37.51, 40.48 [145] [144]	0.34 [144]	...	150, 161 [145] [144]	50.4 [135]	...	2 [145]	5 [145]
Avocado	at 15°C: 877.68 [146]	4.9581 [146]	41.33 [146]	184 [146]	-7 [146]
Babassu (Attalea speciosa)	...	3.18 [147]	0.0778 [147]	...	112 [147]
Ben (Moringa oleifera)	at 40°C: 859.6, 869.6 [22] [63] at 15°C: 869.6, 885.8 [56] [86]	4.1264, 5.0735 [86] [27]	40.05 [22]	39.888, 40.115 [86] [27]	...	0.05, 0.185 [56] [26]	...	150.5, 180.5 [63] [56]	56, 67.07 [56] [26]	4.45, 26.2 [56] [63]	0, 19 [86] [56]	19 [27]

Macadamia	at 15°C: 859.2, 868 [114] [115]	4.4, 4.57 [37] [115]	39.88 [115]	39.9 [114]	...	0.08 & 0.055, 0.15 [116] [115]	135, 178.5 [115] [114]	55, 59.5 & 57.5 [37] [116]	1.97 & 2.06, 3.35 [116] [114]	-1, 8 [117] [114]	-3, 5 & 1 [115] [116]
Mongongo nut (Manketti, Schinziophyton rautaneuii)	at 15°C: 876, 878 [118] [119] at 25°C: 869 [120]	3.72, 4.43 [120] [118]	36.97, 37.82 [118] [119]	41.2 [120]	...	0.08, 0.35 [118] [120]	150, 165 [119] [120]	49.1, 51 [118] [120]	4.75 [119]	-3.8, 1.2 [119] [118]	-6.2, -4.1 [119] [118]
Pistachio	at 15°C: 860, 880 [121] [122]	3.44, 4.71 [121] [122]	0.16, 0.187 [122] [121]	148, 168 [122] [121]	52, 53.94 [122] [121]	8.3 [122]	-2 [122]	-1 [122]
Walnut	at 40°C: 864 [36] at 15°C: 864 [89]	3.88, 4.11 [68] [89]	39.47, 41.18 [68] [89]	41.32 [36]	...	0.03 [68]	170 [36]	...	2.9 [68]	-6.1 [68]	-10 [68]
Lemon	at 40°C: 853 [123]	1.06 [123]	41.510 [123]
Orange	at 40°C: 812, 876 [124] [125] at 32°C: 892 [126] at 15°C: 852 [127]	1.04, 5.6 [127] [126]	38.158, 42.8 [125] [124]	0.25 [126]	29, 78.5 [127] [125]	37, 49 [125] [124]	-25, -10 [126] [125]
Bitter ground (Momordica charantia)	at 15°C: 889 ± 10.50 [128]	4.48 ± 1.02 [128]	0.40 ± 0.39 [128]	162 ± 11.20 [128]	64.0 ± 4.80 [128]	1.99 ± 0.85 [128]	9.00 ± 2.12 [128]	15.00 ± 1.82 [128]
Egusi seed (Cucumeropsis mannii naudin)	at 15°C: 883, 884±8.75 [129] [99]	3.83, 3.91±0.014 [129] [99]	...	39.97, 42±1.2 [129] [99]	...	0.11±0.01, 0.19 [99] [129]	142±2.82 [99]	53.66, 55 [99] [108]	1.32±0.1, 1.41 [99] [129]	0.5±0.1 [99]	...
Pumpkin seed	at 15°C: 883.7 [130]	4.1, 4.41 [37] [130]	40.21 [131]	38.08, 40.84 [130] [132]	...	0.48 [130]	120, 175 [130] [131]	47, 60.01 [37] [132]	7.2 [131]	-18 [131]	-32 [131]
Watermelon seed (Citrullus vulgaris)	at 15°C: 800, 893 [133] [134] at 20°C: 880.6 [135]	1.05, 5.33 [133] [134]	39.74, 39.85 [135] [136]	39.36195 [134]	...	1.387, 3.66 [136] [137]	107, 143 [133] [134]	44.47, 54.682 [133] [136]	...	-1 [133]	-3 [133]

Soybean	at 40°C: 855, 877 [33] [32] at 20°C: 871±20, 885 [30] [90] at 15°C: 881.1, 887 [53] [41]	3.5, 5.75 [32] [91]	39.72 to 40.08, 40.297 [44] [41]	39.85, 45.07 [92] [53]	37.38, 37.75 [92] [33]	0.04±0.01, 0.45 [44] [91]	152, 202.5 [41] [27]	58.1, 46 [93] [37]	1.3, 6±0.3 [38] [30]	-2, 4 [33] [93]	-6, 1 [33] [27]
Sunflower	at 40°C: 863, 874 [36] [32] at 20°C: 832±4 [30] at 15°C: 880, 888.4, [41] [94]	3.2, 5.241 [49] [95]	38.1 to 38.472, 42.02 [44] [96]	41.03, 41.33 [32] [36]	37.532 [25]	0.04±0.01, 1.3 [44] [95]	125, 192±2 [41] [30]	47, 54 to 58 [37] [44]	0.5, 1.73 [95] [93]	-14, 5±1 [96] [44]	-16, -2±1 [44] [96]
Almond	at 25°C: 881 [58] at 15°C: 882±0.011, 887 [97] [98]	4.2, 4.726 [38] [58]	41.761 [58]	0.10±0.02, 0.44 [97] [98]	145, 173 [58] [98]	44.6, 58 [98] [99]	3, 3.1 [38] [99]	-3, 11 [99] [88]	-9±0.5, -2 [97] [88]
Beech nut (Fagus sylvatica)	at 15°C: 876 [100]	3.8 to 4 [100]	173 [100]	49 [100]
Brazilnut (Bertholletia excelsa)	...	4.3, 4.56 [37] [101]	0.15 [101]	170.3 [101]	53 [37]
Cashew	at 40°C: 863.6, 909.3 [102] [103] at 15°C: 880, 884 [104] [105] at 18°C: 882, 883.5 [106] [107] at 20°C: 890 [19]	3.85, 10.3 [107] [103]	37.51, 39.52 [105] [54]	34.3 [103]	...	0.25, 3.69 [104] [103]	113, 170 [104] [103]	51, 63 [37] [108]	5.6 [104]	-5, 10 [103] [107]	-1, 7.9 [103] [107]
Hazelnut	at 40°C: 875 [36] at 15°C: 863, 884.8 [109] [110]	3.097, 5.48 [109] [110]	37.23, 41.172 [111] [110]	39.889, 41.12 [112] [36]	39.75 [109]	0.05, 0.23 [68] [96]	152, 183 [36] [89]	52.6, 55 [109] [111]	7.6 [68]	-14.7±0.5, -9.3 [113] [68]	-17, -13 [96] [68]

Olive	at 40°C: 860 [36]	4.18, 6.18 [36] [54]	39.92 [54]	41.35 [36]	...	0.13 [38]	178, 204 [38] [36]	57, 61.72 [38] [27]	3.3 [38]
Palm	at 40°C: 858.9, 875 [22] [55] at 15°C: 858.9, 879.7 [56] [57] at 25°C: 884 [58]	2.8, 5.64 [23] [25]	36.5, 40.322 [23] [58]	38.3, 41.3 [59] [60]	37.13, 37.92 [61] [57]	0.01±0.01, 0.4 [44] [21]	125, 214.5 [23] [27]	44, 72.09 [59] [25]	2.41, 23.56 [22] [27]	3, 17±1 [62] [44]	3, 15±1 [63] [44]
Peanut (Arachis hypogea, Ground nut)	at 40°C: 861.1, 992 [39] [64] at 15°C: 848.5, 884 [65] [41] at 20°C: 877.1±2, 877.3 [66] [67]	4.29, 5.908 [39] [64]	39.98, 40.297 [68] [41]	0.08, 0.28 [68] [65]	156, 192 [67] [64]	53, 58.9±4.1 [38] [66]	2, 21.1 [38] [68]	0, 17.8 [65] [68]	-8, 15 [65] [68]
Rapeseed	at 40°C: 857, 872 [36] [32] at 20°C: 879.35 [29] at 15°C: 884, 880 [69] [70]	3.3, 5.51 [49] [71]	37.23, 38.7 [69] [72]	41.31, 41.55 [32] [36]	38.177, 39 [71] [73]	0.03, 0.33 [69] [74]	130, 180 [74] [36]	51, 55 [71] [38]	2, 8.1 [38] [74]	-5 [70]	-13, -10 [70] [69]
Safflower	at 40°C: 866 [36] at 15°C: 860, 890.73 [75] [76] at 20°C: 888, 891 [77] [78]	2.9, 5.8 [49] [79]	38.122, 40.71 [80] [41]	40.604, 45.21±1.58 [81] [82]	38.122, 39.322 [77] [79]	0.06, 0.39 [83] [75]	129, 187 [84] [85]	42.07, 53.16 [76] [75]	0.9, 2.66 [70] [81]	-5, -2±0.05 [76] [82]	-24, -7 [85] [81]
Sesame	at 40°C: 866.9, 884.8 [86] [26] at 15°C: 867.2, 884.8 [87] [88]	3.04, 4.3998 [36] [26]	39.996, 40.7 [26] [89]	39.996, 40.9 [86] [32]	...	0.3 [87]	145, 208.5 [36] [26]	50.48 [87]	1.135, 1.14 [86] [26]	-6 to 1 [26]	-14 to 1 [26]

Table 1: Properties of the first generations of biodiesels.

Biodiesels	Physicochemical property ranges										
	Density (kg/m ³)	Kin. Viscosity 40°C (mm ² /s)	Calorific Value (Mj/Kg)	Higher (gross) Heating Value (Mj/Kg)	Lower (net) Heating Value (Mj/Kg)	Acid Value (Neutralization number) (mg KOH/g)	Flash Point (°C)	Cetane Number	Oxidation Stability (h)	Cloud Point (°C)	Pour Point (°C)
Coconut (Coconut pulm, Copra)	at 40°C: 800, 869.5 [16] [17] at 15°C: 873.3 [18] at 20°C: 870 [19]	2.61, 4.0927 [20] [21]	38, 41.9 [22] [23]	37.26, 39.95 [24] [16]	57.743 [25]	0.106, 0.35 [26] [21]	108, 120.5 [25] [22]	37, 63.73 [24] [19]	5.12, 9.2 [22] [18]	-3, 1 [18] [27]	-12, -4 [18] [22]
Corn (Maize)	at 15°C: 864.2, 879.65 [28] [29] at 20°C: 865±12, 890 [30] [31] at 40°C: 868, 880 [32] [33]	2.45 to 2.56, 4.89 [34] [29]	39.12, 44.92 to 45.06 [31] [34]	39.93, 41.14 [35] [32]	37.679, 38.48 [25] [33]	0.03±0.002, 0.3 [30] [28]	154, 188±4 [36] [30]	46, 58.37 to 59 [37] [34]	1.2, 6.5±0.3 [38] [30]	-15, -3 [28] [33]	-5 to -2 [34]
Canola	at 40°C: 853.6, 875±0.6 [39, 40] at 15°C: 883, 883.2 [41] [42]	2.56 to 2.84, 4.6 [34] [43]	44.65 to 44.93, 37.3 to 39.87 [34] [44]	25.11±0.29, 40.195 [40] [27]	...	0.01±0.01, 0.35 [44] [43]	105, 186.5 [41] [27]	48 to 56 [44]	7.08, 12 [27] [42]	-3, 1 to 2 [27] [40]	-4 to -1, -8 to -10 [34] [40]
Cotton seed	at 40°C: 871 [36] at 20°C: 878±9, 882±1 [30] [45] at 15°C: 850, 887 [46] [47] at 25°C: 875±15.7 [48]	3.1, 6 [49] [46]	39.19, 39.524 [47] [41]	41.68, 48.18 [46] [36]	36.896 [50]	0.02, 0.58 [51] [47]	150±3, 200 [48] [50]	48, 57.1 [52] [53]	1.83±0.12, 4.9±0.8 [48] [45]	-5, 7±0.11 [52] [48]	-4, 6±0.15 [46] [48]

Nerium oleander(Thevetia peruviana)	at 15°C: 875,880 [240] [241]	4.3, 4.33 [241] [240]	42.4, 44.986 [241] [240]	42.279 [240]	...	0.057, 0.66 [240] [241]	75, 178 [240] [241]	61.5, 71.5 [240] [241]	6.5 [241]	-3, 15 [241] [240]	-7, 3 [241] [240]
cebera odollam(sea mango)	at 15°C: 880 [242]	4.5 [242]	39.095 [242]	138 [242]
nagchampa	at 40°C: 876.8,877 [243] [244]	2.64, 4.72 [244] [243]	39.882 [243]	0.41, 0.76 [244] [243]	151, 159 [243] [244]	51.9 [243]	...	13 [243]	10 [243]
Croton megalocarpus	at 40°C: 867.2, 870.4 [22] [27] at 15°C: 883 [245]	4.05, 4.78 [22] [245]	37.24, 39.53 [245] [22]	39.786 [27]	...	0.16 & 0.2 [26]	164, 192 [27] [245]	46.6, 47.52 [26] [245]	0.71, 2.88 [27] [245]	-6, -3 [245] [27]	-3, -2 [22] [27]
Patchouli (Pogostemon cablin)	at 40°C: 922.1 [27]	6.0567 [27]	...	44.18 [27]	118.5 [27]	...	0.022 [27]	-33 [27]	-33 [27]
Sterculia foetida	at 40°C: 877.6 [27]	6.3717 [27]	...	40.001 [27]	130.5 [27]	...	1.46 [27]	1 [27]	2 [27]
greenseed	at 40°C: 877±1 [40]	20.95±0.2 [40]	...	0.162±0.021 [40]	1 to 3 [40]	-12 to -10 [40]
Aphanamixis polystachya	at 40°C: 873.5 [26]	4.7177 [26]	39.960 [26]	0.448 [26]	188.5 [26]	...	0.16 [26]	8 [26]	8 [26]

Honesty (<i>Lunaria annua</i>)	at 15°C: 877±2 [223]	6.815±0.034 [223]	0.42±0.02 [223]	78.9±0.8 [223]
Mango	at 40°C: 873, 882 [224] [225]	4.3, 4.73 [224] [225]	40.453 [225]	...	0.78 [224]	54 [225]
Neem (<i>Azadirachta indica</i>)	at 40°C: 868, 895.2±0.59 [26] [226]	3.58, 5.53 [227] [228]	39.810 [26]	...	0.42, 0.649 [227] [26]	...	76 to 120, 175 [26] [227]	48 to 53, 57 [26] [229]	7.1 [26]	9 to 14.4, 6 [26] [228]	2, 10 [26] [227]
Passion fruit (<i>Passiflora edulis</i>)	...	4.1 [138]	172 [138]	...	3.1 [138]
Rubber seed (<i>Hevea brasiliensis</i>)	at 15°C: 877.3 [230]	5.77 [230]	...	40.3 [230]	0.448 [230]	37.15 [230]	141 [230]	52.56 [230]	8 [230]	9 [230]	7 [230]
Sea buckthorn (<i>Hippophae rhamnoides</i>)	at 20°C: 886.5 [135]	3.79, [135]	39.64 [135]	40.9 [135]
Tall	at 15°C: 878 to 885, 922 [231] [232]	4.1 to 5.3, 7.1 [231] [232]	40.023 [232]	89 [232]	54 [232]	...	1 [232]	...
Tamanu (<i>Polanga</i> , <i>Foraha</i> , <i>Calophyllum tacamahaca</i>)	at 40°C: 869, 894 & 892 & 893 [197] [233] at 15°C: 868.7, 873 [56] [234] at 32°C: 905 [235]	5.74, 3.99 [22] [197]	38.33, 41.397 [234] [197]	39.38, 41.5 [56] [235]	0.3, 0.88 & 1 & 0.76 [120] [233]	...	93.5, 162.5 [22] [27]	56.53 & 58.42 & 58.39, 50 [233] [235]	3.58, 9.42 [56] [22]	7, 13.2 [56] [234]	4.3, 13 [234] [27]
Tucumã butter (<i>Astrocaryum vulgare</i>)	at 40°C: 877 [236]	3.7, 4.54 [237] [236]	0.23 [237]	...	124 [237]
sapindus mukorossi (soapnut)	at 15°C: 875, 876 [238] [239] at 20°C: 870 [238]	4.63 [239]	40.02 [239]	...	0.14 [239]	...	140 [239]	56 [239]	...	-1 [239]	-4 [239]

Stillingia (Chinese vegetable tallow, Sapium sebiferum)	at 15°C: 892, 900 [209] [210]	3.698, 4.81 [209] [210]	0.007, 0.15 [211] [209]	137, 180 [211] [209],	40.2, 50 [209] [210]	0.6, 0.8 [211] [209]	-13 [211]	...
Artichoke	at 25°C: 880 [212] at 15°C: 889 [213]	3.56, 5.101 [212] [213]	...	39.8 [212]	175, 182 [212] [213]	59 [213]	...	-4, -1 [213] [212]	...
Astrocaryum murumuru butter	at 20°C: 877.9 ± 1.6 [172]	3.1 ± 0.03 [214]	0.6±0.1 [214]	40 [172]
Balanos (Balanites aegyptiaca)	at 40°C: 860, 874.8 [215] [216]	3.98, 4.46 [215] [216]	42.5 [216]	39.65 [215]	...	0.34, 1.26 [215] [216]	75, 160 [215] [216]	42 [216]	-2.5, 0 [215] [216]
Brucea javanica	at 40°C: 871 [217]	3.556 [217]	0.027 [217]	164 [217]	...	3 [217]	2 [217]	1 [217]
Buriti (Mauritia flexuosa)	at 40°C: 877.3 ± 6 [172]	5.22 [218]	190 [218]	...	16.5 [218]
Candlenut (Kukui nut)	at 40°C: 886.9 [219] at 20°C: 885.7 [220]	4.8, 4.819 [220] [219]	40.33 [220]	0.4 [220]	160, 161 [219] [220]	...	5.9 [220]	6 [220]	6.667, 6.84 [219] [220]
Chaulmoogra (Hydnocarpus wightiana)	at 40°C: 893 [221]	5.4 [221]	40.7 [221]	163 [221]	2 [221]
Crambe (Crambe abyssinica)	at 40°C: 848 [32]	5.1, 5.12 [32] [36]	...	41.98, 42.26 [36] [32]	190 [36]
Croton (tigilium, Croton tigilium)	at 40°C: 865 [16]	4.78 [16]	...	39.95 [16]	46.6 [16]
Cuphea	...	2.38 & 2.4 [222]	56.07 & 55.06 [222]	3.09 & 3.57 [222]	-9.1 & -10.1 [222]	-21.5 & -22.5 [222]
Cupuaçu butter	...	4.8 [138]	176 [138]	...	72.7 [138]

A wide range of second-generation oils, all of which are non-edible, are reported in the table. Table 2 shows the second generation oils that have been the most researched. The limitations of second generation oils compared to first generation oils are quite evident and 39 oils have been studied to date.

Table 2: Properties of the second generations of biodiesel.

Biodiesels	Physicochemical property ranges										
	Density (kg/m ³)	Kin. Viscosity 40°C (mm ² /s)	Calorific Value (Mj/Kg)	Higher (gross) Heating Value (Mj/Kg)	Lower (net) Heating Value (Mj/Kg)	Acid Value (Neutralization number) (mg KOH/g)	Flash Point (°C)	Cetane Number	Oxidation Stability (h)	Cloud Point (°C)	Pour Point (°C)
Tung	at 15°C: 883, 903 [195] [196]	4.64, 7.84 [195] [196]	38.8 [195]	0.124 [196]	197 [195]	39 [196]	0.3 [196]
Jatropha	at 40°C: 864.2, 873 [27] [197] at 20°C: 784, 875.9 [198] [199] at 15°C: 865.7, 880 [56] [104]	2.35 to 2.47, 6.1 to 6.7 [34] [200]	39.56, 42.673 [200] [152]	39.738, 39.84 [27] [16]	...	0.05, 1.05 [56] [104]	108, 194 [199] [124]	49, 60.74 to 63.27 [26] [34]	3.02, 13.51 [56] [199]	1, 10.2 [104] [197]	-6 to 2, 10 [34] [27]
Jojoba (Simmondsia chinensis)	at 40°C: 830 [201]	2.2, 5.86 [201] [202]	35.66 [202]	0.22 to 0.45 [202]	100, 150 [201] [202]	7 [201]	5 [201]
Nahor (Mesua ferrea)	at 40°C: 873 [203]	4.1, 5.525 [204] [203]	35 [203]	1.8 [203]	113, 142 [203] [204]	6.1 [204]	-1.2 [204]
Paradise (Simarouba glauca)	at 40°C: 875.2 [205] at 15°C: 865 [206]	4.68, 5.4 [206] [205]	0.1, 0.7 [205] [207]	141.2, 165 [205] [206]	64 [205]	...	19 [206]	14.5 [206]
Pongamia (Karanja, Honge, Milletia pinnata)	at 15°C: 880, 890 [208] [104] at 40°C: 883 [197]	3.99, 5.52 to 5.79 [208] [34]	37.8 to 39.69, 42.133 [34] [197]	0.1, 0.72 to 0.76 [93] [34]	141, 163 [93] [197]	55.1, 59.68 to 60.9 [93] [34]	2.35, 4.5 [93] [104]	-1, 14.6 [104] [197]	-3, 5.1 [104] [197]

Mutton fat	at 40°C: 856 [270]	0.65 [270]	59 [270]	-4 [270]	-5 [270]
Poultry fat	...	4.32, 4.71 [271] [278]	0.298 [279]	0.52 [271]	7 [278]	...	1, 3 [278] [280]
Waste cooking oil	at 15°C: 874.9, 880 [281] [282] at 40°C: 863.7, 890 [283] [284]	2.72, 6.1 [284] [285]	37.2, 37.9 [286] [285]	35.401, 41.2 [284] [282]	...	0.6, 0.71 [286] [283]	134, 186 [281] [284]	51.48, 64.2 [286] [281]	...	15.9 [281]	10.5 [286]	...	-1, 1 [282] [286]
Waste fat oil	10.91 [287]
Waste fried oil	at 25°C: 855 [288] at 15°C: 888 [289]	4.318, 4.57 [289] [288]	39.55, 40.5 [289] [288]	1.31±0.06 [287]	126, 156 [288] [289]	52 [289]	-8.3, 3 [288] [289]	...	-2.5 [289]
Waste frying palm oil	at 15°C: 875 [290]	4.401 [290]	38.73 [290]	0.15 [290]	70.6 [290]	60.4 [290]
Waste mixed vegetable oil	at 15°C: 878.9 [280]	4.83 [280]	59.7 [280]	...	14.12 [280]
Waste sunflower oil	at 15°C: 887.5 [280]	4.42 [280]	51.5 [280]	...	0.43 [280]
Trout oil	at 15°C: 885 [291]	4.25 [291]	37.8 [291]	51.3 [291]
Larvae grease (housefly)	at 15°C: 881 [292]	5.64 [292]	0.63 [292]	145 [292]	52 [292]
Plastic pyrolysis oil	at 15°C: 981 [293]	1.918 [293]	38.3 [293]	41 [293]	13 [293]
Sludge pyrolysis oil	at 22°C: 890 [294]	8.2 [294]	...	39.29 [294]	36.49 [294]	0.489 [294]	170 [294]
Neem seed pyrolysis oil	at 40°C: 982 [295]	9.38 [295]	...	20.8 [295]

Schizochytrium mangrovei	...	5.22 [259]	7.59 [259]	186.53 [259]	68.80 [259]	0.05 [259]	19.12 [259]	...
Spirulina	at 40°C: 860 [258]	5.66 [258]	41.36 [258]	0.45 [258]	130 [258]	-18 [258]
Spirulina platensis	at 15°C: 863.7 [260]	12.4 [260]	45.63 [260]	0.75 [260]	189 [260]	70 [260]	...	-3 [260]	-9 [260]
Animal fat	at 15°C: 871, 877 [261] [262]	4.03, 4.42 [261, 262]	...	36.83, 89.49 [261] [262]	36.73 [263]	0.38 [263]	161.5 [261]	57.49, 65.6 [261] [262]	13.03 [263]
Animal fat traps	at 15°C: 870 [264]	4.7 [264]	...	37 [264]	...	0.3 [264]	128 [264]	53 [264]
Beef tallow	at 17°C: 877 [265] at 20°C: 832 266] at 40°C: 0.890 [267]	4.89, 5.47±0.005 [266] [268]	39.858 [265]	...	39.931 [267]	0.495±0.007 [268]	152, 210 [266] [268]	58.8, 64.8 [267] [267]	1.99 [268]	...	0 [265]
Camelus dromedaries fat (Camel fat)	at 15°C: 871 [269]	3.39 [269]	39.52 [269]	...	158 [269]	58.7 [269]	...	12.7 [269]	15.5 [269]
Chicken fat	at 40°C: 867, 881.6 [270] [267] at 15°C: 869, 889.7 [271] [272]	5.3 [272]	39.911 [267]	0.25, 0.43 [270] [272]	169 [272]	57.61 [267] [270]	...	-5 [270]	-6 [270]
Fish oil	at 40°C: 880,885 [273] [274]	4, 4.96 [273] [275]	37.8, 42.241 [275] [273]	114, 176 [274] [273]	51, 52.4 [275] [276]	...	-5 [275]	-14, 4 [275, 276]
Fleshing oil	at 15°C: 876.7 [272]	4.7 [272]	0.28 [272]	168 [272]	58.8 [272]
inedible animal tallow	at 17°C: 877 [265]	5.072 [265]	39.858 [265]
Lard	at 40°C: 879.5 [267]	4.64 to 7.73 [277]	39.932 [267]	1.13 [277]	...	57.8 [267]

The third generation of biofuels, which has done less to date than the previous two generations. The values are reported in Table 3. 33 oils in the third generation have been studied to date and the incidence rate is lower than the previous two generations due to the novelty.

Table 3: Properties of the third generations of biodiesel.

Biodiesels obtained from	Physicochemical property ranges										
	Density (kg/m ³)	Kin. Viscosity 40°C (mm ² /s)	Calorific Value (Mj/Kg)	Higher (gross) Heating Value (Mj/Kg)	Lower (net) Heating Value (Mj/Kg)	Acid Value (Neutralization number) (mg KOH/g)	Flash Point (°C)	Cetane Number	Oxidation Stability (h)	Cloud Point (°C)	Pour Point (°C)
Ankistrodesmus braunii and Nannochloropsis	at 40°C: 869 [246]	4.19 [246]	40.72 [246]	144 [246]	...	7 [246]	-6 [246]	
Auxenochorella protothecoides	at 15°C: 876.9 [247]	4.354 [247]	0.2 [247]	...	1.2 [247]	
Chlorella protothecoides	at 15°C: 882 [248] at 40°C: 864 [249]	4.41, 5.2 [249] [250]	39.01, 41 [249, 250]	0.29, 0.374 [248] [249]	115 [249]	4.52 [248]	
Chlorella variabilis	at 15°C: 867 [251]	4.85 [251]	38.78 [251]	157 [251]	
Chlorella vulgaris	at 40°C: 860, 895 [252] [253]	4.1, 5.2 [253] [252]	42.7 [253]	0.51 [253]	115 [252]	-7 [253]	
Euglena sanguinea	at 15°C: 861, 868 [254] [255]	4.483, 4.545 [254, 255]	0.29, 0.32 [255] [254]	169, 172 [254] [255]	6.20 [255]	15 [255]	13 [255]	
Heterotrophic microalgae (Sugar plant)	at 15°C: 778 [256]	2.748 [256]	44 [256]	
Melanothamnus atafhusainii	at 40°C: 870 [257]	3.67 [257]	0.75 [257]	-1 [257]	-2 [257]	
Pond water algae	at 40°C: 872 [258]	5.82 [258]	40.80 [258]	0.40 [258]	-16 [258]	

2. Density

Density is one of the most important factors in biodiesels. Most studies have reported temperatures of density between 15 and 40 °C for all three generations because the temperature has a direct effect on density. Also, the free fatty acid content, molar mass, temperature, the water content can effect on the density of esters. The cetane number, viscosity, heating value, fuel performance, and the quality of combustion and atomization are strongly connected to the density. The density of diesel fuel is lower than biodiesels. The unit quantity of all reports is converted to kg/m³.

The maximum density of the first generation of biodiesel is reported for Castor biodiesel (at 20°C: around 917). Peanut sample also showed the highest density of about 992 and 884 at 40 and 15°C. The minimum density of the biodiesel was stated for the Watermelon seed around 800 and 880.6 at 15 and 20°C.

Patchouli and Tall biodiesels (second generation of biodiesel) could show the highest amount of density at 15 and 40°C around 922 and 922.1. The minimum of this generation was shown by the jojoba and jatropha at 40, 20 and 15°C (830, 865.7, and 874).

Plastic pyrolysis and neem seed pyrolysis were shown the highest amount of density at 15 and 40°C around 981 and 982. The majority of reports were done at 15 and 40°C, however, some reports were performed at 22, 17, and 25°C (Table 3). The lowest of the density for the third generation of biodiesels were shown by the heterotrophic microalgae at 15°C (778).

Comparison between three generations of biodiesel shows that the highest of density was reported by Peanut biodiesel (first generation) and it was higher than second and third generation of biodiesel around 8 and 2 % and heterotrophic microalgae (third-generation) had the lowest density compared to other generation to approximately 3 and 8%.

2.1. Viscosity

The viscosity plays a leading role in the engine performance of biodiesels. It can affect the size of the particles, spray quality, starting the engine, the quality of the fuel-air mixture combustion, and penetration of the injected jet. Also, the viscosity can affect the lubricity. The amount viscosity has a limitation due to several reasons. The high viscosity makes the formation of too big drops, the increase of combustion chamber deposits, the increase of needed fuel pumping energy and wear of the pump and the injector elements. Also, the high viscosity causes operational issues at the low temperatures due to that the viscosity enhances with reducing the temperature. The low viscosity makes the inadequate penetration and the formation of the black smoke specific to combustion (during the absence of oxygen). Biodiesel is more polar compared to the diesel fuel, so, the viscosity of biodiesel is greater than diesel

fuel. Table 1, 2, and 3 show the viscosity of different feedstocks. The viscosity was measured at 40°C for three generations of biodiesel.

The unit quantity of all reports is converted to mm²/s.

The maximum viscosity was reported for castor biodiesel around 14.4 for the first generation of biodiesel. Orange and watermelon seed biodiesels also showed the lowest amount of viscosity to roughly 1.04 and 1.05.

The highest amount of viscosity for the second generation of biodiesel was shown by tung. It could show the viscosity around 7.84. The minimum of the viscosity was displayed by the jatropha and jojoba biodiesel. It was only around 2.35 and 2.2.

Spirulina platensis and neem seed pyrolysis were shown the highest amount of viscosity for the third generation of biodiesel. They were around 12.4 and 9.38. On the other hand, the lowest amount of viscosity was reported for plastic pyrolysis and it was about 1.91.

The comparison between all generations could show that the maximum viscosity was shown by the castor and it was higher than the second and third generation of biodiesels to approximately 46 and 14%. The minimum of viscosity also reported by the first generation of biodiesels and it was 52 and 45% lower than the second and third generation of biodiesels.

2.2. Calorific Value

The heating value or calorific value of the fuel is defined as the amount of energy released through the combustion of the unit value of the fuel. The unit quantity of all reports is converted to Mj/Kg. The upper heating value is gained while all products of the combustion are cooled down to the temperature before the water vapor combustion formed over combustion is condensed. The lower heating value is achieved by subtracting the latent heat of vaporization of the water vapor formed with the combustion from the upper heating value. Some of the reports indicated only to the calorific value and others referred to higher and lower heating values.

The highest calorific value for the first generation was gained by the false flax biodiesel. It was from 45.05 to 46.15. The maximum higher heating value was shown by the cottonseed and it was about 48.18. Also, the greatest amount of the lower heating values reported by the coconut biodiesel (57.74). On the other hand, the minimum of calorific value was displayed by the thistle biodiesel (36.5). The lowest amount of the higher and lower heating values were gained by the canola and cottonseed biodiesels (25.11±0.29 and 36.89).

Nerium oleander had the highest amount of calorific value between all second generations of the biodiesel (44.98) and the lowest heating value was shown by the nahor biodiesel [35].

The higher heating value of the patchouli was greater than other second generations (44.18). The lowest higher heating value was shown by the green seed biodiesel and it was only around 20.95 ± 0.2 .

Spirulina platensis showed the maximum of calorific value between all third generations of biodiesel (45.63). The lowest calorific value was gained by waste cooking and it was around 37.2. Lard biodiesel showed the highest higher heating values around 39.93 and neem seed pyrolysis was gained 20.8. The greatest lower heating value of the third generation of biodiesels was gained by the heterotrophic microalgae. It was around 44. The minimum lower heating value was reported sludge pyrolysis biodiesel (36.49).

Comparison between all biodiesel generations could display that the *spirulina platensis* could achieve the maximum of calorific value and it was higher than nerium oleander and false flax to approximately 3 and 2%. Also, the highest higher heating value of the first generation of biodiesel (cottonseed) was higher than the second and third generation of biodiesel around 9 and 18%.

2.3. Acid Value

The acidic value (acid number or neutralization number) in chemistry is the amount of mg of potassium hydroxide needed to neutralize one gram of a substance. An acidic number is a measure of the number of carboxylic acid groups in a compound, such as a fatty acid or a mixture of compounds. The upper amount of free fatty acid contributes to the elevated acid value which in turn causes severe corrosion in fuel supply lines of the engine. Besides, the acid value can be observed as the indication of the level of lubrication in fuel lines. The unit quantity of all reports is mg KOH/g.

The castor biodiesel showed the highest acid value between all biodiesels of the first generation (3.9). The lowest acid value was displayed by the dika biodiesel and it was merely around 0.01.

The maximum of acid value for the second generation of biodiesel was gained by the nahor (1.8) and the minimum of the acid value was shown by the stillingia (0.007).

Plastic pyrolysis biodiesel from the third generation could display the highest amount of the acid value (41) and the lowest of acid value was gained by the waste frying palm (0.15).

The third generation of the biodiesel could show the much higher acid value compared to other generations and the second generation of biodiesel could gain the minimum of the acid value compared to others.

2.4. Flash Point

Flashpoint is the smallest temperature at which the fuel will ignite on the application of the ignition source under particular situations. Every liquid has a vapor pressure which is a function of its temperature. As the temperature rises, the vapor pressure increases. As the vapor pressure increases, the density of flammable liquid vapor increases. Therefore, temperature determines the amount of combustible liquid vapor in the air. Flashpoint measurement is done in two main ways: open cup and closed cup. The diesel fuel has a flashpoint around 50-65°C. Mostly, the flashpoint of the biodiesel is much higher than diesel fuel. The high flash point of biodiesel increases the security of fuel storage and transportation. The unit quantity of all reports is Celsius (centigrade).

The palm biodiesel had the maximum of the flashpoint between all first generation of biodiesel (214.5°C). On the other hand, the minimum of the flashpoint was gained by the orange biodiesel (29°C) and it was lower than diesel fuel.

Tung biodiesel (second generation of biodiesel) had the highest amount of the flashpoint around 197°C. However, the flashpoint of the nerium oleander was merely about 73°C.

In the third generation of biodiesel, the beef tallow had the greatest flashpoint between all samples (around 210°C) while the plastic pyrolysis biodiesel was only around 13°C.

The flashpoint of palm biodiesel was higher than tung and beef tallow around 8 and 3% and the plastic pyrolysis biodiesel was lower than the nerium oleander and orange biodiesel around 82 and 55%

2.5. Cetane Number

The cetane number represents the delay between the start of the injection into the combustion chamber and the start of the fuel combustion. During this delay, the fuel accumulates and then ignites and the combustion explodes to produce a powerful effect. Reducing the delay time makes the combustion more uniform. The increase of the cetane number causes the quick ignition of the fuel and it makes less non-ignited fuels building up in the combustion chamber and also further complete fuel combustion. The low cetane number affect the incomplete combustion and it causes the enhancement of the exhaust emissions and extreme deposits in the engine. Normally, biodiesels have a higher cetane number due to greater oxygen content compared to the diesel fuel.

Papaya seed biodiesel (first generation) showed the highest amount of the cetane number compared to other biodiesels. The value of the cetane number was around 77.3. On the contrary, pomegranate seed biodiesel had the lowest cetane number around (26.1).

Honesty biodiesel (second generation) displayed the highest amount of cetane number around 78.9±0.8 compared to other biodiesels in this generation, while tung biodiesel could

reach only around 39.

Heterotrophic microalgae were shown the maximum of the cetane number in the third generation of biodiesel (almost 75). However, the cetane number of the fish biodiesel was around 51.

The highest cetane number between all generations was shown by the honesty biodiesel and it was 3 and 6 % higher than the maximum of the other generations. Pomegranate seed biodiesel was shown the lowest amount of cetane number compared to other generations and it was 33 and 48% than the minimum of the other generations.

2.6. Oxidation Stability

The oxidation can affect the quality of the biodiesel over storage in contact with air. The storage stability is extremely important for the biodiesel and it indicates the ability of the fuel to stand chemical changes over the long term storage due to the connection with the oxygen from the air. The oxidation stability of biodiesel is subject to the number of bis-allylic sites in unsaturated compounds. The primitive oxidation is started by the radical formation at bis-allylic sites and it forms peroxides. Then, the secondary oxidation generates the aldehydes, volatile organic compounds, and ketones with ruing the methyl ester which polymerizes to form waste sludge that can detriment the engine fuel injection system.

This feature is not mentioned in all reports but Ben biodiesel had the highest in first-generation biodiesel (26.2 h). On the contrary, the kenaf seed biodiesel had the lowest oxidation stability and it was around 0.35 h.

The honesty biodiesel showed the maximum of the oxidation stability between all second generations of biodiesel (72 h). *Sterculia foetida* biodiesel displayed the minimum of the oxidation stability around 0.022 h.

The waste mixed vegetable biodiesel presented the greatest amount of oxidation stability in the third generation of biodiesel about 14.12 h. However, the lowest of the oxidation stability was shown by the waste sunflower biodiesel (around 0.43 h).

Comparing between all generations, the honesty biodiesel was higher than the maximum of the first and third generation of biodiesel around 63 and 80%. Also, *Sterculia foetida* biodiesel (second generation of biodiesel) was lower than the minimum of the first and third generation of biodiesel to approximately 93 and 94%.

2.7. Cloud and Pour Point

The minimum temperature at which a cloud of paraffin crystals appears inside the oil product is called the cloud point. At this temperature, the sample does not lose its fluidity and

is usable. The pour point of a hydrocarbon material is when it cools under certain conditions and is defined as the lowest temperature at which the hydrocarbon flows. This temperature is somewhat higher than the solidification point temperature. It is difficult to define precisely the pour or solidification point since the transition from the liquid phase to the solid phase is gradual. The unit quantity of all reports is Celsius (centigrade).

Ben biodiesel showed the highest cloud and pour point between all first generations of biodiesel (19°C). However, pumpkin seed had the minimum of cloud point around -18°C. Also, it had the lowest pour point between all biodiesel samples around -32°C.

The maximum of cloud and pour point was shown by the paradise biodiesel between the second generation of biodiesel (19 and 14.5°C). On the other hand, the minimum of cloud and pour point was reported by the patchouli biodiesel (-33°C).

Euglena sanguinea biodiesel had the greatest cloud and pour point between all third generations of biodiesel and it was around 15 and 13°C. Waste fried oil and *Spirulina* showed the lowest cloud and pour point to roughly -8.3 and -18°C.

Paradise and ben biodiesel had the highest cloud point and it was 21% higher than *Euglena sanguinea* biodiesel. However, the ben biodiesel had the maximum pour point compared to the all second and third generation of biodiesel. It was 23 and 31% higher than the maximum of other generations. The patchouli biodiesel indicated the minimum of the cloud and pour point between all generations and it was lower than the lowest cloud and pour point of other generations between 4-74%.

3. Conclusion and Future Trend

This review article attempts to provide comprehensive information on the physical properties of the majority of biodiesel used in all three generations and propose the best biodiesel concerning their physical properties. Biodiesel has many advantages over fossil fuels. One of the most important reasons for choosing biodiesel is its impact on the economy, environment and energy security in the world. Some of the most important benefits of biodiesel on the economy can be sustainability, job opportunities in the rural area, fuel diversity, more income taxes, development of agriculture, International competitiveness, decreasing the dependency on the imported petroleum, and improving investments in equipment and plant. Reducing air contamination, Biodegradability, Greenhouse gas reductions, better combustion efficiency, and carbon sequestration are some of the environmental impacts of biodiesel. One of the most important impacts of biodiesel on energy security can also be addressed renewability, ready availability, domestic distribution, supply reliability, domestic targets, and decreasing use of fossil fuels [296].

Although biodiesel has superior properties over fossil fuels, choosing the right biodiesel has many difficulties. Choosing the right biodiesel depends on various factors including standards set in different countries, raw material production policies, weather conditions, engine biodiesel performance, initial production costs, and physical properties of biodiesel available in that region and so on. Therefore, choosing the best option among all the studied biofuels is almost impossible, and choosing the best feedstock has to take into account all the physical, chemical and product conditions, and so on.

4. Acknowledgments

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

5. Conflict Of Interest Statement

The authors have confirmed that there is no conflict of interest.

6. References

1. Armas, O., J.J. Hernández, and M.D. Cárdenas, Reduction of diesel smoke opacity from vegetable oil methyl esters during transient operation. *Fuel*, 2006. 85(17-18): p. 2427-2438.
2. Atadashi, I., M. Aroua, and A.A. Aziz, High quality biodiesel and its diesel engine application: a review. *Renewable and sustainable energy reviews*, 2010. 14(7): p. 1999-2008.
3. Baskar, G., et al., Microalgae A Source for Third-Generation Biofuels. *Bioprocess Engineering for a Green Environment*, 2018: p. 297-306.
4. Klass, D.L., *Biomass for renewable energy, fuels, and chemicals*. 1998: Elsevier.
5. Ahmad, A., et al., Microalgae as a sustainable energy source for biodiesel production: a review. *Renewable and Sustainable Energy Reviews*, 2011. 15(1): p. 584-593.
6. Zhu, L., C. Cheung, and Z. Huang, Impact of chemical structure of individual fatty acid esters on combustion and emission characteristics of diesel engine. *Energy*, 2016. 107: p. 305-320.
7. Knothe, G., J. Krahl, and J. Van Gerpen, *The biodiesel handbook*. 2015: Elsevier.
8. Knothe, G., C.A. Sharp, and T.W. Ryan, Exhaust emissions of biodiesel, petrodiesel, neat methyl esters, and alkanes in a new technology engine. *Energy & Fuels*, 2006. 20(1): p. 403-408.
9. Ranganathan, S.V., S.L. Narasimhan, and K. Muthukumar, An overview of enzymatic production of biodiesel. *Bioresource technology*, 2008. 99(10): p. 3975-3981.
10. Barabás, I. and I.-A. Todoruț, Biodiesel quality, standards and properties. *Biodiesel-quality, emissions and by-products*, 2011: p. 3-28.
11. Casanave, D., J.-L. Duplan, and E. Freund, Diesel fuels from biomass. *Pure and Applied Chemistry*, 2007. 79(11): p. 2071-2081.
12. Erdmann, M., M. Böhning, and U. Niebergall, Physical and chemical effects of biodiesel storage on high-density polyethylene: Evidence of co-oxidation. *Polymer Degradation and Stability*, 2019. 161: p. 139-149.

13. Hoekman, S.K., et al., Review of biodiesel composition, properties, and specifications. *Renewable and sustainable energy reviews*, 2012. 16(1): p. 143-169.
14. Naik, S.N., et al., Production of first and second generation biofuels: a comprehensive review. *Renewable and sustainable energy reviews*, 2010. 14(2): p. 578-597.
15. Ramírez-Verduzco, L.F., J.E. Rodríguez-Rodríguez, and A. del Rayo Jaramillo-Jacob, Predicting cetane number, kinematic viscosity, density and higher heating value of biodiesel from its fatty acid methyl ester composition. *Fuel*, 2012. 91(1): p. 102-111.
16. Lujaji, F., et al., Cetane number and thermal properties of vegetable oil, biodiesel, 1-butanol and diesel blends. *Journal of Thermal Analysis and Calorimetry*, 2010. 102(3): p. 1175-1181.
17. Khot, M., et al., Fungal production of single cell oil using untreated copra cake and evaluation of its fuel properties for biodiesel. *J Microbiol Biotechnol*, 2015. 25(4): p. 459-63.
18. Kumar, G., et al., Continuous Low Cost Transesterification Process for the Production of Coconut Biodiesel. *Energies*, 2010. 3(1): p. 43-56.
19. Lafont, J.J., A.A. Espitia, and J.R. Sodr , Potential vegetable sources for biodiesel production: cashew, coconut and cotton. *Materials for Renewable and Sustainable Energy*, 2015. 4(1).
20. Duncan, A.M., et al., High-Pressure Viscosity of Biodiesel from Soybean, Canola, and Coconut Oils. *Energy & Fuels*, 2010. 24(10): p. 5708-5716.
21. Habibullah, M., et al., Biodiesel production and performance evaluation of coconut, palm and their combined blend with diesel in a single-cylinder diesel engine. *Energy Conversion and Management*, 2014. 87: p. 250-257.
22. Atabani, A.E., et al., Effect of Croton megalocarpus, Calophyllum inophyllum, Moringa oleifera, palm and coconut biodiesel–diesel blending on their physico-chemical properties. *Industrial Crops and Products*, 2014. 60: p. 130-137.
23. Benjapornkulaphong, S., C. Ngamcharussrivichai, and K. Bunyakiat, Al₂O₃-supported alkali and alkali earth metal oxides for transesterification of palm kernel oil and coconut oil. *Chemical Engineering Journal*, 2009. 145(3): p. 468-474.
24. Kalam, M.A., M. Husnawan, and H.H. Masjuki, Exhaust emission and combustion evaluation of coconut oil-powered indirect injection diesel engine. *Renewable Energy*, 2003. 28(15): p. 2405-2415.
25. Pinzi, S., et al., Multiple response optimization of vegetable oils fatty acid composition to improve biodiesel physical properties. *Bioresour Technol*, 2011. 102(15): p. 7280-8.
26. Wakil, M.A., et al., Influence of biodiesel blending on physicochemical properties and importance of mathematical model for predicting the properties of biodiesel blend. *Energy Conversion and Management*, 2015. 94: p. 51-67.
27. Atabani, A.E., et al., A comparative evaluation of physical and chemical properties of biodiesel synthesized from edible and non-edible oils and study on the effect of biodiesel blending. *Energy*, 2013. 58: p. 296-304.
28. Aydin, F., et al., The Basic Properties of Transesterified Corn Oil and Biodiesel-Diesel Blends. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, 2011. 33(8): p. 745-751.
29. Tesfa, B., et al., Prediction models for density and viscosity of biodiesel and their effects on fuel supply system in CI engines. *Renewable Energy*, 2010. 35(12): p. 2752-2760.
30. Serqueira, D.S., et al., Influence of blending soybean, sunflower, colza, corn, cottonseed, and residual cooking oil methyl biodiesels on the oxidation stability. *Fuel*, 2014. 118: p. 16-20.
31. Hazar, H. and U. Ozturk, The effects of Al₂O₃–TiO₂ coating in a diesel engine on performance and emission of corn oil methyl ester. *Renewable Energy*, 2010. 35(10): p. 2211-2216.

32. Demirbas, A., Prediction of Higher Heating Values for Biodiesels from Their Physical Properties. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, 2009. 31(8): p. 633-638.
33. Shehata, M.S., A.M.A. Attia, and S.M. Abdel Razek, Corn and soybean biodiesel blends as alternative fuels for diesel engine at different injection pressures. *Fuel*, 2015. 161: p. 49-58.
34. Patil, P.D. and S. Deng, Optimization of biodiesel production from edible and non-edible vegetable oils. *Fuel*, 2009. 88(7): p. 1302-1306.
35. Gülüm, M. and A. Bilgin, Density, flash point and heating value variations of corn oil biodiesel–diesel fuel blends. *Fuel Processing Technology*, 2015. 134: p. 456-464.
36. Demirbas, A., Relationships derived from physical properties of vegetable oil and biodiesel fuels. *Fuel*, 2008. 87(8-9): p. 1743-1748.
37. Prestes, R.A., et al., A rapid and automated low resolution NMR method to analyze oil quality in intact oilseeds. *Anal Chim Acta*, 2007. 596(2): p. 325-9.
38. Ramos, M.J., et al., Influence of fatty acid composition of raw materials on biodiesel properties. *Bioresour Technol*, 2009. 100(1): p. 261-8.
39. Ahmed, M., et al., Effect of biodiesel fuel properties and its blends on atomization. 2006, SAE Technical Paper.
40. Dmytryshyn, S.L., et al., Synthesis and characterization of vegetable oil derived esters: evaluation for their diesel additive properties. *Bioresource Technology*, 2004. 92(1): p. 55-64.
41. Eryilmaz, T., et al., Biodiesel production potential from oil seeds in Turkey. *Renewable and Sustainable Energy Reviews*, 2016. 58: p. 842-851.
42. Ozsezen, A.N., et al., Performance and combustion characteristics of a DI diesel engine fueled with waste palm oil and canola oil methyl esters. *Fuel*, 2009. 88(4): p. 629-636.
43. Albuquerque, M.C.G., et al., Properties of biodiesel oils formulated using different biomass sources and their blends. *Renewable Energy*, 2009. 34(3): p. 857-859.
44. Moser, B.R., Influence of blending canola, palm, soybean, and sunflower oil methyl esters on fuel properties of biodiesel. *Energy & fuels*, 2008. 22(6): p. 4301-4306.
45. Fernandes, D.M., et al., Preparation and characterization of methylic and ethylic biodiesel from cottonseed oil and effect of tert-butylhydroquinone on its oxidative stability. *Fuel*, 2012. 97: p. 658-661.
46. Nabi, M.N., M.M. Rahman, and M.S. Akhter, Biodiesel from cotton seed oil and its effect on engine performance and exhaust emissions. *Applied thermal engineering*, 2009. 29(11-12): p. 2265-2270.
47. Alhassan, Y., et al., Co-solvents transesterification of cotton seed oil into biodiesel: Effects of reaction conditions on quality of fatty acids methyl esters. *Energy Conversion and Management*, 2014. 84: p. 640-648.
48. Rashid, U., F. Anwar, and G. Knothe, Evaluation of biodiesel obtained from cottonseed oil. *Fuel Processing Technology*, 2009. 90(9): p. 1157-1163.
49. Demirbaş, A., Biodiesel from vegetable oils via transesterification in supercritical methanol. *Energy conversion and management*, 2002. 43(17): p. 2349-2356.
50. Karabektas, M., G. Ergen, and M. Hosoz, The effects of preheated cottonseed oil methyl ester on the performance and exhaust emissions of a diesel engine. *Applied Thermal Engineering*, 2008. 28(17-18): p. 2136-2143.
51. Moser, B.R., Fuel property enhancement of biodiesel fuels from common and alternative feedstocks via complementary blending. *Renewable Energy*, 2016. 85: p. 819-825.

52. Lingfeng, C., et al., Transesterification of cottonseed oil to biodiesel by using heterogeneous solid basic catalysts. *Energy & Fuels*, 2007. 21(6): p. 3740-3743.
53. Keera, S.T., S.M. El Sabagh, and A.R. Taman, Transesterification of vegetable oil to biodiesel fuel using alkaline catalyst. *Fuel*, 2011. 90(1): p. 42-47.
54. Okoro, L.N., et al., Thermodynamic and viscometric evaluation of biodiesel and blends from olive oil and cashew nut oil. *Research journal of chemical sciences*, 2011. 1(4): p. 90-97.
55. Rizwanul Fattah, I.M., et al., Effect of antioxidant on the performance and emission characteristics of a diesel engine fueled with palm biodiesel blends. *Energy Conversion and Management*, 2014. 79: p. 265-272.
56. Mofijur, M., et al., Assessment of Physical, Chemical, and Tribological Properties of Different Biodiesel Fuels, in *Clean Energy for Sustainable Development*. 2017. p. 441-463.
57. Salamanca, M., et al., Influence of palm oil biodiesel on the chemical and morphological characteristics of particulate matter emitted by a diesel engine. *Atmospheric Environment*, 2012. 62: p. 220-227.
58. Abu-Hamdeh, N.H. and K.A. Alnefaie, A comparative study of almond and palm oils as two bio-diesel fuels for diesel engine in terms of emissions and performance. *Fuel*, 2015. 150: p. 318-324.
59. Bunyakiat, K., et al., Continuous production of biodiesel via transesterification from vegetable oils in supercritical methanol. *Energy & Fuels*, 2006. 20(2): p. 812-817.
60. Kalam, M. and H. Masjuki, Biodiesel from palmoil—an analysis of its properties and potential. *Biomass and Bioenergy*, 2002. 23(6): p. 471-479.
61. Benjumea, P., J. Agudelo, and A. Agudelo, Effect of altitude and palm oil biodiesel fuelling on the performance and combustion characteristics of a HSDI diesel engine. *Fuel*, 2009. 88(4): p. 725-731.
62. Trakarnpruk, W. and S. Porntangjitlikit, Palm oil biodiesel synthesized with potassium loaded calcined hydrotalcite and effect of biodiesel blend on elastomer properties. *Renewable Energy*, 2008. 33(7): p. 1558-1563.
63. Mofijur, M., et al., Comparative evaluation of performance and emission characteristics of *Moringa oleifera* and Palm oil based biodiesel in a diesel engine. *Industrial Crops and Products*, 2014. 53: p. 78-84.
64. Ahmad, M., et al., Optimization of base catalyzed transesterification of peanut oil biodiesel. *African Journal of Biotechnology*, 2009. 8(3).
65. Kaya, C., et al., Methyl ester of peanut (*Arachis hypogea* L.) seed oil as a potential feedstock for biodiesel production. *Renewable Energy*, 2009. 34(5): p. 1257-1260.
66. Silveira Junior, E.G., et al., Potential of Virginia-type peanut (*Arachis hypogaea* L.) as feedstock for biodiesel production. *Industrial Crops and Products*, 2016. 89: p. 448-454.
67. Pinto, L.M., et al., Comparative evaluation of the effect of antioxidants added into peanut (*arachis hypogae* l.) oil biodiesel by P-DSC and rancimat. *Journal of Thermal Analysis and Calorimetry*, 2014. 120(1): p. 277-282.
68. Moser, B.R., Preparation of fatty acid methyl esters from hazelnut, high-oleic peanut and walnut oils and evaluation as biodiesel. *Fuel*, 2012. 92(1): p. 231-238.
69. Labeckas, G. and S. Slavinskas, The effect of rapeseed oil methyl ester on direct injection Diesel engine performance and exhaust emissions. *Energy Conversion and Management*, 2006. 47(13-14): p. 1954-1967.
70. Xin, J., H. Imahara, and S. Saka, Oxidation stability of biodiesel fuel as prepared by supercritical methanol. *Fuel*, 2008. 87(10-11): p. 1807-1813.
71. Kegl, B. and A. Hribernik, Experimental analysis of injection characteristics using biodiesel fuel. *Energy & fuels*,

2006. 20(5): p. 2239-2248.

72. Azcan, N. and A. Danisman, Microwave assisted transesterification of rapeseed oil. *Fuel*, 2008. 87(10-11): p. 1781-1788.

73. Tsolakis, A., et al., Engine performance and emissions of a diesel engine operating on diesel-RME (rapeseed methyl ester) blends with EGR (exhaust gas recirculation). *Energy*, 2007. 32(11): p. 2072-2080.

74. Dzida, M. and P. Prusakiewicz, The effect of temperature and pressure on the physicochemical properties of petroleum diesel oil and biodiesel fuel. *Fuel*, 2008. 87(10-11): p. 1941-1948.

75. Duz, M.Z., A. Saydut, and G. Ozturk, Alkali catalyzed transesterification of safflower seed oil assisted by microwave irradiation. *Fuel Processing Technology*, 2011. 92(3): p. 308-313.

76. Eryilmaz, T. and M.K. Yesilyurt, Influence of blending ratio on the physicochemical properties of safflower oil methyl ester-safflower oil, safflower oil methyl ester-diesel and safflower oil-diesel. *Renewable Energy*, 2016. 95: p. 233-247.

77. Çelebi, Y. and H. Aydın, Investigation of the effects of butanol addition on safflower biodiesel usage as fuel in a generator diesel engine. *Fuel*, 2018. 222: p. 385-393.

78. Işık, M.Z. and H. Aydın, Analysis of ethanol RCCI application with safflower biodiesel blends in a high load diesel power generator. *Fuel*, 2016. 184: p. 248-260.

79. Aydın, H., Scrutinizing the combustion, performance and emissions of safflower biodiesel–kerosene fueled diesel engine used as power source for a generator. *Energy Conversion and Management*, 2016. 117: p. 400-409.

80. İlkılıç, C., et al., Biodiesel from safflower oil and its application in a diesel engine. *Fuel Processing Technology*, 2011. 92(3): p. 356-362.

81. Al-Samarae, R., et al., Perspective of safflower (*Carthamus tinctorius*) as a potential biodiesel feedstock in Turkey: characterization, engine performance and emissions analyses of butanol–biodiesel–diesel blends. *Biofuels*, 2017: p. 1-17.

82. Rashid, U. and F. Anwar, Production of biodiesel through base-catalyzed transesterification of safflower oil using an optimized protocol. *Energy & Fuels*, 2008. 22(2): p. 1306-1312.

83. Karabas, H., Application of the Taguchi Method for the Optimization of Effective Parameters on the Safflower Seed Oil Methyl Ester Production. *International Journal of Green Energy*, 2014. 11(9): p. 1002-1012.

84. Özçelik, A.E. Determination of the Effects of Safflower Biodiesel and Its Blends with Diesel Fuel on Engine Performance and Emissions in a Single Cylinder Diesel Engine. in *International Conference on Software Technology and Engineering*, 3rd (ICSTE 2011). 2011. ASME Press.

85. Hamamci, C., et al., Biodiesel Production via Transesterification from Safflower (*Carthamus tinctorius*L.) Seed Oil. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, 2011. 33(6): p. 512-520.

86. Wakil, M.A., et al., Evaluation of rice bran, sesame and moringa oils as feasible sources of biodiesel and the effect of blending on their physicochemical properties. *RSC Adv.*, 2014. 4(100): p. 56984-56991.

87. Saydut, A., et al., Transesterified sesame (*Sesamum indicum* L.) seed oil as a biodiesel fuel. *Bioresour Technol*, 2008. 99(14): p. 6656-60.

88. Atapour, M. and H.-R. Kariminia, Optimization of Biodiesel Production from Iranian Bitter Almond Oil Using Statistical Approach. *Waste and Biomass Valorization*, 2013. 4(3): p. 467-474.

89. Rao, G.L.N., et al., Relationships among the physical properties of biodiesel and engine fuel system design requirement. *International journal of energy and environment*, 2010. 1(5): p. 919-926.

90. Valente, O.S., et al., Fuel consumption and emissions from a diesel power generator fuelled with castor oil and soybean biodiesel. *Fuel*, 2010. 89(12): p. 3637-3642.
91. Candeia, R.A., et al., Influence of soybean biodiesel content on basic properties of biodiesel–diesel blends. *Fuel*, 2009. 88(4): p. 738-743.
92. Moser, B.R. and S.F. Vaughn, Coriander seed oil methyl esters as biodiesel fuel: Unique fatty acid composition and excellent oxidative stability. *Biomass and Bioenergy*, 2010. 34(4): p. 550-558.
93. Sarin, R., et al., Jatropha–Palm biodiesel blends: An optimum mix for Asia. *Fuel*, 2007. 86(10-11): p. 1365-1371.
94. Vujicic, D., et al., Kinetics of biodiesel synthesis from sunflower oil over CaO heterogeneous catalyst. *Fuel*, 2010. 89(8): p. 2054-2061.
95. Porte, A.F., et al., Sunflower biodiesel production and application in family farms in Brazil. *Fuel*, 2010. 89(12): p. 3718-3724.
96. Saydut, A., et al., Process optimization for production of biodiesel from hazelnut oil, sunflower oil and their hybrid feedstock. *Fuel*, 2016. 183: p. 512-517.
97. Fadhil, A.B., A.M. Aziz, and M.H. Altamer, Potassium acetate supported on activated carbon for transesterification of new non-edible oil, bitter almond oil. *Fuel*, 2016. 170: p. 130-140.
98. Atapour, M. and H.-R. Kariminia, Characterization and transesterification of Iranian bitter almond oil for biodiesel production. *Applied Energy*, 2011. 88(7): p. 2377-2381.
99. Giwa, S.O., L.A. Chuah, and N.M. Adam, Fuel properties and rheological behavior of biodiesel from egusi (*Colocynthis citrullus* L.) seed kernel oil. *Fuel Processing Technology*, 2014. 122: p. 42-48.
100. Aburas, H. and A. Demirbas, Evaluation of beech for production of bio-char, bio-oil and gaseous materials. *Process Safety and Environmental Protection*, 2015. 94: p. 29-36.
101. Gonçalves, J.D., M. Aznar, and G.R. Santos, Liquid–liquid equilibrium data for systems containing Brazil nut biodiesel+methanol+glycerin at 303.15K and 323.15K. *Fuel*, 2014. 133: p. 292-298.
102. Latinwo, G.K., D.S. Aribike, and S.A. Kareem, Comparative study of biodiesels produced from unrefined vegetable oils. *Nature and Science*, 2010. 8(9): p. 102-106.
103. Vedharaj, S., et al., Experimental and finite element analysis of a coated diesel engine fueled by cashew nut shell liquid biodiesel. *Experimental Thermal and Fluid Science*, 2014. 53: p. 259-268.
104. Phoo, Z.W.M.M., et al., Physico-Chemical Properties of Biodiesel from Various Feedstocks, in *Zero-Carbon Energy Kyoto 2012*. 2013. p. 113-121.
105. Radhakrishnan, S., et al., Effect of nanoparticle on emission and performance characteristics of a diesel engine fueled with cashew nut shell biodiesel. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, 2018. 40(20): p. 2485-2493.
106. Devarajan, Y., D.B. Munuswamy, and B. Nagappan, Emissions analysis on diesel engine fuelled with cashew nut shell biodiesel and pentanol blends. *Environ Sci Pollut Res Int*, 2017. 24(14): p. 13136-13141.
107. Senthilkumar, G., et al., Evaluation of emission, performance and combustion characteristics of dual fuelled research diesel engine. *Environ Technol*, 2018: p. 1-8.
108. Bello, E., F. Out, and A. Osasona, Cetane number of three vegetable oils, their biodiesels and blends with diesel fuel. *Journal of Petroleum Technology and Alternative Fuels*, 2012. 3(5): p. 52-57.
109. Koçak, M.S., E. Ileri, and Z. Utlu, Experimental study of emission parameters of biodiesel fuels obtained from

canola, hazelnut, and waste cooking oils. *Energy & fuels*, 2007. 21(6): p. 3622-3626.

110. Çelikten, İ., E. Mutlu, and H. Solmaz, Variation of performance and emission characteristics of a diesel engine fueled with diesel, rapeseed oil and hazelnut oil methyl ester blends. *Renewable Energy*, 2012. 48: p. 122-126.

111. Gumus, M., Evaluation of hazelnut kernel oil of Turkish origin as alternative fuel in diesel engines. *Renewable Energy*, 2008. 33(11): p. 2448-2457.

112. Gülüm, M. and A. Bilgin, Measurements and empirical correlations in predicting biodiesel-diesel blends' viscosity and density. *Fuel*, 2017. 199: p. 567-577.

113. Xu, Y.X. and M.A. Hanna, Synthesis and characterization of hazelnut oil-based biodiesel. *Industrial Crops and Products*, 2009. 29(2-3): p. 473-479.

114. Rahman, M.M., et al., Effect of small proportion of butanol additive on the performance, emission, and combustion of Australian native first- and second-generation biodiesel in a diesel engine. *Environ Sci Pollut Res Int*, 2017. 24(28): p. 22402-22413.

115. Azad, A.K., et al., Biodiesel From Queensland Bush Nut (*Macadamia integrifolia*), in *Clean Energy for Sustainable Development*. 2017. p. 419-439.

116. Knothe, G., Biodiesel Derived from a Model Oil Enriched in Palmitoleic Acid, *Macadamia Nut Oil*. *Energy & Fuels*, 2010. 24(3): p. 2098-2103.

117. Azad, A.K., et al., *Macadamia Biodiesel as a Sustainable and Alternative Transport Fuel in Australia*. *Energy Procedia*, 2017. 110: p. 543-548.

118. Kivevele, T.T. and M.M. Mbarawa, Experimental Investigations of Oxidation Stability of Biodiesel Produced from Manketti Seeds Oil (*Schinziophyton rautanenii*). *Energy & Fuels*, 2011. 25(5): p. 2341-2346.

119. Kivevele, T.T. and Z. Huan, An Analysis of Fuel Properties of Fatty Acid Methyl Ester from Manketti Seeds Oil. *International Journal of Green Energy*, 2014. 12(4): p. 291-296.

120. Rutto, H.L. and C.C. Enweremadu, Optimization of Production Variables of Biodiesel from Manketti Using Response Surface Methodology. *International Journal of Green Energy*, 2011. 8(7): p. 768-779.

121. Khiari, K., et al., Experimental investigation of pistacia lentiscus biodiesel as a fuel for direct injection diesel engine. *Energy Conversion and Management*, 2016. 108: p. 392-399.

122. Samani, B.H., et al., Ultrasonic-assisted production of biodiesel from *Pistacia atlantica* Desf. oil. *Fuel*, 2016. 168: p. 22-26.

123. Ashok, B., et al., Lemon peel oil – A novel renewable alternative energy source for diesel engine. *Energy Conversion and Management*, 2017. 139: p. 110-121.

124. Kumar, P. and N. Kumar, Comparative study of biodiesel from *Jatropha* and orange peel oils as pilot fuels in a dual-fuel engine. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, 2016. 38(23): p. 3491-3496.

125. Tüccar, G., et al., Diesel engine emissions and performance from blends of citrus *sinensis* biodiesel and diesel fuel. *Fuel*, 2014. 132: p. 7-11.

126. Agarry, S., et al., Alkali-catalysed production of biodiesel fuel from Nigerian Citrus seeds oil. *International Journal of Engineering Science and Technology*, 2013. 5(9): p. 1682.

127. Deep, A., et al., Potential Utilization of the Blend of Orange Peel Oil Methyl Ester and Isopropyl Alcohol in CI Engine, in *SAE Technical Paper Series*. 2014.

128. Rashid, U., et al., *Momordica Charantia* Seed Oil Methyl Esters: A Kinetic Study And Fuel Properties. *International*

Journal of Green Energy, 2014. 11(7): p. 727-740.

129. Giwa, S., L.C. Abdullah, and N.M. Adam, Investigating “Egusi” (*Citrullus Colocynthis* L.) Seed Oil as Potential Biodiesel Feedstock. *Energies*, 2010. 3(4): p. 607-618.

130. Schinas, P., et al., Pumpkin (*Cucurbita pepo* L.) seed oil as an alternative feedstock for the production of biodiesel in Greece. *Biomass and Bioenergy*, 2009. 33(1): p. 44-49.

131. Bello, E., S. Anjorin, and M. Agge, Production of biodiesel from fluted pumpkin (*Telfairia occidentalis* Hook F.) seeds oil. *International Journal of Mechanical Engineering*, 2011. 1(2).

132. Sokoto, M., et al., Influence of fatty acid methyl esters on fuel properties of biodiesel produced from the seeds oil of *Curcubita pepo*. *Nigerian Journal of Basic and Applied Sciences*, 2011. 19(1).

133. Ogunwale, O.A., Production of biodiesel from watermelon (*Citrullus lanatus*) seed oil. *Leonardo J. Sci*, 2015. 27: p. 63-74.

134. Asokan, M.A., et al., Performance, combustion and emission characteristics of diesel engine fuelled with papaya and watermelon seed oil bio-diesel/diesel blends. *Energy*, 2018. 145: p. 238-245.

135. Górnaś, P. and M. Rudzińska, Seeds recovered from industry by-products of nine fruit species with a high potential utility as a source of unconventional oil for biodiesel and cosmetic and pharmaceutical sectors. *Industrial Crops and Products*, 2016. 83: p. 329-338.

136. Panneerselvam, N., et al., Performance, emissions and combustion characteristics of CI engine fuel with watermelon (*Citrullus vulgaris*) methyl esters. *International Journal of Ambient Energy*, 2015. 38(3): p. 308-313.

137. Usta, N., et al., Properties and quality verification of biodiesel produced from tobacco seed oil. *Energy Conversion and Management*, 2011. 52(5): p. 2031-2039.

138. Pantoja, S.S., et al., Oxidative stability of biodiesels produced from vegetable oils having different degrees of unsaturation. *Energy Conversion and Management*, 2013. 74: p. 293-298.

139. Yunus Khan, T.M., et al., *Ceiba pentandra*, *Nigella sativa* and their blend as prospective feedstocks for biodiesel. *Industrial Crops and Products*, 2015. 65: p. 367-373.

140. Knothe, G., Fuel properties of methyl esters of borage and black currant oils containing methyl γ -linolenate. *European Journal of Lipid Science and Technology*, 2013. 115(8): p. 901-908.

141. Hoseini, S.S., et al., Characterization of biodiesel production (ultrasonic-assisted) from evening-primroses (*Oenothera lamarckiana*) as novel feedstock and its effect on CI engine parameters. *Renewable Energy*, 2019. 130: p. 50-60.

142. Demirbas, A., Production of biodiesel fuels from linseed oil using methanol and ethanol in non-catalytic SCF conditions. *Biomass and Bioenergy*, 2009. 33(1): p. 113-118.

143. Gumus, M. and S. Kasifoglu, Performance and emission evaluation of a compression ignition engine using a biodiesel (apricot seed kernel oil methyl ester) and its blends with diesel fuel. *Biomass and Bioenergy*, 2010. 34(1): p. 134-139.

144. Hotti, S.R. and O.D. Hebbal, Biodiesel Production Process Optimization from Sugar Apple Seed Oil (*Annona squamosa*) and Its Characterization. *Journal of Renewable Energy*, 2015. 2015: p. 1-6.

145. Sriram, V., J. Jeevahan, and A. Poovannan, Comparative study of performance of the DI diesel engine using custard apple biodiesel. *International Journal of Ambient Energy*, 2017. 40(1): p. 54-56.

146. Rachimoallah, H., et al., Production of biodiesel through transesterification of avocado (*Persea gratissima*) seed oil using base catalyst. *Jurnal Teknik Mesin*, 2010. 11(2): p. 85-90.

147. Nogueira Jr, C.A., et al., Densities and viscosities of binary mixtures of babassu biodiesel+ cotton seed or soybean biodiesel at different temperatures. *Journal of Chemical & Engineering Data*, 2010. 55(11): p. 5305-5310.
148. Cesur, C., et al., Cocklebur (*Xanthium strumarium* L.) seed oil and its properties as an alternative biodiesel source. *Turkish Journal of Agriculture and Forestry*, 2018. 42: p. 29-37.
149. Chapagain, B.P., Y. Yehoshua, and Z. Wiesman, Desert date (*Balanites aegyptiaca*) as an arid lands sustainable bioresource for biodiesel. *Bioresour Technol*, 2009. 100(3): p. 1221-6.
150. Bello, E., et al., Characterization and evaluation of African bush mango Nut (Dika nut)(*Irvingia gabonensis*) oil biodiesel as alternative fuel for diesel engines. *Journal of Petroleum Technology and Alternative Fuels*, 2011. 2(9): p. 176-180.
151. Nwufu, O.C., Effect of temperature on the biodiesel yield from Nigerian physic nut, castor bean, dika nut and sandbax seed oils. *International Journal of Ambient Energy*, 2013. 37(1): p. 16-19.
152. Patil, P.D., V.G. Gude, and S. Deng, Biodiesel production from *Jatropha curcas*, waste cooking, and *Camelina sativa* oils. *Industrial & Engineering Chemistry Research*, 2009. 48(24): p. 10850-10856.
153. Li, S.Y., et al., The feasibility of converting *Cannabis sativa* L. oil into biodiesel. *Bioresour Technol*, 2010. 101(21): p. 8457-60.
154. Ahmad, M., et al., Physicochemical Analysis of Hemp Oil Biodiesel: A Promising Non Edible New Source for Bioenergy. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, 2011. 33(14): p. 1365-1374.
155. Ragit, S.S., et al., Brown hemp methyl ester: Transesterification process and evaluation of fuel properties. *Biomass and Bioenergy*, 2012. 41: p. 14-20.
156. Yang, R., et al., One-pot process combining transesterification and selective hydrogenation for biodiesel production from starting material of high degree of unsaturation. *Bioresour Technol*, 2010. 101(15): p. 5903-9.
157. Stamenković, O.S., et al., Optimization of KOH-catalyzed methanolysis of hempseed oil. *Energy Conversion and Management*, 2015. 103: p. 235-243.
158. Silitonga, A.S., et al., Experimental study on performance and exhaust emissions of a diesel engine fuelled with *Ceiba pentandra* biodiesel blends. *Energy Conversion and Management*, 2013. 76: p. 828-836.
159. Vallinayagam, R., et al., Pine oil–biodiesel blends: A double biofuel strategy to completely eliminate the use of diesel in a diesel engine. *Applied Energy*, 2014. 130: p. 466-473.
160. Knothe, G., L.F. Razon, and F.T. Bacani, Kenaf oil methyl esters. *Industrial Crops and Products*, 2013. 49: p. 568-572.
161. Modiba, E., P. Osifo, and H. Rutto, The use of impregnated perlite as a heterogeneous catalyst for biodiesel production from marula oil. *Chemical Papers*, 2014. 68(10).
162. Moser, B.R., G. Knothe, and S.C. Cermak, Biodiesel from meadowfoam (*Limnanthes alba* L.) seed oil: oxidative stability and unusual fatty acid composition. *Energy & Environmental Science*, 2010. 3(3): p. 318-327.
163. Yerranguntla, R., et al., Production of Biodiesel from *Guizotia abyssinica* seed oil using crystalline Manganese carbonate ($MnCO_3$) a Green catalyst. *Catalysis for Sustainable Energy*, 2012. 1.
164. Sarin, R., M. Sharma, and A.A. Khan, Studies on *Guizotia abyssinica* L. oil: biodiesel synthesis and process optimization. *Bioresour Technol*, 2009. 100(18): p. 4187-92.
165. Anwar, M., M.G. Rasul, and N. Ashwath, Production optimization and quality assessment of papaya (*Carica papaya*) biodiesel with response surface methodology. *Energy Conversion and Management*, 2018. 156: p. 103-112.

166. Wong, C. and R. Othman, Biodiesel production by enzymatic transesterification of papaya seed oil and rambutan seed oil. *Int. J. Eng. Technol*, 2014. 6: p. 2773-2777.
167. De Melo, M.L.S., et al., Use of thermal analysis techniques for evaluation of the stability and chemical properties of papaya biodiesel (*Carica Papaya L.*) at low temperatures. *Journal of Thermal Analysis and Calorimetry*, 2011. 106(3): p. 831-836.
168. Agunbiade, F.O. and T.A. Adewole, Methanolysis of *Carica papaya* Seed Oil for Production of Biodiesel. *Journal of Fuels*, 2014. 2014: p. 1-6.
169. Silva, T.A., et al., Methyl and ethyl biodiesels from pequi oil (*Caryocar brasiliense Camb.*): Production and thermogravimetric studies. *Fuel*, 2014. 136: p. 10-18.
170. Borges, K.A., et al., Production of methyl and ethyl biodiesel fuel from pequi oil (*Caryocar brasiliensis Camb.*). *Chemistry and Technology of Fuels and Oils*, 2012. 48(2): p. 83-89.
171. Tüccar, G. and E. Uludamar, Emission and engine performance analysis of a diesel engine using hydrogen enriched pomegranate seed oil biodiesel. *International Journal of Hydrogen Energy*, 2018. 43(38): p. 18014-18019.
172. Lima, R.P., et al., Murumuru (*Astrocaryum murumuru Mart.*) butter and oils of buriti (*Mauritia flexuosa Mart.*) and pracaxi (*Pentaclethra macroloba (Willd.) Kuntze*) can be used for biodiesel production: Physico-chemical properties and thermal and kinetic studies. *Industrial crops and products*, 2017. 97: p. 536-544.
173. Rashid, U., et al., Optimization of alkaline transesterification of rice bran oil for biodiesel production using response surface methodology. *Journal of Chemical Technology & Biotechnology*, 2009. 84(9): p. 1364-1370.
174. Ahmad, M., et al., Physicochemical Characterization of Eco-friendly Rice Bran Oil Biodiesel. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, 2011. 33(14): p. 1386-1397.
175. El Boulifi, N., et al., Optimization and oxidative stability of biodiesel production from rice bran oil. *Renewable Energy*, 2013. 53: p. 141-147.
176. Sinha, S., A.K. Agarwal, and S. Garg, Biodiesel development from rice bran oil: Transesterification process optimization and fuel characterization. *Energy Conversion and Management*, 2008. 49(5): p. 1248-1257.
177. Zuleta, E.C., L.A. Rios, and P.N. Benjumea, Oxidative stability and cold flow behavior of palm, sachal-inchi, jatropha and castor oil biodiesel blends. *Fuel Processing Technology*, 2012. 102: p. 96-101.
178. Pochareddy, Y.K., et al., Performance and emission characteristics of a stationary direct injection compression ignition engine fuelled with diethyl ether–sapote seed oil methyl ester–diesel blends. *Biofuels*, 2016. 8(2): p. 297-305.
179. Kumar, A.R., G.J. Raju, and K.H. Reddy, Biodiesel production by alkaline transesterification of Mamey Sapote oil. *International Journal for Research in Applied Science and Engineering Technology*, 2016. 4(2): p. 391-396.
180. Enweremadu, C., H. Rutto, and J. Oladeji, Investigation of the relationship between some basic flow properties of shea butter biodiesel and their blends with diesel fuel. *International Journal of Physical Sciences*, 2011. 6(4): p. 758-767.
181. Ajala, E.O., et al., Optimization of a two stage process for biodiesel production from shea butter using response surface methodology. *Egyptian Journal of Petroleum*, 2017. 26(4): p. 943-955.
182. Bello, E.I. and A. Mamman, Shea Butter (*Vitellaria paradoxa*) Biodiesel. *Journal: JOURNAL OF ADVANCES IN BIOTECHNOLOGY*, 2015. 4(3).
183. Chakrabarti, M.H. and R. Ahmad, Investigating possibility of using least desirable edible oil of *Eruca sativa L.*, in biodiesel production. *Pakistan Journal of Botany*, 2009. 41(1): p. 481-7.

184. Mumtaz, M.W., et al., Biodiesel production using *Eruca sativa* oil: optimization and characterization. *Pakistan journal of botany*, 2012. 44(3): p. 1111-1120.
185. Serin, H. and Ş. Yıldızhan, Hydrogen addition to tea seed oil biodiesel: Performance and emission characteristics. *International Journal of Hydrogen Energy*, 2018. 43(38): p. 18020-18027.
186. Takase, M., et al., *Silybum marianum* oil as a new potential non-edible feedstock for biodiesel: A comparison of its production using conventional and ultrasonic assisted method. *Fuel Processing Technology*, 2014. 123: p. 19-26.
187. Fadhil, A.B., A.M. Aziz, and M.H. Al-Tamer, Biodiesel production from *Silybum marianum* L. seed oil with high FFA content using sulfonated carbon catalyst for esterification and base catalyst for transesterification. *Energy Conversion and Management*, 2016. 108: p. 255-265.
188. Ahmad, M., et al., The Optimization of Biodiesel Production from a Novel Source of Wild Non-Edible Oil Yielding Plant *Silybum Marianum*. *International Journal of Green Energy*, 2013. 11(6): p. 589-594.
189. Usta, N., Use of tobacco seed oil methyl ester in a turbocharged indirect injection diesel engine. *Biomass and Bioenergy*, 2005. 28(1): p. 77-86.
190. Rao, B.S., et al., Studies on exhaust emissions and combustion characteristics of tobacco seed oil in crude form and biodiesel from a high grade low heat rejection diesel engine. *International Journal of Industrial Engineering and Technology*, 2013. 3(1): p. 27-36.
191. Veljkovic, V., et al., Biodiesel production from tobacco (*Nicotiana tabacum* L.) seed oil with a high content of free fatty acids. *Fuel*, 2006. 85(17-18): p. 2671-2675.
192. Sivasubramanian, H., et al., Investigation of biodiesel obtained from tomato seed as a potential fuel alternative in a CI engine. *Biofuels*, 2017: p. 1-9.
193. Dias, J.M., et al., Biodiesel production from raw castor oil. *Energy*, 2013. 53: p. 58-66.
194. Barminas, J., et al., A preliminary investigation into the biofuel characteristics of tigernut (*Cyperus esculentus*) oil. *Bioresource technology*, 2001. 79(1): p. 87-89.
195. Kaur, A., M. Roy, and K. Kundu, Transesterification process optimization for tung oil methyl ester (*Aleurites fordii*) and characterization of fuel as a substitute for diesel. *IJCS*, 2017. 5(6): p. 632-638.
196. Chen, Y.H., et al., Biodiesel production from tung (*Vernicia montana*) oil and its blending properties in different fatty acid compositions. *Bioresour Technol*, 2010. 101(24): p. 9521-6.
197. Sahoo, P.K. and L.M. Das, Process optimization for biodiesel production from *Jatropha*, *Karanja* and *Polanga* oils. *Fuel*, 2009. 88(9): p. 1588-1594.
198. Balan, K.N., et al., Investigation on emission characteristics of alcohol biodiesel blended diesel engine. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, 2018. 41(15): p. 1879-1889.
199. Diana da Silva Araújo, F., et al., Study of degumming process and evaluation of oxidative stability of methyl and ethyl biodiesel of *Jatropha curcas* L. oil from three different Brazilian states. *Renewable Energy*, 2014. 71: p. 495-501.
200. Mazumdar, P., et al., Physico-chemical characteristics of *Jatropha curcas* L. of North East India for exploration of biodiesel. *Biomass and Bioenergy*, 2012. 46: p. 546-554.
201. Shah, M., et al., Catalytic conversion of jojoba oil into biodiesel by organotin catalysts, spectroscopic and chromatographic characterization. *Fuel*, 2014. 118: p. 392-397.
202. Al-Hamamre, Z. and A. Al-Salaymeh, Physical properties of (jojoba oil+biodiesel), (jojoba oil+diesel) and (biodiesel+diesel) blends. *Fuel*, 2014. 123: p. 175-188.

203. Aslam, M., P. Saxena, and A.K. Sarma, Green Technology for Biodiesel Production From *Mesua Ferrea* L. Seed Oil. *Energy and Environment Research*, 2014. 4(2).
204. De, B. and D. Bhattacharyya, Biodiesel from minor vegetable oils like karanja oil and nahor oil. *Lipid/Fett*, 1999. 101(10): p. 404-406.
205. Devan, P.K. and N.V. Mahalakshmi, Utilization of unattended methyl ester of paradise oil as fuel in diesel engine. *Fuel*, 2009. 88(10): p. 1828-1833.
206. Mishra, S., et al., Production of bio-diesel (Methyl Ester) from *Simarouba glauca* oil. *Research Journal of Chemical Sciences* ISSN, 2012. 2231: p. 606X.
207. Mishra, S.R., et al., Optimisation of base-catalysed transesterification of *Simarouba glauca* oil for biodiesel production. *International Journal of Sustainable Energy*, 2013. 33(6): p. 1033-1040.
208. Baiju, B., M.K. Naik, and L.M. Das, A comparative evaluation of compression ignition engine characteristics using methyl and ethyl esters of Karanja oil. *Renewable Energy*, 2009. 34(6): p. 1616-1621.
209. Wang, R., et al., Production and selected fuel properties of biodiesel from promising non-edible oils: *Euphorbia lathyris* L., *Sapium sebiferum* L. and *Jatropha curcas* L. *Bioresour Technol*, 2011. 102(2): p. 1194-9.
210. Li, Q. and Y. Yan, Production of biodiesel catalyzed by immobilized *Pseudomonas cepacia* lipase from *Sapium sebiferum* oil in micro-aqueous phase. *Applied Energy*, 2010. 87(10): p. 3148-3154.
211. Liu, Y., H.-l. Xin, and Y.-j. Yan, Physicochemical properties of *Stillingia* oil: Feasibility for biodiesel production by enzyme transesterification. *Industrial Crops and Products*, 2009. 30(3): p. 431-436.
212. Encinar, J.M., et al., Preparation and properties of biodiesel from *Cynara Cardunculus* L. oil. *Industrial & Engineering Chemistry Research*, 1999. 38(8): p. 2927-2931.
213. Fernández, J., M.D. Curt, and P.L. Aguado, Industrial applications of *Cynara cardunculus* L. for energy and other uses. *Industrial Crops and Products*, 2006. 24(3): p. 222-229.
214. Pereira Lima, R., et al., Murumuru (*Astrocaryum murumuru* Mart.) butter and oils of buriti (*Mauritia flexuosa* Mart.) and pracaxi (*Pentaclethra macroloba* (Willd.) Kuntze) can be used for biodiesel production: Physico-chemical properties and thermal and kinetic studies. *Industrial Crops and Products*, 2017. 97: p. 536-544.
215. Deshmukh, S.J. and L.B. Bhuyar, Transesterified Hingan (*Balanites*) oil as a fuel for compression ignition engines. *Biomass and Bioenergy*, 2009. 33(1): p. 108-112.
216. Naik, N.S. and B. Balakrishna, Experimental evaluation of a diesel engine fueled with *Balanites aegyptiaca* (L.) Del biodiesel blends. *Biofuels*, 2016. 7(6): p. 603-609.
217. Hasni, K., et al., Optimization of biodiesel production from *Brucea javanica* seeds oil as novel non-edible feedstock using response surface methodology. *Energy Conversion and Management*, 2017. 149: p. 392-400.
218. Pantoja, S.S., et al., High-Quality Biodiesel Production from Buriti (*Mauritia flexuosa*) Oil Soapstock. *Molecules*, 2018. 24(1).
219. Sulisty, H., et al., Biodiesel production from high iodine number candlenut oil. *World Acad. Sci. Eng. Technol*, 2008. 48: p. 485-488.
220. Imdadul, H.K., et al., Experimental assessment of non-edible candlenut biodiesel and its blend characteristics as diesel engine fuel. *Environ Sci Pollut Res Int*, 2017. 24(3): p. 2350-2363.
221. Math, M.C. and H.L. Hegde, Experimental studies on performance and combustion characteristics of a compression ignition engine fuelled with *Hydnocarpus wightiana* methyl ester and its blends with conventional diesel and domestic kerosene. *Biofuels*, 2017. 9(6): p. 677-684.

222. Knothe, G., S.C. Cermak, and R.L. Evangelista, Cuphea oil as source of biodiesel with improved fuel properties caused by high content of methyl decanoate. *Energy & fuels*, 2009. 23(3): p. 1743-1747.
223. Dodos, G.S., et al., Renewable fuels and lubricants from *Lunaria annua* L. *Industrial Crops and Products*, 2015. 75: p. 43-50.
224. Ogunsuyi, H., Acid and base catalysed transesterification of mango (*mangifera indica*) seed oil to biodiesel. *IOSR Journal of Applied Chemistry*, 2012. 2(2): p. 18-22.
225. Velmurugan, K. and A.P. Sathiyagnanam, Impact of antioxidants on NO_x emissions from a mango seed biodiesel powered DI diesel engine. *Alexandria Engineering Journal*, 2016. 55(1): p. 715-722.
226. Pillay, A.E., et al., A comparison of trace metal profiles of neem biodiesel and commercial biofuels using high performance ICP-MS. *Fuel*, 2012. 97: p. 385-389.
227. Anya, U.A., N.N. Chioma, and O. Obinna, Optimized reduction of free fatty acid content on neem seed oil, for biodiesel production. *Journal of Basic and Applied Chemistry*, 2012. 2(4): p. 21-28.
228. Aransiola, E.F., Production of biodiesel from crude neem oil feedstock and its emissions from internal combustion engines. *African Journal of Biotechnology*, 2012. 11(22).
229. Muthu, H., et al., Synthesis of biodiesel from neem oil using sulfated zirconia via transesterification. *Brazilian Journal of Chemical Engineering*, 2010. 27(4): p. 601-608.
230. Bello, E.I., F. Otu, and S. Rao, Physicochemical Properties of Rubber (*Hevea brasiliensis*) Seed Oil, Its Biodiesel and Blends with Diesel. *British Journal of Applied Science & Technology*, 2015. 6(3): p. 261-275.
231. Demirbas, A., Production of Biodiesel from Tall Oil. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, 2008. 30(20): p. 1896-1902.
232. Altiparmak, D., et al., Alternative fuel properties of tall oil fatty acid methyl ester-diesel fuel blends. *Bioresour Technol*, 2007. 98(2): p. 241-6.
233. Jahirul, M.I., et al., Physio-chemical assessment of beauty leaf (*Calophyllum inophyllum*) as second-generation biodiesel feedstock. *Energy Reports*, 2015. 1: p. 204-215.
234. Peer, M.S., et al., Experimental evaluation on oxidation stability of biodiesel/diesel blends with alcohol addition by rancimat instrument and FTIR spectroscopy. *Journal of Mechanical Science and Technology*, 2017. 31(1): p. 455-463.
235. Mohanraj, T. and K.M. Mohan Kumar, Operating Characteristics of a Variable Compression Ratio Engine Using Esterified Tamanu Oil. *International Journal of Green Energy*, 2013. 10(3): p. 285-301.
236. Lima, J.R.d.O., et al. Indian-Nut (*Aleurites Moluccana*) and Tucum (*Astrocaryum Vulgare*); Non Agricultural Sources for Biodiesel Production Using Ethanol: Composition; Characterization and Optimization of the Reactional Production Conditions. in *World Renewable Energy Congress-Sweden*; 8-13 May; 2011; Linköping; Sweden. 2011. Linköping University Electronic Press.
237. Lima, J.R.d.O., et al., Biodiesel of tucum oil, synthesized by methanolic and ethanolic routes. *Fuel*, 2008. 87(8-9): p. 1718-1723.
238. Pelegrini, B.L., et al., Thermal and rheological properties of soapberry *Sapindus saponaria* L. (*Sapindaceae*) oil biodiesel and its blends with petrodiesel. *Fuel*, 2017. 199: p. 627-640.
239. Chakraborty, M. and D.C. Baruah, Production and characterization of biodiesel obtained from *Sapindus mukorossi* kernel oil. *Energy*, 2013. 60: p. 159-167.
240. Deka, D.C. and S. Basumatary, High quality biodiesel from yellow oleander (*Thevetia peruviana*) seed oil. *Biomass and Bioenergy*, 2011. 35(5): p. 1797-1803.

241. Yadav, A.K., et al., Biodiesel production from *Nerium oleander* (*Thevetia peruviana*) oil through conventional and ultrasonic irradiation methods. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, 2016. 38(23): p. 3447-3452.
242. Ang, G.T., et al., Optimization and kinetic studies of sea mango (*Cerbera odollam*) oil for biodiesel production via supercritical reaction. *Energy Conversion and Management*, 2015. 99: p. 242-251.
243. Nayak, S.K. and P.C. Mishra, Application of Nagchampa biodiesel and rice husk gas as fuel. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, 2016. 38(14): p. 2024-2030.
244. Gole, V.L. and P.R. Gogate, Intensification of Synthesis of Biodiesel from Nonedible Oils Using Sonochemical Reactors. *Industrial & Engineering Chemistry Research*, 2012. 51(37): p. 11866-11874.
245. Kivevele, T.T. and M.M. Mbarawa, Comprehensive Analysis of Fuel Properties of Biodiesel from *Croton megalocarpus* Oil. *Energy & Fuels*, 2010. 24(11): p. 6151-6155.
246. Haik, Y., M.Y.E. Selim, and T. Abdulrehman, Combustion of algae oil methyl ester in an indirect injection diesel engine. *Energy*, 2011. 36(3): p. 1827-1835.
247. Xiao, Y., et al., Industrial Fermentation of *Auxenochlorella protothecoides* for Production of Biodiesel and Its Application in Vehicle Diesel Engines. *Front Bioeng Biotechnol*, 2015. 3: p. 164.
248. Chen, Y.-H., et al., Fuel properties of microalgae (*Chlorella protothecoides*) oil biodiesel and its blends with petroleum diesel. *Fuel*, 2012. 94: p. 270-273.
249. Satputaley, S.S., D.B. Zodpe, and N.V. Deshpande, Performance, combustion and emission study on CI engine using microalgae oil and microalgae oil methyl esters. *Journal of the Energy Institute*, 2017. 90(4): p. 513-521.
250. Xu, H., X. Miao, and Q. Wu, High quality biodiesel production from a microalga *Chlorella protothecoides* by heterotrophic growth in fermenters. *J Biotechnol*, 2006. 126(4): p. 499-507.
251. Singh, D., et al., Transient performance and emission characteristics of a heavy-duty diesel engine fuelled with microalga *Chlorella variabilis* and *Jatropha curcas* biodiesels. *Energy Conversion and Management*, 2015. 106: p. 892-900.
252. Kalhor, A.X., et al., Biodiesel production in crude oil contaminated environment using *Chlorella vulgaris*. *Bioresource technology*, 2016. 222: p. 190-194.
253. Mathimani, T., L. Uma, and D. Prabakaran, Homogeneous acid catalysed transesterification of marine microalga *Chlorella* sp. BDUG 91771 lipid – An efficient biodiesel yield and its characterization. *Renewable Energy*, 2015. 81: p. 523-533.
254. Arunachalam Sivagurulingam, A.P., et al., Optimization and kinetic studies on biodiesel production from microalgae (*Euglena sanguinea*) using calcium methoxide as catalyst. *Energy Sources, Part A: Recovery, Utilization, and Environmental Effects*, 2018. 41(12): p. 1497-1507.
255. Kings, A.J., et al., Cultivation, extraction and optimization of biodiesel production from potential microalgae *Euglena sanguinea* using eco-friendly natural catalyst. *Energy Conversion and Management*, 2017. 141: p. 224-235.
256. Petersen, J., et al., Combustion Characterization and Ignition Delay Modeling of Low- and High-Cetane Alternative Diesel Fuels in a Marine Diesel Engine. *Energy & Fuels*, 2014. 28(8): p. 5463-5471.
257. Khan, A.M. and N. Fatima, Biodiesel synthesis via metal oxides and metal chlorides catalysis from marine alga *Melanothamnus afaqusainii*. *Chinese Journal of Chemical Engineering*, 2016. 24(3): p. 388-393.
258. Nautiyal, P., K.A. Subramanian, and M.G. Dastidar, Production and characterization of biodiesel from algae. *Fuel Processing Technology*, 2014. 120: p. 79-88.

259. Hong, D.D., et al., Biodiesel production from Vietnam heterotrophic marine microalga *Schizochytrium mangrovei* PQ6. *J Biosci Bioeng*, 2013. 116(2): p. 180-5.
260. Mostafa, S.S.M. and N.S. El-Gendy, Evaluation of fuel properties for microalgae *Spirulina platensis* bio-diesel and its blends with Egyptian petro-diesel. *Arabian Journal of Chemistry*, 2017. 10: p. S2040-S2050.
261. Barrios, C.C., et al., Effects of animal fat based biodiesel on a TDI diesel engine performance, combustion characteristics and particle number and size distribution emissions. *Fuel*, 2014. 117: p. 618-623.
262. Armas, O., A. Gómez, and Á. Ramos, Comparative study of pollutant emissions from engine starting with animal fat biodiesel and GTL fuels. *Fuel*, 2013. 113: p. 560-570.
263. Ballesteros, R., E. Monedero, and J. Guillén-Flores, Determination of aldehydes and ketones with high atmospheric reactivity on diesel exhaust using a biofuel from animal fats. *Atmospheric Environment*, 2011. 45(16): p. 2690-2698.
264. Awad, S., K. Loubar, and M. Tazerout, Experimental investigation on the combustion, performance and pollutant emissions of biodiesel from animal fat residues on a direct injection diesel engine. *Energy*, 2014. 69: p. 826-836.
265. Öner, C. and Ş. Altun, Biodiesel production from inedible animal tallow and an experimental investigation of its use as alternative fuel in a direct injection diesel engine. *Applied Energy*, 2009. 86(10): p. 2114-2120.
266. Teixeira, L.S.G., et al., Comparison between conventional and ultrasonic preparation of beef tallow biodiesel. *Fuel Processing Technology*, 2009. 90(9): p. 1164-1166.
267. Sander, A., et al., The influence of animal fat type and purification conditions on biodiesel quality. *Renewable Energy*, 2018. 118: p. 752-760.
268. Araújo, B.Q., et al., Synthesis and Characterization of Beef Tallow Biodiesel. *Energy & Fuels*, 2010. 24(8): p. 4476-4480.
269. Sbihi, H.M., et al., Production and characterization of biodiesel from *Camelus dromedarius* (Hachi) fat. *Energy Conversion and Management*, 2014. 78: p. 50-57.
270. Bhatti, H., et al., Biodiesel production from waste tallow. *Fuel*, 2008. 87(13-14): p. 2961-2966.
271. de Guzman, R., et al., Synergistic Effects of Antioxidants on the Oxidative Stability of Soybean Oil- and Poultry Fat-Based Biodiesel. *Journal of the American Oil Chemists' Society*, 2009. 86(5): p. 459-467.
272. Alptekin, E., et al., Using waste animal fat based biodiesels–bioethanol–diesel fuel blends in a DI diesel engine. *Fuel*, 2015. 157: p. 245-254.
273. Godiganur, S., C. Suryanarayana Murthy, and R.P. Reddy, Performance and emission characteristics of a Kirloskar HA394 diesel engine operated on fish oil methyl esters. *Renewable Energy*, 2010. 35(2): p. 355-359.
274. Gnanasekaran, S., N. Saravanan, and M. Ilangkumaran, Influence of injection timing on performance, emission and combustion characteristics of a DI diesel engine running on fish oil biodiesel. *Energy*, 2016. 116: p. 1218-1229.
275. Swaminathan, C. and J. Sarangan, Performance and exhaust emission characteristics of a CI engine fueled with biodiesel (fish oil) with DEE as additive. *Biomass and Bioenergy*, 2012. 39: p. 168-174.
276. Behçet, R., Performance and emission study of waste anchovy fish biodiesel in a diesel engine. *Fuel Processing Technology*, 2011. 92(6): p. 1187-1194.
277. Dias, J.M., M.C. Alvim-Ferraz, and M.F. Almeida, Production of biodiesel from acid waste lard. *Bioresour Technol*, 2009. 100(24): p. 6355-61.
278. Ramalho, E.F.S.M., et al., Low temperature behavior of poultry fat biodiesel:diesel blends. *Fuel*, 2012. 93: p. 601-605.

279. Tang, H., S.O. Salley, and K.S. Ng, Fuel properties and precipitate formation at low temperature in soy-, cottonseed-, and poultry fat-based biodiesel blends. *Fuel*, 2008. 87(13-14): p. 3006-3017.
280. Can, Ö., Combustion characteristics, performance and exhaust emissions of a diesel engine fueled with a waste cooking oil biodiesel mixture. *Energy Conversion and Management*, 2014. 87: p. 676-686.
281. Cai, Z.-Z., et al., A two-step biodiesel production process from waste cooking oil via recycling crude glycerol esterification catalyzed by alkali catalyst. *Fuel Processing Technology*, 2015. 137: p. 186-193.
282. Lin, Y.-C., K.-H. Hsu, and C.-B. Chen, Experimental investigation of the performance and emissions of a heavy-duty diesel engine fueled with waste cooking oil biodiesel/ultra-low sulfur diesel blends. *Energy*, 2011. 36(1): p. 241-248.
283. Maddikeri, G.L., P.R. Gogate, and A.B. Pandit, Intensified synthesis of biodiesel using hydrodynamic cavitation reactors based on the transesterification of waste cooking oil. *Fuel*, 2014. 137: p. 285-292.
284. Muralidharan, K. and D. Vasudevan, Performance, emission and combustion characteristics of a variable compression ratio engine using methyl esters of waste cooking oil and diesel blends. *Applied Energy*, 2011. 88(11): p. 3959-3968.
285. Lin, Y.-S. and H.-P. Lin, Study on the spray characteristics of methyl esters from waste cooking oil at elevated temperature. *Renewable Energy*, 2010. 35(9): p. 1900-1907.
286. Sahar, et al., Biodiesel production from waste cooking oil: An efficient technique to convert waste into biodiesel. *Sustainable Cities and Society*, 2018. 41: p. 220-226.
287. Sanek, L., et al., Pilot-scale production of biodiesel from waste fats and oils using tetramethylammonium hydroxide. *Waste Manag*, 2016. 48: p. 630-637.
288. Atmanli, A., Comparative analyses of diesel-waste oil biodiesel and propanol, n-butanol or 1-pentanol blends in a diesel engine. *Fuel*, 2016. 176: p. 209-215.
289. Utlu, Z. and M.S. Koçak, The effect of biodiesel fuel obtained from waste frying oil on direct injection diesel engine performance and exhaust emissions. *Renewable Energy*, 2008. 33(8): p. 1936-1941.
290. Ozsezen, A.N. and M. Canakci, The emission analysis of an IDI diesel engine fueled with methyl ester of waste frying palm oil and its blends. *Biomass and Bioenergy*, 2010. 34(12): p. 1870-1878.
291. Buyukkaya, E., et al., Effects of trout-oil methyl ester on a diesel engine performance and emission characteristics. *Energy Conversion and Management*, 2013. 69: p. 41-48.
292. Yang, S., et al., Biodiesel production from swine manure via housefly larvae (*Musca domestica* L.). *Renewable Energy*, 2014. 66: p. 222-227.
293. Kalargaris, I., G. Tian, and S. Gu, Combustion, performance and emission analysis of a DI diesel engine using plastic pyrolysis oil. *Fuel Processing Technology*, 2017. 157: p. 108-115.
294. Hossain, A.K., et al., Experimental investigation of performance, emission and combustion characteristics of an indirect injection multi-cylinder CI engine fuelled by blends of de-inking sludge pyrolysis oil with biodiesel. *Fuel*, 2013. 105: p. 135-142.
295. Alagu, R.M. and E. Ganapathy Sundaram, Preparation and characterization of pyrolytic oil through pyrolysis of neem seed and study of performance, combustion and emission characteristics in CI engine. *Journal of the Energy Institute*, 2018. 91(1): p. 100-109.
296. Demirbas, A., Political, economic and environmental impacts of biofuels: A review. *Applied energy*, 2009. 86: p. S108-S117.